

Review

Gender Differences in Oxidative Stress in Relation to Cancer Susceptibility and Survival

Alessandro Allegra 1,* [,](https://orcid.org/0000-0001-6156-8239) Santino Caserta ¹ [,](https://orcid.org/0000-0002-0698-1781) Sara Genovese ² [,](https://orcid.org/0000-0002-2639-4680) Giovanni Pioggia 2,[†](https://orcid.org/0000-0002-8089-7449) and Sebastiano Gangemi 3,†

- ¹ Division of Hematology, Department of Human Pathology in Adulthood and Childhood 'Gaetano Barresi', University of Messina, 98125 Messina, Italy; 132588@polime.it
- 2 Institute for Biomedical Research and Innovation (IRIB), National Research Council of Italy (CNR), 98164 Messina, Italy; sara.genovese@cnr.it (S.G.); giovanni.pioggia@irib.cnr.it (G.P.)
- ³ Allergy and Clinical Immunology Unit, Department of Clinical and Experimental Medicine, University of Messina, 98100 Messina, Italy; gangemis@unime.it
- ***** Correspondence: aallegra@unime.it
- † These authors equally contributed to the work.

Abstract: Genetic, developmental, biochemical, and environmental variables interact intricately to produce sex differences. The significance of sex differences in cancer susceptibility is being clarified by numerous studies. Epidemiological research and cancer registries have revealed over the past few years that there are definite sex variations in cancer incidence, progression, and survival. However, oxidative stress and mitochondrial dysfunction also have a significant impact on the response to treatment of neoplastic diseases. Young women may be more protected from cancer than men because most of the proteins implicated in the regulation of redox state and mitochondrial function are under the control of sexual hormones. In this review, we describe how sexual hormones control the activity of antioxidant enzymes and mitochondria, as well as how they affect several neoplastic diseases. The molecular pathways that underlie the gender-related discrepancies in cancer that have been identified may be better understood, which may lead to more effective precision medicine and vital information on treatment options for both males and females with neoplastic illnesses.

Keywords: gender differences; cancer; oxidative stress; estrogens; testosterone; antioxidant; reactive oxygen species; sex hormones; mitochondria

1. Introduction

1.1. General Considerations on Gender Differences in Cancer Susceptibility

Epidemiological studies consistently demonstrate that there are gender variations in cancer incidence and mortality $[1,2]$ $[1,2]$. An analysis of the IARC's global cancer statistics highlighted that men were more likely than women to develop cancer in 32 out of 35 tumor sites. The authors concluded that the causes of the significant gender differences are unknown; in fact, in 13 of these sites, the discrepancies could not be accounted for by known risk variables [\[3\]](#page-17-2). Another study found that the incidence of cancer in nonreproductive organs is 1.8 times higher in men than in women [\[4\]](#page-17-3). Animal studies have revealed gender disparities in cancer incidences, even in rodents that were not exposed to any harmful substances [\[5\]](#page-17-4). One study on rats found 68 "male-specific" and 19 "female-specific" carcinogens that caused cancer, though the exact causes of this difference were not clear [\[6\]](#page-17-5).

A preliminary review of the literature on these gender-specific carcinogens revealed that oxidative stress might be a key mechanism, particularly for male-specific carcinogens. In fact, it has already been hypothesized that oxidative stress might affect a patient's susceptibility to developing cancer from chemical carcinogens, which is supported by the literature on gender disparities [\[7\]](#page-17-6).

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1.2. Oxidative Stress and Cancer For the production of th

The imbalance between the production of toxic reactive species (TRS) and antioxidant defense mechanisms is referred to as "oxidative stress". Reactive oxygen species (ROS) and reactive nitrogen species of TRS. Superoxide and reactive original reactive original reactive original reactive original reactive o reactive nitrogen species (RNS) are two categories of TRS. Superoxide anion, hydrogen peroxide, singlet oxygen, hydrogen superoxide, and reactive hydroxyl radical are the main components of ROS. The two RNS that are most known are nitric oxide and peroxynitrite [\[8\]](#page-17-7). Since ROS can damage DNA, proteins, and lipids within cells, oxidative stress is regarded as a serious condition. In cancer cells, TRS can come from a variety of sources. The chief contributors to the generation of ROS are thought to be an active metabolism and issues with the mitochondrial respiratory chain [\[9\]](#page-17-8). Additionally, activated macrophages infiltrate cancer tissue, increasing the inflammatory state and escalating the production of ROS and cytokines [\[10\]](#page-17-9). Moreover, the activation of oncogenes such as RAS2 or c-Myc is one example of a disorder in cellular signaling that is thought to be a significant generator of ROS [\[11](#page-17-10)[,12\]](#page-17-11). For cells, oxidative stress may be damaging, but intrinsic oxidative stress in cancer cells in malignant neoplasms may have dramatic effects, including cancer cell proliferation, the promotion of genetic instability, and changes in cellular sensitivity to anticancer agents, and the modulation of cellular redox parameters is a real possibility [\[13](#page-17-12)-18] (Figure [1\)](#page-1-0).

Figure 1. Cancer cells stimulate the activation of macrophages that infiltrate neoplastic tissue, with **Figure 1.** Cancer cells stimulate the activation of macrophages that infiltrate neoplastic tissue, with the consequent production of ROS, RNS, and cytokines production and, in turn, oxidative stress. the consequent production of ROS, RNS, and cytokines production and, in turn, oxidative stress. Oxidative stress, as in a cycle, promotes cancerogenesis. Oxidative stress, as in a cycle, promotes cancerogenesis.

1.3. Gender Differences, Oxidative Stress, and Cancer 1.3. Gender Differences, Oxidative Stress, and Cancer

In one review, the literature on twenty-six recognized human carcinogens (IARC In one review, the literature on twenty-six recognized human carcinogens (IARC group 1) was examined. The analysis was based on about $600,000$ abstracts and Tox21 screening assays and suggested a connection between testosterone, oxidative stress, and male-specific cancers. This would seem to indicate that the higher susceptibility to cancer seen in men may be due to a cellular response to oxidative stress that is only found in men [19]. 1) was examined. The analysis was based on about 600,000 abstracts and Tox21 screening
assays and suggested a connection between testosterone, oxidative stress, and male-specific
cancers. This would seem to indicate that t

as confirmed in many other species, including flies, mice, and rats [\[21](#page-17-16)[–23\]](#page-17-17). However, this hypothesis seems to depend on the cell type or tissue [\[22–](#page-17-18)[28\]](#page-17-19). Males are thought to experience oxidative stress more frequently than females [\[20\]](#page-17-15),

Other experimental data seem to confirm this hypothesis. NADPH oxidase activity and function appear to be lower in females, according to several studies [\[29\]](#page-18-0). Firstly, estrogen can directly cause reduced NADPH oxidase activity in females. Secondly, females have lower levels of p47, which is necessary for the assembly of the NADPH oxidase enzyme, as well as lower levels of superoxide production independently of estrogen. Females with lower levels of oxidative stress thus have lower levels of superoxide (Table [1\)](#page-2-0).

Table 1. Differences in oxidative stress in females.

In addition to gender differences in ROS generation, clinical and experimental studies have indicated that women have stronger antioxidant potential than men [\[30\]](#page-18-1). This may be because estrogen has antioxidant qualities, making women less vulnerable to oxidative stress [\[31\]](#page-18-2). Since postmenopausal women do not benefit from the anti-inflammatory and antioxidant protective properties of estrogen, they are more likely to experience increased oxidative stress [\[32\]](#page-18-3). Superoxide dismutase (SOD) transforms superoxide anions into hydrogen peroxide, and it seems that different tissues may react differently to this process.

Female rats apparently have higher heart SOD activity levels than male rats, and the levels of SOD activity in the brain and lungs seem to be higher in females [\[22\]](#page-17-18). Surprisingly, castration significantly reduced the levels of SOD activity in both male and female rats compared with the corresponding controls [\[22\]](#page-17-18), thus indicating that sex hormones may be related to SOD activity levels. SOD activity was reported to be higher in female erythrocytes than male erythrocytes in one human experiment [\[33\]](#page-18-4) and higher in female plasma than male plasma in another [\[33\]](#page-18-4).

Glutathione peroxidase (GPx) is another enzyme that detoxifies hydrogen peroxide into water and oxygen. Progesterone and testosterone and other sex hormones control how active the GPx enzyme is. They are tissue-dependent in terms of sex-related differences, while male mice seem to display higher GPx activity than females in the heart, and in females GPx, activity is primarily in the kidney and brain [\[34\]](#page-18-5). Previous research has also revealed higher GPx activity in the brain and liver of female rats [\[35\]](#page-18-6) and that the activity of GPx was more than twice as high in the hepatic mitochondria of female rats as compared with male rats of the same age [\[36\]](#page-18-7). In addition, female rats had more GPx activity in their livers than did male rats [\[37\]](#page-18-8), and in humans, teenage girls had higher GPx activity in their blood than did men [\[38\]](#page-18-9).

Since estrogen replacement therapy appears to increase erythrocyte GPx activity significantly in postmenopausal women and shows a positive correlation between GPx and serum estrogen levels in both premenopausal and postmenopausal women, it is likely that estrogen stimulates GPx expression. In fact, total hysterectomy in premenopausal females reduced the mRNA expression of SOD and GPx, which was then restored by estrogen replacement treatment, but had no effect on the expression of catalase [\[39\]](#page-18-10).

Hepatic mitochondrial GSH levels in female rats are higher than in male rats, which appears to lead to a lower glutathione (GSH) concentration.

However, after ovariectomization, the levels of mitochondrial glutathione in the rats dropped to levels comparable to those in males.

Adolescent girls likewise had a greater blood GSH/oxidized glutathione (GSSG) ratio than did men [\[38\]](#page-18-9), while men presented statistically significantly higher values of oxidized-reduced GSSG/GSH and GSSG in terms of concentration [\[40\]](#page-18-11). Additionally, premenopausal women that had had hysterectomies showed a decrease in GSH concentration

and an increase in GSSG and GSSG/GSH ratio after 30 days. Interestingly, estrogen replacement therapy brought glutathione levels back to what they were prior to hysterectomy, highlighting once again the critical role of estrogen in the glutathione cycle [\[39\]](#page-18-10).

Obesity is also a factor in the possible link between gender differences, oxidative stress, and the development of cancer. The burden of cancer attributable to obesity, expressed as population attributable fraction (PAF), is 11.9% in men and 13.1% in women for all obesityrelated malignancies worldwide—though this clearly varies from region to region [\[41\]](#page-18-12). The highest PAF is typically seen in cases of esophageal adenocarcinoma in men and endometrial cancer in women.

The overall strong relationship between obesity and gynecological cancer (endometrial, postmenopausal breast, and ovarian cancers) suggests that female sex hormones have a role in the etiology of cancer. The risks for various cancers, including colon, rectal, gallbladder, kidney, and pancreatic cancers, are associated differently by gender depending on BMI and other somatometric factors [\[42,](#page-18-13)[43\]](#page-18-14). This discovery emphasizes the harmful impact of visceral adiposity and insulin resistance in colon cancer, as well as the protective benefits of endogenous estrogenic effects against colon cancer in women [\[44–](#page-18-15)[46\]](#page-18-16). Men are more likely than women to develop visceral adiposity.

Given that it is a chronic inflammatory state [\[47\]](#page-18-17), obesity has been implicated in the initiation and progression of cancer [\[48\]](#page-18-18). This is due to the presence of numerous inflammatory components in the tumor microenvironment that support a malignant phenotype. Obese patients with metabolic abnormalities and adipose inflammation have a greater chance of developing cancer [\[49\]](#page-18-19).

Tumor promotion is aided by ROS generation, which has also been linked to obesity [\[50\]](#page-18-20). The formation of ROS and the release of proinflammatory cytokines are induced by hyperglycemia combined with increased levels of free fatty acids, which together cause mitochondrial and DNA damage [\[51\]](#page-18-21). Furthermore, the folding of proteins is affected by oxidative stress. Obese people have higher amounts of free fatty acids (FFAs), which are linked to endoplasmic reticulum (ER) stress in adipocytes [\[52\]](#page-18-22). FFAs cause the production of ROS, which oxidize proteins and raise the proportion of unfolded proteins in ER. An inflammatory reaction is brought on by the build-up of unfolded proteins [\[53\]](#page-18-23). Cytokines have been connected to colon cancer in this build-up [\[54\]](#page-18-24).

All these factors could help explain the different incidence of obesity-induced neoplasms in the two sexes.

Finally, the molecular processes underlying the links between gender disparities, oxidation, and the development of cancer have been defined in certain research. The main operating factor in the defense against cancer is the tumor suppressor protein p53, which plays a crucial role in protecting against long-term DNA damage. As a transcription factor, p53 promotes the expression of its target genes by interacting with DNA responsive sequences in their regulatory regions [\[55\]](#page-19-0). Through the p53-DREAM pathway, which includes its primary transcriptional target, the cyclin-dependent kinase inhibitor 1A, also known as P21, which is encoded in CDKN1A, can also suppress other sets of genes [\[56\]](#page-19-1). There is growing evidence that certain cancer sex disparities are related to differences in p53 functional abilities between males and females [\[57\]](#page-19-2). This suggests that either innate or externally imposed effects prevent p53 from conducting its functions equally between the sexes [\[58\]](#page-19-3) (Figure [2\)](#page-4-0). A change in p53 function results in a decrease in the quantity of mitochondria, and wild-type p53 has the ability to affect mitochondrial biogenesis. Additionally, p53 appears to be able to stop mitochondrial DNA mutation [\[59,](#page-19-4)[60\]](#page-19-5). Interestingly, mitochondria may be more adapted to female circumstances than to male situations because they are exclusively of maternal origin [\[61\]](#page-19-6). As a result, an increase in ROS promotes the accumulation of p53, because p53 is tightly tied to redox processes [\[62\]](#page-19-7). The p53 activation pathways that result in ferroptosis, or programmed cell death, depend heavily on acute ROS stimulation [\[63\]](#page-19-8). Wild-type p53 can be made ready to activate repair pathways during a brief cell cycle arrest by sublethal ROS concentrations. However, DNA alteration, such as the TP53 mutation, puts the development of cancer at risk from

sustained ROS levels at sublethal dosages [64]. Therefore, [p5](#page-19-9)3 mutation may play a role in the molecular pathways underlying sex differences in cancer risk.

during a brief cell cycle arrest by sublethal ROS concentrations. However, DNA alteration,

Figure 2. Mutant p53 is associated with a reduction in ATP production, elevated oxidative stress, **Figure 2.** Mutant p53 is associated with a reduction in ATP production, elevated oxidative stress, and persisting DNA damage, which causes cancerogenesis.

1.4. Gender, Oxidative Stress, and Immunity 1.4. Gender, Oxidative Stress, and Immunity

Innate and adaptive immune responses differ between males and females, as do their Innate and adaptive immune responses differ between males and females, as do their immunological responses. Immunological sex differences between men and women can
be seen throughout their life but particularly after puberty and before reproductive senesbe seen throughout their life but particularly after puberty and before reproductive senescence, which suggests that hormones may be at play in some cases. Females tend to have higher innate and adaptive (humoral and cellular) immune responses than males. Genetic and epigenetic factors, sex hormones, and a distinct response to inflammatory stimuli (e.g., oxidative stress) may be the cause of the higher immune response in females. cence, which suggests that hormones may be at play in some cases. Females tend to have
higher innate and adaptive (humoral and cellular) immune responses than males. Genetic
and epigenetic factors, sex hormones, and a dist

The leukocyte cell count in peripheral blood is similar in men and women [\[65\]](#page-19-10). Alhough monocyte and lymphocyte counts have not been found to differ by sex overall [65], though monocyte and lymphocyte counts have not been found to differ by sex overall [\[65\]](#page-19-10), gender dimorphism has been noted in several lymphocyte subsets and natural killer cells. gender dimorphism has been noted in several lymphocyte subsets and natural killer cells. As a result, men seem to have a greater percentage of CD3-CD56+ natural killer cells and As a result, men seem to have a greater percentage of CD3-CD56+ natural killer cells and CD8+ T-cytotoxic lymphocytes, whereas women exhibit a greater proportion of CD4 + T-CD8+ T-cytotoxic lymphocytes, whereas women exhibit a greater proportion of CD4 + helper cells, which leads to a higher CD4/CD8 ratio [66]. T-helper cells, which leads to a higher CD4/CD8 ratio [\[66\]](#page-19-11).

When discussing cytokine differences between the sexes, it is important to differentiate tiate between cytokine levels in the absence of a challenge and cytokine production in between cytokine levels in the absence of a challenge and cytokine production in response to various stimuli. Men seem to have higher amounts of proinflammatory cytokines in relation to basal inflammation $[67,68]$ $[67,68]$.

Oxidative stress is known to activate and stimulate the NF-B transcription factor, Oxidative stress is known to activate and stimulate the NF-B transcription factor, which in turn causes the synthesis of a number of proinflammatory cytokines [\[69\]](#page-19-14), and this might be because of their greater levels of oxidative stress. Females have a higher immunological and inflammatory response than males in terms of the sex differences in immunological and inflammatory response than males in terms of the sex differences in cytokine production in response to a stimulus [\[70\]](#page-19-15). As a result, specific immune cells in \mathbb{Z}^3 women respond to a challenge by producing more TNF and IFN than those in men [71,72]. women respond to a challenge by producing more TNF and IFN than those in men [\[71,](#page-19-16)[72\]](#page-19-17).

Estrogen action has been linked to a greater synthesis of proinflammatory mediators in response to a challenge in females. As a result, normal estrogen concentrations increase the concentration of $\Pi \in \Pi$ and TR in general to the stimulation of homogeneous to the and murine macrophages [\[73\]](#page-19-18). On the other hand, removing endogenous estrogen tends to and murine macrophages [73]. On the other hand, removing endogenous estrogen tends lower the proinflammatory response of the immune cells [\[74\]](#page-19-19). Estrogen action has been linked to a greater synthesis of proinflammatory mediators the production of IL-6, IL-1, and TNF in response to the stimulation of human monocytes

The above findings suggest that differences in the susceptibility and severity of malig-
The above findings suggest that differences in the susceptibility and severity of malig-The above findings suggest that differences in the susceptibility and severity of ma-nancies could be caused by sex differences in immune system performance.

In the next section, we assess how the differences in gender and oxidative stress can Γ affect the start and progression of neoplastic disorders.

2. Sex Differences in Oxidative Stress and Neoplastic Diseases

2.1. Glioma, Oxidative Stress, and Gender Differences

Although it is the second most common cancer in children, brain cancer is an uncommon condition in comparison with other cancer types [\[75\]](#page-19-20). Men are twice as likely to develop medulloblastoma, ependymoma, and gliomas than women, according to epidemiologic research [\[76\]](#page-19-21). In addition, a recent study found that women outlived males and responded better to standard treatment, identifying transcriptome signatures for glioblastomas in women [\[77\]](#page-19-22).

Since oxidative stress and inflammation are also involved in the onset and progression of brain cancer, substances able to modify oxidative stress such as phytoestrogens have been considered good candidates for brain cancer prevention due to their antioxidant and anti-inflammatory properties. In fact, consuming foods containing phytoestrogens, particularly daidzein, appears to have a protective effect against gliomagenesis, according to an epidemiologic study conducted in 2006 [\[78\]](#page-19-23). Additionally, new research has shown that the phytoestrogens formononetin or biochanin A and the cytotoxic drug temozolomide combined have an enhanced anticancer effect in glioblastoma multiforme cells, with greater inhibition of cell signaling and invasion pathways and restoration of mitochondrial function [\[79,](#page-19-24)[80\]](#page-19-25).

Furthermore, long-term research has been conducted on the effects of gender on oxidative stress in the brain, including free radical generation, oxidative damage, and antioxidant enzyme levels and/or activity [\[81\]](#page-19-26). According to certain studies [\[82–](#page-19-27)[91\]](#page-20-0), male rats have greater DNA, protein, and lipid oxidative damage than female rats. The increased ROS generation in male rats [\[92,](#page-20-1)[93\]](#page-20-2) and the decreased levels and/or activities of antioxidant enzymes [\[94](#page-20-3)[–98\]](#page-20-4) are the causes of this oxidative damage. However, although these studies suggest that female rats have better redox homeostasis than male rats, other reports [\[99](#page-20-5)[–102\]](#page-20-6) have found no differences.

In terms of sex hormones, 17-estradiol (E2) and progesterone, which are produced by females, have neuroprotective effects in vivo and in vitro at physiological concentrations [\[103–](#page-20-7)[106\]](#page-20-8), but androgens and testosterone, which are produced by males, typically have neurotoxic effects [\[107\]](#page-20-9). The ability of some neurons and glial cells to create neurosteroids—sex hormones that are often produced de novo and independently of peripheral tissues—is particularly intriguing. These neurosteroids are equivalent to circulating steroids in both chemical and biological terms [\[108,](#page-21-0)[109\]](#page-21-1).

Along with oxidative stress, brain tumorigenesis has also been linked to decreased responses from nonenzyme (reduced glutathione, GSH) and enzyme antioxidant systems (SOD, catalase, and GPx) [\[110\]](#page-21-2). Since the central nervous system (CNS) is extremely susceptible to free radical damage, an imbalance between the production of free radicals and the effectiveness of the antioxidant defense systems is able to initiate the neoplastic process [\[111\]](#page-21-3). This theory is supported by numerous research works. For instance, research has shown that subcutaneous administration of hydroxytyrosol, but not oleuropein or a combination of both compounds, resulted in a significant inhibition of tumor growth through mechanisms involving endogenous enzymatic and nonenzymatic antioxidant defense systems [\[112](#page-21-4)[–114\]](#page-21-5).

Thus, the existence of gender differences in processes related to brain tumors, such as the management of redox status, suggested that research on brain cancer should take gender differences into account in preclinical studies, screening, and prevention programs, as well as in therapeutic approaches.

2.2. Liver Cancer, Oxidative Stress, and Gender Differences

Liver cancer is currently the second most common cancer type [\[115\]](#page-21-6). The 5-year survival rate for people with liver cancer only oscillates by 10%, despite the use of intensive treatments [\[116\]](#page-21-7). In total, 90% of liver cancer cases are caused by hepatocellular carcinoma (HCC).

Even after accounting for variations in exposure to risk factors, there is a two- to four-fold higher incidence of liver cancer in men than in women in humans [\[117](#page-21-8)[,118\]](#page-21-9). Additionally, males predominate in transgenic mouse models of hepatitis virus infection and models of liver tumor induction in mice after exposure to chemical carcinogens such as AFB1, 4-aminobiphenyl (ABP), and diethylnitrosamine (DEN) [\[119\]](#page-21-10). Additionally, numerous human and animal studies on HCC confirmed sexual dimorphism during the onset and development of alcohol liver disease (ALD). It is likely that variations in the expression of genes that code for ethanol-metabolizing enzymes have an impact on the development and progression of ALD and liver cancer [\[120\]](#page-21-11). Alcohol dehydrogenase (ADH) activity varies between sexes; it is lower in men than in women, which leads to less acetaldehyde build-up. Additionally, studies reveal that estrogens positively affect CYP2E1 and ADH, indicating that ethanol should be metabolized more quickly in females than in males [\[121\]](#page-21-12).

Some studies demonstrated that male mice are more vulnerable than female mice to HCC [\[122\]](#page-21-13).

It should be mentioned that lipid peroxide levels in the liver and serum are decreased by estradiol and its derivatives, which are potent endogenous antioxidants [\[123,](#page-21-14)[124\]](#page-21-15). The loss of SOD and glutathione peroxidase activity, as well as iron (ferric nitrilotriacetate) induced ROS production, lipid peroxidation, activation of AP-1 and NF-B, are all suppressed by estradiol in cultured rat hepatocytes, according to recent research [\[125,](#page-21-16)[126\]](#page-21-17). In isolated rat liver mitochondria, estradiol also reduces the lipid peroxidation brought on by iron [\[125\]](#page-21-16). These results imply that the inhibitory impact of estradiol on AP-1 and NF-B activation may result from scavenging ROS and/or from lowering intracellular ROS generation by inducing antioxidant enzymes.

Male sex, like the viral risk factor for hepatic fibrosis, is a significant risk factor for HCC [\[127\]](#page-21-18), while it is unknown whether males and females differ in their susceptibility to the integration of viral DNA, which causes the malignant transformation of hepatocytes. In contrast, premenopausal women are least susceptible to HCC because they lack the risk factors of older age and male sex. In a study, 901 individuals with HBV-associated HCC had their age-specific male-to-female ratios looked at. The younger group had a smaller percentage of females (10.5%) than the older group when the subjects were split into two age groups based on whether they were younger or older than the menopausal age of 50 years.

The differences in hepatic damage were connected to alterations in cellular GSH, ROS production, and cell REDOX status brought on by the metabolism of ethanol. The imbalance between acetaldehyde and ALDH is accentuated by CYP2E1 induction, which also leads to the production of ROS, the subsequent depletion of GSH, and oxidative damage [\[122\]](#page-21-13). Similar results were obtained employing a different experimental model exposing mice to aminobiphenyl (ABP) [\[128](#page-21-19)[–131\]](#page-21-20).

In contrast, levels of the hepatotoxicity biomarker alanine aminotransferase (ALT) were acutely two-fold higher in male adult mice exposed to ABP, DEN, or carbon tetrachloride (CCl4) than in female adult mice [\[132\]](#page-21-21), while levels of the inflammatory biomarker interleukin-6 (IL-6) did not differ based on sex. While CCl4 produced a 40-fold ALT elevation but without sex differences, treatment of immature mice with either ABP or DEN using conventional tumor-inducing postnatal exposure protocols did not result in an increase in serum ALT or IL-6 levels in either males or females. There was no sex difference in the baseline expression of Ggt1 or Hmox1, but adult females expressed the NRF2-responsive gene Nqo1 at higher levels than adult males. Animals that were still developing sexually revealed no sex difference in the three genes' baseline expression. While CCl4 slightly increased the expression of Ggt1 in both males and females and Nqo1 only in females, postnatal DEN exposure slightly increased the expression of Ggt1 only in male mice and Nqo1 in both sexes. Together, these findings rule out the possibility that postnatal carcinogen exposure in mice results in acute hepatotoxic, inflammatory, or NRF2-activated gene responses that are responsible for the male predominance in liver tumor growth [\[132\]](#page-21-21). These results also imply that when extrapolating putative processes to liver carcinogenesis

models that frequently employ postnatally exposed mice, acute toxicity studies conducted in adult mice should be read with caution. However, the various experimental setups used could be the cause of the disparate results found in the various studies.

Aflatoxin B1 (AFB1) is a strong hepatotoxin and hepatocarcinogen for humans and most other mammalian species, although adult mice are remarkably resistant to it [\[133\]](#page-21-22). *Aspergillus flavus*, a mold that develops on groundnuts, grain, and maize that mice frequently consume, produces AFB1. Cytochrome P450 (CYP) transforms AFB1 in both humans and mice into a reactive AFB1-epoxide that can damage DNA by attaching to the N-7 atom of guanine [\[134\]](#page-22-0). Once produced, the glutathione S-transferase (GST) enzymes in the cytosol can catalyze the conjugation of the AFB1-epoxide with reduced glutathione to detoxify it. Water-soluble aflatoxin mercapturic acids (AFB1-NAC) are eliminated in urine as glutathione conjugates of AFB1-epoxides [\[135\]](#page-22-1). Mice's inherent resistance to AFB1 may be due to CYP isoenzymes' poor capacity to produce reactive epoxides and/or GST isoenzymes' great capacity to produce glutathione conjugates.

The important function of GSTA3 in AFB1 resistance was confirmed by a study that produced glutathione S-transferase (GST) A3 knockout (KO) mice. GSTA3 KO mice are vulnerable to the acute cytotoxic and genotoxic effects of AFB1 [\[136\]](#page-22-2). In contrast to the known higher incidence of liver cancer in males in humans, this study shows that initial vulnerability to AFB1 is greater in female mice and that oval cell response and GSTA3 peroxidase activity may affect susceptibility to cancer development (Table [2\)](#page-7-0).

Table 2. Different effects of experimental models of liver damage in the two sexes.

Other information supports the notion that oxidative stress plays a part in the different onset of liver cancer in the two sexes. According to a study, age-related TBARS accumulation in the liver may be sex-related, because it was more noticeable in old male mice compared with old female mice. Gonadotropic hormones, particularly estrogens, may be the cause of these sex-related variations in the TBARS level [\[137\]](#page-22-3). The connection between estrogens and liver oxidative damage has been shown by numerous in vitro investigations [\[138\]](#page-22-4). Since females at that age are in a reproductive decline stage, hormonal changes alone cannot account for the fact that TBARS in 18-month-old females were higher than in males of the same age. The growth of tumors seen in aged male mice may be linked to gender-specific changes in TBARS. These findings are consistent with some published studies that link declining lipid peroxidation (LPO) levels to increasing tumor size [\[139\]](#page-22-5).

Researchers have studied the activities of total superoxide dismutase (tSOD), Gpx, and catalase (CAT). LPO, quantified in terms of TBARS, was determined by the authors to be a marker of liver oxidative damage. LPO increased with aging in both sexes. In both mouse sexes, tSOD appears to be a dormant antioxidative enzyme. The principal alterations in the liver's antioxidant capacity of aging mice were connected to sex-related increases in CAT and Gpx that were only seen in males. Surprisingly, hepatic tumors developed in more than 60% of 18-month-old men (but not girls), which first appeared at 10 months. The findings indicate that increased liver antioxidant capacity of CAT and Gpx in male mice may be an indication of oxidative stress; increases in CAT and Gpx activities in male mice are strongly correlated with the incidence of hepatic tumors; and significantly increased SOD activity in tumor-bearing mice may have been caused by damage from accumulated hydrogen peroxide H₂O₂ [\[140\]](#page-22-6).

The varied ways that oxidative stress behaves in the two sexes is also intriguing. An experiment revealed that during male senescence, CAT and Gpx significantly changed. In contrast to this, there was little to no change in CAT activity and no appreciable change in Gpx activity in female mice. In general, CAT and Gpx activity were 50% and 85% higher in males than in females. Tumor-bearing mice displayed elevated tSOD activity in contrast to the antioxidant enzyme status of tumor-free mice (inert tSOD activity). Antioxidant enzyme activities are typically thought to vary during or after tumor development [\[141\]](#page-22-7). Most past investigations have suggested that cancer has poor antioxidant enzyme activity [\[142\]](#page-22-8). However, most of them used cell lines, and in some of them, conclusions were reached based on blood sample activity measurements that did not accurately reflect the enzyme levels in the tumor or the affected organ. Manganese superoxide dismutase (MnSOD) expression has been shown to be high in many human cancers and, in some tumors, the level of MnSOD is directly correlated with the tumor grade [\[143\]](#page-22-9). Additionally, Manna et al. demonstrated that MnSOD overexpression in tumors may give tumor cells a survival advantage [\[144\]](#page-22-10). Another author's theory is that tumor cells produce a significant amount of H_2O_2 [\[145\]](#page-22-11), and research showing that tSOD overexpression promotes H_2O_2 generation supports this idea. To fulfil the demands of the increased LPO and H_2O_2 buildup brought on by the increased SOD activity, these facts may explain why males generally have higher CAT and Gpx activities [\[146\]](#page-22-12).

Numerous studies have demonstrated that oxidative stress restricts the ability of cells to undergo mitosis, suggesting that oxidative stress may also condition a different proliferative capacity of cancerous cells [\[147\]](#page-22-13). Based on higher antioxidant enzyme levels and the oxidative stress situation prevalent in men, it is possible to infer that cell division favoring clonal growth can occur. Such a phenomenon might aid in the development of cancer. Similar findings have been published from Gonzales, where higher antioxidant levels have been linked to a faster rate of cell division [\[148\]](#page-22-14). Like the gender difference in the incidence of liver cancer in humans, postnatal exposure of mice to ABP causes a higher incidence of liver tumors in males than in females. ABP-DNA adducts that start tumor growth are produced because of first N-hydroxylation that is initially mediated by CYP1A2, according to a conventional theory of ABP carcinogenesis. CYP2E1 was found to be a key ABP N-hydroxylating enzyme in isozyme-selective inhibition tests employing liver microsomes from wild-type and genetically engineered mice. Oxidative stress was brought on by the N-hydroxylation of ABP by transiently expressed CYP2E1 in cultured mouse hepatoma cells. Male wild-type mice exposed postnatally to a tumor-causing dosage of ABP also experienced oxidative stress, but neither male Cyp2e1(/) mice nor female mice did. However, females showed a stronger NRF2-associated antioxidant response [\[149\]](#page-22-15). These results imply that CYP2E1 is a novel ABP-N-oxidizing enzyme and that sex differences in tumor incidence and cell proliferation may be related to sex differences in oxidative stress and antioxidant responses to ABP.

Finally, a particularly exciting area of research focuses on the relationships between gender differences, obesity, oxidative stress, and liver cancers. Recent population-based studies have repeatedly demonstrated that obese men are far more likely to acquire HCC. Men with a BMI of 35 kg/m² showed a severe 4.52-fold increase in relative risk of mortality from liver cancer, although women only showed a small 1.68-fold increase, according to prospective research involving more than 900,000 persons [\[150\]](#page-22-16). The large gender-based variation in HCC incidence has been further validated by a cohort study of 5.24 million persons in the UK [\[151\]](#page-22-17). According to the studies, BMI and HCC in males

were correlated [\[151\]](#page-22-17), and increased and disordered ROS production in extra adipose tissue during obesity may increase oxidative stress and the likelihood of developing HCC [\[152\]](#page-22-18). In contrast to subcutaneous fat accumulation, visceral fat deposition is substantially higher in males than in females [\[153\]](#page-22-19). In numerous datasets [\[154](#page-22-20)[,155\]](#page-22-21), men were found to have larger visceral fat and liver fat contents than women, despite having similar total fat and BMI values. Liver cancer is facilitated by visceral fat, which actively secretes carcinogenic adipokines that cause persistent inflammation. High androgen receptor density may be the root cause of the differences between liver cancer and visceral fat accumulation [\[156\]](#page-22-22). As people get older, their visceral body fat increases, while their subcutaneous body fat decreases, which is correlated with an increase in the incidence of HCC [\[157\]](#page-22-23).

2.3. Colorectal Cancer, Oxidative Stress, and Gender Differences

Colorectal cancer (CRC) is the second most prevalent cause of cancer mortality among men and women globally [\[158\]](#page-22-24). Drug resistance and adverse reactions continue to hinder the success of treatment, despite the fact that the overall survival rate of CRC patients has increased because of advancements in treatment methods such as chemotherapy.

According to certain research, the disease affects people of various sexes at different rates, and this could be due to oxidative stress. For instance, neutrophils and monocytes both contain the lysosomal enzyme myeloperoxidase (MPO) [\[159\]](#page-22-25). Hypochlorous acid, a potent oxidant produced by MPO for its microbicidal function, can target proteins, nucleic acids, and unsaturated lipids by simultaneously releasing ROS [\[160\]](#page-22-26). A-463 G>A transition, which is situated in the consensus binding location of the SP1 transcription factor, is a frequently occurring polymorphism in the MPO gene promoter region. In vitro, the MPO G wild-type allele confers approximately twenty-five times more transcriptional activation than the -463 A variant. According to reports, this polymorphism raises the likelihood of developing laryngeal, lung, breast, and stomach cancers [\[161–](#page-23-0)[165\]](#page-23-1). According to a study, those with the genotype GA/AA were considerably less likely to get colorectal cancer than people with the GG genotype. The reduced risk was particularly significant among men according to the stratified analysis. For male individuals with the GA/AA genotype compared with GG genotype, the adjusted OR was 0.47. However, among women, the OR was not statistically significant. The possibility that estrogen-induced increased MPO-463 A promoter activity is the cause of the MPO-463 A variant's lack of protective effect in female patients is therefore plausible [\[166\]](#page-23-2).

Oxidative stress and cancer have been linked in other research. Bilirubin is more than only the byproduct of heme catabolism. It is now thought to be an essential blood component that forms endogenously and has anti-inflammatory and antioxidant activities [\[167](#page-23-3)[–172\]](#page-23-4). Recent research has indicated that bilirubin, particularly unconjugated bilirubin (UCB), may provide protection against oxidative stress-related illnesses such as CRC. In vitro research outcomes also demonstrated that UCB has antimutagenic qualities [\[173\]](#page-23-5), which may be especially pertinent for gut health. Tetrapyrroles, a family of bile pigments that are abundant in the intestine, reduced the genotoxicity brought on by poly-/heterocyclic amines and triggered apoptosis in cancer cells [\[174](#page-23-6)[–176\]](#page-23-7). Higher circulating UCB concentrations were positively linked with CRC risk in males and negatively associated with risk in women, according to a study that examined relationships between UCB and CRC risk in the European Prospective Investigation into Cancer and Nutrition (EPIC) study [\[177\]](#page-23-8). According to one study, every one standard deviation increase in log-UCB was associated with a lower risk of CRC in males and a higher risk in women (heterogeneity = 0.4 for differences between men and women) [\[178\]](#page-23-9). Finally, it has been demonstrated that UCB may easily cross cell membranes in vivo, infiltrate colon cancer cells to stop tumor cell growth [\[179\]](#page-23-10), trigger death in cancer cells in vitro [\[180\]](#page-23-11), and control gene transcription (via ERK, p53, and p27) [\[181\]](#page-23-12). Strogen, lower NADPH-oxidase activity, or other previously described mechanisms may make women less susceptible to oxidative stress [\[23\]](#page-17-17).

2.4. Lung Cancer, Oxidative Stress, and Gender Differences

Lung cancer is the most common cancer in the world [\[182](#page-23-13)[,183\]](#page-23-14). There may be gender disparities in lung cancer incidence, according to epidemiologic data [\[184–](#page-23-15)[186\]](#page-23-16). Agreeing to several studies, women may be more likely than men to acquire lung and colon cancer from smoking cigarettes [\[187,](#page-23-17)[188\]](#page-24-0).

The expression of genes relevant to cancer and the immune system is altered by genetic and epigenetic alterations, as well as by the abnormal expression of noncoding RNAs, which predisposes the lung epithelium to carcinogenesis. Smoking-related oxidative stress contributes to decreased genomic integrity and promotes epithelial–mesenchymal transition and the creation of a chronic inflammatory milieu. Although not all smokers develop lung cancer, this results in abnormal immune reactions that support the development of cancer. Females are more likely to accumulate oxidative stress damage due to gender differences in the metabolism of cigarette smoke, which increases their risk of developing lung cancer [\[189\]](#page-24-1). Additionally, ROS and RNS can activate signaling molecules such as HIF1, which is a key regulator of angiogenesis and a driving force behind the development of tumors [\[190\]](#page-24-2). Furthermore, it has been demonstrated that the byproducts of ROS and inflammation can inactivate PTEN, a tumor suppressor gene that is frequently altered in lung cancer, by creating an intramolecular disulfide bond [\[191,](#page-24-3)[192\]](#page-24-4).

Large epidemiological studies have demonstrated that for every pack-year of smoking, women are two to three times more likely to die from COPD than males [\[193\]](#page-24-5) and are 50% more likely to develop COPD than men. One explanation is that because women's lungs are smaller than men's with comparable smoking histories, the harm from oxidative stress is more obvious in women [\[193\]](#page-24-5). Another is sex variations in the metabolism of tobacco: women have higher liver CYP1A1 and CYP1B1 activity levels, which activate specific tobacco smoke components to create ROS [\[194\]](#page-24-6). Strogen's role in activating CYP enzymerelated pathways is a contributing factor in the enhanced CYP expression in females [\[195\]](#page-24-7). For instance, a study of smokers who developed lung cancer showed that females had higher levels of CYP1A1 expression and a commensurate rise in DNA adducts, even in lung tissue that was not cancerous [\[196\]](#page-24-8). Additionally, studies on animals showed that the injection of naphthalene—a substance found in tobacco smoke—caused more airway damage in female mice than in male mice. This was due to increased CYP enzyme expression and the production of metabolites, which led to a more severe inflammatory response in the airways and produced more ROS than in male mice [\[197\]](#page-24-9). Because women are more frequently exposed to biomass smoke, exposure to indoor and outdoor air pollution is also a significant risk factor for the development of lung cancer in nonsmokers [\[198,](#page-24-10)[199\]](#page-24-11).

The varied ways that oxidative stress affects the incidence of pulmonary neoplasia in the two sexes could be explained by other processes, as reported in studies performed employing a class of pervasive environmental pollutants known as polycyclic aromatic hydrocarbons (PAHs) [\[200–](#page-24-12)[202\]](#page-24-13).

A study identified sixteen environmental PAHs in workplaces and assessed that women who worked in the office, next to the coke oven, or on its bottom or side, respectively, had significantly higher urine 8-OHdG and 8-isoPGF2a levels and lymphocytic micronucleus frequencies than men who worked in those locations. Gender and BPDE-Alb adducts had a strong impact on rising micronucleus frequencies. The foregoing gender disparities were more pronounced in the median- and high-exposure groups, according to authors who further stratified all workers based on the tertiles of urinary ROH-PAHs or plasma BPDE-Alb adducts [\[203\]](#page-24-14). As a result, women were more vulnerable than males to the oxidative stress and chromosomal damage caused by PAHs, which could be additional evidence for gender differences in PAH-exposure-related lung carcinogenesis (Table [3\)](#page-11-0).

2.5. Melanoma, Oxidative Stress, and Gender Differences

Since the middle of the 1950s, malignant melanoma prognoses for cases with advanced metastases have remained dismal [\[204](#page-24-15)[,205\]](#page-24-16). Gender has been shown to be an independent prognostic factor of melanoma survival in numerous studies, as it remains significant after adjusting for nearly all known prognostic indicators, including age, Breslow thickness, Clark level of invasion, body site, histological subtype, and recently, emerged prognostic indicators, such as ulceration, sentinel node status, and mitotic rate [\[206,](#page-24-17)[207\]](#page-24-18). Both the incidence and survival of malignant melanoma differ significantly across gender. Male patients advance more quickly to stage III [\[208\]](#page-24-19) and maybe even stage IV melanoma [\[209,](#page-24-20)[210\]](#page-24-21); male original melanomas appear to grow more quickly than those in females; and men present with nodal and visceral metastases more frequently than women [\[206\]](#page-24-17). Instead, women are more likely to present with tumors that are in an earlier stage, have longer survival times, and experience better outcomes [\[211](#page-24-22)[–215\]](#page-25-0).

Table 3. Effects of different experimental models of lung damage.

More and more evidence points to the involvement of oxidative stress, which is brought on by high amounts of ROS, such as superoxide anions and hydrogen peroxide, in the development of melanoma [\[216](#page-25-1)[,217\]](#page-25-2). When compared with nearby tissues or melanocytes, melanoma cells produce a lot of ROS, which they then excrete into extracellular space [\[218\]](#page-25-3). Additionally, melanoma cells have elevated intracellular ROS levels [\[219\]](#page-25-4).

High amounts of oxidative stress are known to exist in the initial melanoma tumor environment [\[220](#page-25-5)[–222\]](#page-25-6); tumor-related immune cells release ROS [\[223\]](#page-25-7), and ultraviolet (UV) radiation further intensifies oxidative stress in the skin and melanocytes [\[224\]](#page-25-8). In contrast to surrounding nontumor tissue, benign melanocytic nevi, and control subject skin, Sander et al. discovered a considerable upregulation in antioxidant enzymes in human melanoma biopsies, indicating that the melanoma cells were responding to increasing oxidative stress [\[221\]](#page-25-9).

According to a different study, the advantage that females have in terms of melanoma survival is likely due to sex differences in the capacity to counteract the oxidative stress brought on by ROS [\[220\]](#page-25-5). In fact, it appears that the oxidative environment in the skin of male and female mice has different baseline characteristics; UV-induced oxidative stress amplifies these differences. In comparison with female hairless mice, the skin of males had a lower baseline level of antioxidant enzymes and a roughly 10-fold lower antioxidant functional capacity. In comparison with levels found in the skin of male mice exposed to UVB radiation, the skin of female mice showed a significantly higher induction in antioxidant level, greater antioxidant functional capacity, and lower levels of 8-oxo-deoxyguanosine, the most common type of DNA damage caused by ROS [\[225\]](#page-25-10). These findings were supported by an experiment that looked at gender differences in the development of cancer linked to UV-induced chronic inflammation [\[226\]](#page-25-11). According to the finding's, photoaging damage was present in both male and female mice at the ninth week. However, only male mice in the third week developed skin tumors. Additionally, UV increased the expression of the p65, p-p65, IL-6, and TNF proteins in skin, and these factors were more elevated in the male mouse model. The parameters of blood systemic inflammation were altered to variable degrees in the model groups, according to hematology data, whereas the internal organs of both model groups revealed varying degrees of inflammatory cell infiltration, according to pathology results. These findings suggest that UV-induced skin inflammation, carcinogenesis, and systemic damage differ between the sexes.

Additionally, it is possible that men's higher ROS levels encourage the selection of ROS-resistant melanoma cells. Consequently, ROS can promote melanoma cells' capacity for metastatic spread. Additionally, because men have weaker antioxidant defenses, the ROS that melanoma cells produce damage surrounding healthy tissues more severely, which promotes metastasis. As a result, ROS could account for the reported disparities in melanoma survival between males and females [\[220\]](#page-25-5).

After menopause, according to some researchers, the female advantage vanishes [\[207\]](#page-24-18). Others, however, discovered that females continue to live longer even after menopause [\[227\]](#page-25-12). In female rats, ovariectomies boosted peroxide generation in liver cells to levels seen in male cells, decreased antioxidant enzyme levels to those found in male cells, and restored both peroxide and antioxidant enzyme levels in female cells to the control female levels [\[23\]](#page-17-17). This team discovered that 17-b-estradiol decreased hydrogen peroxide production when isolated mitochondrion was incubated with it [\[228\]](#page-25-13).

The effect of antioxidant supplementation on the incidence of melanoma has also been studied; however, due to the small number of events in the trials, no significant effect [\[229\]](#page-25-14), or even a negative effect [\[230\]](#page-25-15), was discovered. More importantly, the effect of antioxidants varied by gender in each of these studies, affecting both the incidence of melanoma [\[230\]](#page-25-15) and all cancers [\[231\]](#page-25-16). This strongly implies that gender has a role in the relationship between melanoma and ROS.

2.6. Non-Hodgkin Lymphoma, Oxidative Stress, and Gender Differences

The Swedish Lymphoma Register was used in a population-based cohort study that looked at gender differences in the incidence of lymphoma subtypes and excess mortality among people diagnosed between 2000 and 2019 [\[232\]](#page-25-17). Poisson regression was used to predict the male-to-female incidence rate ratios (IRRs) and excess mortality ratios (EMRs) after adjusting for age. They discovered 36,795 instances of lymphoma, 20,738 (56.4%) of which were in men and 16,057 (43.6%) in women. Incidence rate ratios (IRRs) ranged from 1.15 in follicular lymphoma to 5.95 in hairy cell leukemia, with men being considerably more at risk for 14 of the 16 subtypes of lymphoma. Although only statistically significant for classical Hodgkin lymphoma 1.26, aggressive lymphoma not otherwise specified 1.29, and small lymphocytic lymphoma 1.52, EMRs > 1 was seen in 13 out of 16 lymphoma subtypes, indicating higher mortality in men. Similar findings were obtained from a related analysis utilizing information from the Danish Lymphoma Register [\[232\]](#page-25-17). In conclusion, researchers found that for the majority of lymphoma subtypes, men had a significantly greater incidence and a tendency toward higher death rates.

The differing levels of oxidative stress experienced by the two sexes may contribute to the development and spread of lymphomas. For instance, a study in [\[233\]](#page-25-18) examined the idea that lymphomagenesis following low-dose radiation is aided by mitochondrial malfunction and elevated superoxide levels in thymocytes overexpressing Bax (Lck-Bax1 and Lck-Bax38&1). Single whole-body doses of 10 or 100 cGy of 137Cs, iron ions, or silicon ions, were administered to Lck-Bax1 single-transgenic and Lck-Bax38&1 double-transgenic mice. In female Lck-Bax1 mice, a 10 cGy dosage of 137Cs markedly increased the incidence and development of thymic lymphomas. In contrast to silicon ions, a 100 cGy dosage of high-LET iron ions significantly and dose-dependently accelerated lymphomagenesis in both male and female Lck-Bax38&1 mice. Lck-Bax38&1 overexpressing animals were bred with Sirtuin 3 knockouts, a mitochondrial protein deacetylase that controls superoxide metabolism, to ascertain the contribution of mitochondrial oxidative metabolism. Significant increases in thymocyte superoxide levels and accelerated lymphomagenesis were seen in Sirt3//Lck-Bax38&1 animals [\[233\]](#page-25-18) (Table [4\)](#page-13-0). These findings demonstrate that radiation exposure increases lymphomagenesis in Bax overexpressing animals in a manner that depends on both LET and gender. These results are consistent with the hypothesis that in Lck-Bax transgenic mice, mitochondrial dysfunction increases superoxide levels and speeds up lymphomagenesis.

Table 4. Different risk rates of cancer according to oxidative stress mechanisms in males and females.

3. Gender Differences, Oxidative Stress, and Responsiveness to Anticancer Therapy

The possibility that oxidative stress in relation to gender may affect the effectiveness and toxicity of chemotherapy and radiotherapy in cancer subjects is unquestionably an exciting area for future research.

By interfering with mitochondrial function, bioenergetics, signaling pathways, and redox balance, anthracyclines can cause cell malfunction and death [\[234\]](#page-25-19). The majority of these targets exhibit sexual dimorphism, including "redox features" of cells, such as altered redox-associated molecules and enzymes in relation to gender differences in terms of intracellular production and biochemical activity of reactive species, as well as expression of genes related to mitochondria [\[235](#page-25-20)[,236\]](#page-25-21). Along with pharmacodynamics, sex-related variations in pharmacokinetics may have significant clinical ramifications, since they impact the side effects of certain drugs.

Men appear to have a considerably higher doxorubicin clearance than women [\[237\]](#page-25-22). In fact, doxorubicinol levels have been found to be higher in men, which may be connected to greater aldoketoreductase activity [\[238\]](#page-25-23). A lower expression of p-glycoprotein in females may also cause doxorubicin and doxorubicinol to accumulate, increasing the risk of cardiotoxicity. The pharmacokinetics of epirubicin have also been shown to differ according to sex [\[238\]](#page-25-23).

Anthracyclines' overproduction of ROS and RNS causes redox stress, which then causes cardiac injury [\[239\]](#page-26-0), DNA damage, lipid peroxidation, membrane injury, and/or apoptosis, as well as changes in the enzymatic activity of the mitochondrial redox system. The respiratory complexes, Krebs cycle enzymes, oxidative phosphorylation, oxidation, and nitric oxide synthases (NOSs) are among the enzymes changed [\[240](#page-26-1)[–242\]](#page-26-2).

The various oxidative stresses that influence cardiotoxicity in a gender-specific way could in theory be sustained by a complicated inter-relationship between estrogen receptors and enzyme activity engaged in redox processes. Adult male SH rats with tumors seem to be more cardiosensitive to doxorubicin treatment than female rats or hormone-deficient rats, which lends credence to this theory [\[243\]](#page-26-3). According to these findings, doxorubicininduced cardiotoxicity is regulated by reproductive hormones, and the selective cytotoxic mechanism probably works by increasing oxidative stress and apoptosis in male SH rats [\[243\]](#page-26-3).

The anthracycline drug doxorubicin (Dox) is highly effective against a number of neoplastic illnesses but also causes dose-limiting cardiotoxicity [\[244,](#page-26-4)[245\]](#page-26-5). Congestive heart failure, for instance, occurs following Dox treatment at a rate of about 4% at doses of 500–550 mg/m 2 , but this rate rises to 18% at doses of 55–600 mg/m 2 , and it reaches 36% in patients receiving $>601 \text{ mg/m}^2$ of Dox [\[246\]](#page-26-6). A continued follow-up of the cardiac condition of patients that received anthracyclines is recommended, since cumulative and late-onset progressive cardiotoxicity might be seen even decades after treatment [\[247\]](#page-26-7).

In the mitochondria, doxorubicin builds up and causes an excessive amount of ROS production. Male adults (15–55 years old) are more likely than females to have cardiovascular disease overall [\[248–](#page-26-8)[252\]](#page-26-9). Doxorubicin treatment for women causes cardiac failure in 6–20% of adults and 40% of pediatric patients [\[13\]](#page-17-12). Additionally, postmenopausal women are more susceptible to cardiac stress than men their own age following Dox treatment [\[253\]](#page-26-10). Finally, Dox-induced cardiotoxicity in prepubescent girls was found to be more severe than in boys of the same age [\[253\]](#page-26-10). Additionally, according to recent clinical reports, male adults and young girls are more cardiosensitive to Dox. Adult male SHRs with tumors are more cardiosensitive to Dox than female or hormone-deficient animals. In fact, the selective cytotoxic mechanism is thought to work because oxidative stress and apoptosis are more strongly activated in male SHRs, and this suggests that Dox-induced cardiotoxicity inhibits or negatively regulates reproductive hormones [\[254\]](#page-26-11).

Estrogen may act as a cardioprotectant by reducing left ventricular hypertrophy, preventing cardiomyocyte death, and protecting against the onset of cardiac fibrosis in females [\[255\]](#page-26-12). Estrogen, and possibly testosterone, may protect the heart against excessive drug-induced oxidative damage.

Although gender is known to affect how the body reacts to radiation, the underlying molecular mechanisms are not clear, and consequently, current risk estimates are uncertain and have low-resolution dose limits [\[256\]](#page-26-13). It is recommended to use male and female reference phantoms for study; however, few health authorities have defined dose limits with sex-specific regimens [\[257\]](#page-26-14).

Predicting cellular outcomes from proteins related to DNA damage and repair has highlighted the qualitative variations in ionic radiation (IR)-induced reactions between age and sex. Juvenile girls and males appear to start separate signaling cascades as opposed to merely altering the response intensity of the same mechanism. Although both share a suppression of cell cycle progression, males appear to shift towards proapoptosis with mitochondrial stress and reduced DNA repair, while females display activated DNA repair and prosurvival mechanisms. Inflammatory regulators also seem to compete for control over the activation and inhibition of immunological responses; however, they were not found in females [\[258\]](#page-26-15).

To comprehend the effects of ionizing radiation, radiobiology is key. The interaction of radiation with targeted and nontargeted cells, tissues, and organs has profound implications on both the early and late development of primary and secondary malignancies, but knowledge of the mechanisms underlying radiation carcinogenesis is still lacking. Some studies, however, have suggested that nontargeted effects may contribute to a higher risk of developing cancer [\[259\]](#page-26-16).

Males and females experience different rates of radiation-induced mutation patterns, subsequent changes in gene expression and epigenetic status, and malignancies [\[260–](#page-26-17)[263\]](#page-26-18). According to research by Korturbash et al., local cerebral irradiation of mice causes DNA damage and changes in global DNA methylation that are tissue- and sex-dependent. They demonstrated that although nontargeted effects can result in skin hypomethylation, they have not been studied for spleen hypomethylation. They also found that males appear to be more obviously hypomethylated than females [\[264\]](#page-26-19). In nontargeted tissues, similar outcomes have been seen for the control of the microRNAome and inflammatory responses [\[265](#page-27-0)[,266\]](#page-27-1).

Following exposure to X or gamma rays, oxidative DNA damage and cell death are brought on by free radical production which is due to the interaction of ionizing radiation with water molecules and redox-mediated biological pathways. In both the nucleus and the mitochondria, the interaction of free radicals with DNA causes various forms of DNA oxidation. Further DNA damage can result from oxidative stress, inflammatory reactions, and cell death due to necrosis or apoptosis. Patients with cancer and those receiving radiation for their malignancies both have higher levels of oxidized cell-free DNA [\[267–](#page-27-2)[269\]](#page-27-3).

Precision radiation oncology in cancer treatment and individualized risk evaluation for ionizing radiation (IR) exposure are still in their infancy [\[270\]](#page-27-4). Improved medicines with fewer toxicities might be developed if there was a better understanding of the sex-related mechanisms of protection and harm [\[271\]](#page-27-5).

Another approach would be to use natural substances for the modification of oxidative stress [\[272,](#page-27-6)[273\]](#page-27-7). As mentioned above, several natural products have an enhanced anticancer effect via a restoration of mitochondrial function [\[79,](#page-19-24)[80\]](#page-19-25). These substances could reduce the negative effects of sex on the progression of neoplastic diseases, perhaps allowing a reduction in the dosage of traditional anticancer drugs.

4. Final Remarks and Conclusions

A vast range of systems involved in the redox characteristics of cells are affected by oxidative damage [\[235](#page-25-20)[,274\]](#page-27-8). Women often live longer than men as a result of the benefits of their X chromosomes, the antioxidant protective properties of estrogen, and a lower exposure to extrinsic risk factors such as alcohol and smoking. Sex hormones alter the expression of several crucial transcription factors that control ROS-induced stress and in vivo responses. Due to estrogen, women have lower levels of ROS generation and mitochondrial damage than men, which is also linked to increased mitochondrial function and disease resistance. Furthermore, estrogen benefits females by influencing NRF 2 activation and the regulation of other antioxidant-related transcription factors through NRF2. Effective cancer treatment necessitates an awareness of the potential of ROS and a focus on the traits of the study target, such as the patient's gender [\[275\]](#page-27-9). ROS have a variety of biochemical targets in cells.

Males exhibit higher rates of oxidative damage than females [\[276–](#page-27-10)[278\]](#page-27-11); in fact, after adjusting for smoking and body mass index, healthy males have a 29% higher level of urinary oxidative damage [\[279\]](#page-27-12). Males also express lower levels of antioxidants than females, such as GSH, catalase, and SOD. Given that oxidative damage can result in cancer and cardiovascular disease, this could explain why women generally live longer than men [\[280\]](#page-27-13).

The genetic overexpression of antioxidant enzymes in females, which may be brought on by estrogen receptor activation, may also be important. In fact, ovariectomy-induced menopause in mice likely increases oxidative damage susceptibility. Although androgens, such as testosterone, appear to weaken these same defense mechanisms, estrogen levels do not entirely account for these variations in antioxidant defense [\[281\]](#page-27-14).

Regarding mitochondrial function, the traditional view of mitochondrial inheritance holds that mtDNA is only passed down through the female line, yet some reports have suggested that it may also be inherited from the father [\[282](#page-27-15)[–284\]](#page-27-16). The different male/female sex hormones that control mitochondrial energy, OXPHOS, and $Ca²⁺$ homeostasis, may be one of the causes of mitochondrial sexual dimorphisms [\[285\]](#page-27-17). Although this varies depending on the tissue in question and the age/hormonal status of the tested subject, female mitochondria typically have a greater functional capacity than male mitochondria [\[286\]](#page-27-18).

The relationship between mitochondrial activity, gender variations, and cancers can thus be explained in a number of ways.

The thymidine phosphorylase (TP) enzyme contributes to the lowering of TP activity during the metabolism of pyrimidines. It disrupts the nucleotide pool by creating a buildup of thymidine and deoxyuridine. The result is aberrant mitochondrial DNA that displays point mutations, many deletions, and depletion [\[287\]](#page-27-19).

In response to cellular stress conditions, such as inflammation and oxidative damage, TP overexpression takes place and stimulates the Pi3 kinase/Akt pathway, the apoptotic caspase 3/9 pathway, and the autophagic BNIP3 gene with an antiapoptotic activity that promotes proliferation [\[288,](#page-27-20)[289\]](#page-27-21). TP is pathologically overexpressed in a number of human tumors and is associated with a bad prognosis [\[290\]](#page-27-22). The plasma of neoplastic patients has also been found to contain TP-related proteins [\[291\]](#page-27-23).

TP overexpression is associated with the carcinogenesis process and may play a predictive function in breast cancer [\[292\]](#page-28-0). During the female menstrual cycle, the glandular and stromal epithelium contain the enzyme TP [\[293\]](#page-28-1). According to one study, cytoplasmic TP overexpression is associated with microvascular density in canine mammary tumors of a severe grade and may be an indicator of breast cancer [\[294\]](#page-28-2).

The actions of hormones on ion channels also form a specific method by which sex hormones, particularly estrogens, influence oncogenesis. The growth and multiplication of cells depend greatly on potassium channels. The cell cycle's G1/S checkpoint requires membrane hyperpolarization, which is accomplished by the outflow of K+ [\[295\]](#page-28-3). Together with its regulatory subunit, KCNE3, the potassium channel KCNQ1 plays a crucial role. In order to recycle potassium at the basolateral membrane, which is necessary for membrane repolarization, KCNQ1/KCNE3 voltage-gated channels are required [\[296\]](#page-28-4). KCNQ1 functions as a tumor suppressor gene that controls cellular growth, innate immunity, signaling pathways that control growth, and inflammation.

When lost or inactive, KCNQ1 acts as a tumor suppressor, because it prevents the development of cancer [\[297\]](#page-28-5). In addition to membrane hyperpolarization, KCNQ1 is also involved in inflammatory response, oxidative stress, stem cell homeostasis, growth regulation signaling pathways, and ion channel function, all of which are associated with oncogenesis. The downregulation of lipid oxidation caused by KCNQ1 knockout has profound impacts on lipid metabolism [\[298\]](#page-28-6), and the transition to lipogenesis as a result of the suppression of fatty acid oxidation appears to be closely linked to oncogenesis.

In KCNQ1 KO and MUC2 KO mice, several oxidative stress-related genes, including cytochrome oxidase P450 enzymes and various glutathione transferases, appear to be dysregulated [\[298\]](#page-28-6). In colon epithelial cells, estrogen encourages the phosphorylation of KCNQ1 and the sequestration of the channel into endocytic vesicles [\[299\]](#page-28-7). The redistributive process only affects females and has no impact on the overall abundance of KCNQ1. In CRC, KCNQ1 is gradually becoming recognized as a key tumor suppressor. Better CRC survival is related to sustained KCNQ1 expression, and KCNQ1 overexpression reduces nuclear catenin accumulation [\[300](#page-28-8)[,301\]](#page-28-9).

Finally, the regulatory sulfonylureas receptor (Sur1, Sur2, and their splicing products Sur2A and 2B) subunits are coupled with the inwardly rectifying K+ subunits Kir6.1 and Kir6.2 and form the ATP-sensitive potassium (KATP) channel complexes. Low intracellular ATP/ADP ratios, second messengers, kinases such as AMPK, and hormones, are all known to trigger KATP channels [\[302\]](#page-28-10). Estrogens may regulate Kir6.2 and Sur2A-B differently in some tissues; for example, they may upregulate them in cardiomyocytes and decrease KATP channel subunits in neurons, as has been shown in female rats with different effects [\[303\]](#page-28-11). The KATP channel subunits are functionally expressed in a variety of cancer cell types, including hepatocellular carcinoma [\[304\]](#page-28-12), human bladder cancer, human gastric cancer, and glioma [\[305\]](#page-28-13). This has been demonstrated by in vitro and ex vivo research. In two animal models of cancer, the Sur2A component was expressed more highly in proliferating cells.

The Kir6.1/2-Sur2A/B subunits are a therapeutic target in breast and kidney malignancies, according to immunohistochemistry/omics/pharmacovigilance data [\[306\]](#page-28-14).

In summary, our survey highlights that greater knowledge of the molecular mechanisms underlying the gender-related disparities in cancer would likely lead to higher levels of precision medicine and improved treatment options for both males and females with neoplastic disorders.

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References

- 1. Dorak, M.T.; Karpuzoglu, E. Gender differences in cancer susceptibility: An inadequately addressed issue. *Front. Genet.* **2012**, *3*, 268. [\[CrossRef\]](https://doi.org/10.3389/fgene.2012.00268) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23226157)
- 2. Ashley, D.J. A male-female differential in tumour incidence. *Br. J. Cancer* **1969**, *23*, 21–25. [\[CrossRef\]](https://doi.org/10.1038/bjc.1969.3)
- 3. Edgren, G.; Liang, L.; Adami, H.O.; Chang, E.T. Enigmatic sex disparities in cancer incidence. *Eur. J. Epidemiol.* **2012**, *27*, 187–196. [\[CrossRef\]](https://doi.org/10.1007/s10654-011-9647-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22212865)
- 4. Cook, M.B.; McGlynn, K.A.; Devesa, S.S.; Freedman, N.D.; Anderson, W.F. Sex disparities in cancer mortality and survival. *Cancer Epidemiol. Biomark. Prev.* **2011**, *20*, 1629–1637. [\[CrossRef\]](https://doi.org/10.1158/1055-9965.EPI-11-0246) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21750167)
- 5. Carlus, M.; Elies, L.; Fouque, M.C.; Maliver, P.; Schorsch, F. Historical control data of neoplastic lesions in the Wistar Hannover Rat among eight 2-year carcinogenicity studies. *Exp. Toxicol. Pathol.* **2013**, *65*, 243–253. [\[CrossRef\]](https://doi.org/10.1016/j.etp.2011.08.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21945048)
- 6. Kadekar, S.; Peddada, S.; Silins, I.; French, J.E.; Högberg, J.; Stenius, U. Gender differences in chemical carcinogenesis in National Toxicology Program 2-year bioassays. *Toxicol. Pathol.* **2012**, *40*, 1160–1168. [\[CrossRef\]](https://doi.org/10.1177/0192623312446527)
- 7. Seifarth, J.E.; McGowan, C.L.; Milne, K.J. Sex and life expectancy. *Gend. Med.* **2012**, *9*, 390–401. [\[CrossRef\]](https://doi.org/10.1016/j.genm.2012.10.001)
- 8. Thannickal, V.J.; Fanburg, B.L. Reactive oxygen species in cell signaling. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2000**, *279*, L1005e28. [\[CrossRef\]](https://doi.org/10.1152/ajplung.2000.279.6.L1005)
- 9. Olinski, R.; Zastawny, T.; Budzbon, J.; Skokowski, J.; Zegarski, W.; Dizaroglu, M. DNA base modifications in chromatin of human cancerous tissues. *FEBS Lett.* **1992**, *309*, 193–198. [\[CrossRef\]](https://doi.org/10.1016/0014-5793(92)81093-2)
- 10. Conner, E.M.; Grisham, M.B. Inflammation, free radicals, and antioxidants. *Nutrition* **1996**, *12*, 274–277. [\[CrossRef\]](https://doi.org/10.1016/S0899-9007(96)00000-8)
- 11. Hlavata, L.; Aguilaniu, H.; Pichova, A.; Nystrom, T. The oncogenic RAS2(val19) mutation locks respiration, independently of PKA, in a mode prone to generate ROS. *EMBO J.* **2003**, *22*, 3337–3345. [\[CrossRef\]](https://doi.org/10.1093/emboj/cdg314) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12839995)
- 12. Vafa, O.; Wade, M.; Kern, S.; Beeche, M.; Pandita, T.K.; Hampton, G.M.; Wahl, G.M. c-Myc can induce DNA damage, increase reactive oxygen species, and mitigate p53 function: A mechanism for oncogene-induced genetic instability. *Mol. Cell* **2002**, *9*, 1031–1044. [\[CrossRef\]](https://doi.org/10.1016/S1097-2765(02)00520-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12049739)
- 13. Ozben, T. Oxidative stress and apoptosis: Impact on cancer therapy. *J. Pharm. Sci.* **2007**, *96*, 2181–2196. [\[CrossRef\]](https://doi.org/10.1002/jps.20874) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17593552)
- 14. Bardelˇcíková, A.; Šoltys, J.; Mojžiš, J. Oxidative Stress, Inflammation and Colorectal Cancer: An Overview. *Antioxidants* **2023**, *12*, 901. [\[CrossRef\]](https://doi.org/10.3390/antiox12040901) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37107276)
- 15. Musolino, C.; Allegra, A.; Saija, A.; Alonci, A.; Russo, S.; Spatari, G.; Penna, G.; Gerace, D.; Cristani, M.; David, A.; et al. Changes in advanced oxidation protein products, advanced glycation end products, and s-nitrosylated proteins, in patients affected by polycythemia vera and essential thrombocythemia. *Clin. Biochem.* **2012**, *45*, 1439–1443. [\[CrossRef\]](https://doi.org/10.1016/j.clinbiochem.2012.07.100)
- 16. Gangemi, S.; Allegra, A.; Alonci, A.; Cristani, M.; Russo, S.; Speciale, A.; Penna, G.; Spatari, G.; Cannavò, A.; Bellomo, G.; et al. Increase of novel biomarkers for oxidative stress in patients with plasma cell disorders and in multiple myeloma patients with bone lesions. *Inflamm. Res.* **2012**, *61*, 1063–1067. [\[CrossRef\]](https://doi.org/10.1007/s00011-012-0498-7)
- 17. Thapa, P.; Jiang, H.; Ding, N.; Hao, Y.; Alshahrani, A.; Wei, Q. The Role of Peroxiredoxins in Cancer Development. *Biology* **2023**, *12*, 666. [\[CrossRef\]](https://doi.org/10.3390/biology12050666) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37237480)
- 18. Jiang, H.; Zuo, J.; Li, B.; Chen, R.; Luo, K.; Xiang, X.; Lu, S.; Huang, C.; Liu, L.; Tang, J.; et al. Drug-induced oxidative stress in cancer treatments: Angel or devil? *Redox Biol.* **2023**, *18*, 102754. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2023.102754)
- 19. Ali, I.; Högberg, J.; Hsieh, J.H.; Auerbach, S.; Korhonen, A.; Stenius, U.; Silins, I. Gender differences in cancer susceptibility: Role of oxidative stress. *Carcinogenesis* **2016**, *37*, 985–992. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgw076)
- 20. Ide, T.; Tsutsui, H.; Ohashi, N.; Hayashidani, S.; Suematsu, N.; Tsuchihashi, M.; Tamai, H.; Takeshita, A. Greater oxidative stress in healthy young men compared with premenopausal women. *ATVB* **2002**, *22*, 438–442. [\[CrossRef\]](https://doi.org/10.1161/hq0302.104515) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11884287)
- 21. Niveditha, S.; Deepashree, S.; Ramesh, S.R.; Shivanandappa, T. Sex differences in oxidative stress resistance in relation to longevity in *Drosophila melanogaster*. *J. Comp. Physiol. B* **2017**, *187*, 899–909. [\[CrossRef\]](https://doi.org/10.1007/s00360-017-1061-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28261744)
- 22. Barp, J.; Araújo, A.S.D.R.; Fernandes, T.R.G.; Rigatto, K.V.; Llesuy, S.; Belló-Klein, A.; Singal, P. Myocardial antioxidant and oxidative stress changes due to sex hormones. *Braz. J. Med. Biol. Res.* **2002**, *35*, 1075–1081. [\[CrossRef\]](https://doi.org/10.1590/S0100-879X2002000900008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12219179)
- 23. Kander, M.C.; Cui, Y.; Liu, Z. Gender difference in oxidative stress: A new look at the mechanisms for cardiovascular diseases. *J. Cell. Mol. Med.* **2017**, *21*, 1024–1032. [\[CrossRef\]](https://doi.org/10.1111/jcmm.13038) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27957792)
- 24. Borras, C.; Sastre, J.; García-Sala, D.; Lloret, A.; Pallardo, F.V.; Vina, J. Mitochondria from females exhibit higher antioxidant gene expression and lower oxidative damage than males. *Free Radic. Biol. Med.* **2003**, *34*, 546–552. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(02)01356-4)
- 25. Miquel, J.; Economos, A.C.; Fleming, J.; Johnson, J.E., Jr. Mitochondrial role in cell aging. *Exp. Gerontol.* **1980**, *15*, 575–591. [\[CrossRef\]](https://doi.org/10.1016/0531-5565(80)90010-8)
- 26. Sohal, R.S.; Sohal, B.H.; Brunk, U.T. Relationship between antioxidant defenses and longevity in different mammalian species. *Mech. Ageing Dev.* **1990**, *53*, 217–227. [\[CrossRef\]](https://doi.org/10.1016/0047-6374(90)90040-M) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2376982)
- 27. Barja, G.; Cadenas, S.; Rojas, C.; Perez-Campo, R.; Lopez-Torres, M. Low mitochondrial free radical production per unit O_2 consumption can explain the simultaneous presence of high longevity and high aerobic metabolic rate in birds (Oct). *Free Radic. Res.* **1994**, *21*, 317–327. [\[CrossRef\]](https://doi.org/10.3109/10715769409056584)
- 28. Barja, G. Updating the mitochondrial free radical theory of aging: An integrated view, key aspects, and confounding concepts. *Antioxid. Redox Signal.* **2013**, *19*, 1420–1445. [\[CrossRef\]](https://doi.org/10.1089/ars.2012.5148)
- 29. Miller, A.A.; Drummond, G.R.; Mast, A.E.; Schmidt, H.H.; Sobey, C.G. Effect of gender on NADPH-oxidase activity, expression, and function in the cerebral circulation: Role of estrogen. *Stroke* **2007**, *38*, 2142–2149. [\[CrossRef\]](https://doi.org/10.1161/STROKEAHA.106.477406)
- 30. Mendoza-Núnez, V.M.; Beristain-Perez, A.; Perez-Vera, S.P.; Altamirano-Lozano, M.A. Age-related sex differences in glutathione peroxidase and oxidative DNA damage in a healthy Mexican population. *J. Women Health* **2010**, *19*, 919–926. [\[CrossRef\]](https://doi.org/10.1089/jwh.2009.1684)
- 31. Kendall, B.; Eston, R. Exercise-induced muscle damage and the potential protective role of estrogen. *Sport. Med.* **2002**, *32*, 103–123. [\[CrossRef\]](https://doi.org/10.2165/00007256-200232020-00003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11817996)
- 32. Karolkiewicz, J.; Michalak, E.; Pospieszna, B.; Deskur-Smielecka, E.; Nowak, A.; Pilaczynska-Szczesniak, L. Response of oxidative stress markers and antioxidant parameters to an 8- week aerobic physical activity program in healthy, postmenopausal women. *Arch. Gerontol. Geriatr.* **2009**, *49*, e67–e71. [\[CrossRef\]](https://doi.org/10.1016/j.archger.2008.09.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18990458)
- 33. Semenova, N.V.; Rychkova, L.V.; Darenskaya, M.A.; Kolesnikov, S.I.; Nikitina, O.A.; Petrova, A.G.; Vyrupaeva, E.V.; Kolesnikova, L.I. Superoxide Dismutase Activity in Male and Female Patients of Different Age with Moderate COVID-19. *Bull. Exp. Biol. Med.* **2022**, *173*, 51–53. [\[CrossRef\]](https://doi.org/10.1007/s10517-022-05491-6)
- 34. Chen, Y.; Ji, L.L.; Liu, T.Y.; Wang, Z.T. Evaluation of gender-related differences in various oxidative stress enzymes in mice. *Chin. J. Physiol.* **2011**, *54*, 385–390. [\[CrossRef\]](https://doi.org/10.4077/CJP.2011.AMM080)
- 35. Sobocanec, S.; Balog, T.; Sverko, V.; Marotti, T. Sex-dependent antioxidant enzyme activities and lipid peroxidation in ageing mouse brain. *Free Radic. Res.* **2003**, *37*, 743–748. [\[CrossRef\]](https://doi.org/10.1080/1071576031000102178)
- 36. Vina, J.; Borras, C.; Gambini, J.; Sastre, J.; Pallardo, F.V. Why females live longer than males: Control of longevity by sex hormones. *Sci. Aging Knowl. Environ.* **2005**, *2005*, pe17. [\[CrossRef\]](https://doi.org/10.1126/sageke.2005.23.pe17)
- 37. Erden Inal, M.; Akgün, A.; Kahraman, A. The effects of exogenous glutathione on reduced glutathione level, glutathione peroxidase and glutathione reductase activities of rats with different ages and gender after whole-body Γ-irradiation. *AGE* **2003**, *26*, 55–58. [\[CrossRef\]](https://doi.org/10.1007/s11357-003-0005-8)
- 38. Alkazemi, D.; Rahman, A.; Habra, B. Alterations in glutathione redox homeostasis among adolescents with obesity and anemia. *Sci. Rep.* **2021**, *11*, 3034. [\[CrossRef\]](https://doi.org/10.1038/s41598-021-82579-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33542364)
- 39. Bellanti, F.; Matteo, M.; Rollo, T.; De Rosario, F.; Greco, P.; Vendemiale, G.; Serviddio, G. Sex hormones modulate circulating antioxidant enzymes: Impact of estrogen therapy. *Redox Biol.* **2013**, *1*, 340–346. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2013.05.003)
- 40. Ferri, J.; Navarro, I.; Alabadí, B.; Bosch-Sierra, N.; Benito, E.; Civera, M.; Ascaso, J.F.; Martinez-Hervas, S.; Real, J.T. Gender differences on oxidative stress markers and complement component C3 plasma values after an oral unsaturated fat load test. *Clin. Investig. Arterioscler.* **2020**, *32*, 87–93. [\[CrossRef\]](https://doi.org/10.1016/j.arteri.2019.11.002)
- 41. Arnold, M.; Pandeya, N.; Byrnes, G.; Renehan, P.A.G.; Stevens, G.A.; Ezzati, P.M.; Ferlay, J.; Miranda, J.J.; Romieu, I.; Dikshit, R.; et al. Global burden of cancer attributable to high body-mass index in 2012: A population-based study. *Lancet Oncol.* **2015**, *16*, 36–46. [\[CrossRef\]](https://doi.org/10.1016/S1470-2045(14)71123-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25467404)
- 42. Kyrgiou, M.; Kalliala, I.; Markozannes, G.; Gunter, M.J.; Paraskevaidis, E.; Gabra, H.; Martin-Hirsch, P.; Tsilidis, K.K. Adiposity and cancer at major anatomical sites: Umbrella review of the literature. *BMJ* **2017**, *356*, j477. [\[CrossRef\]](https://doi.org/10.1136/bmj.j477) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28246088)
- 43. Renehan, A.G.; Zwahlen, M.; Egger, M. Adiposity and cancer risk: New mechanistic insights from epidemiology. *Nat. Rev. Cancer* **2015**, *15*, 484–498. [\[CrossRef\]](https://doi.org/10.1038/nrc3967) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26205341)
- 44. Murphy, N.; Strickler, H.D.; Stanczyk, F.Z.; Xue, X.; Wassertheil-Smoller, S.; Rohan, T.E.; Ho, G.Y.F.; Anderson, G.L.; Potter, J.D.; Gunter, M.J. A prospective evaluation of endogenous sex hormone levels and colorectal cancer risk in postmenopausal women. *J. Natl. Cancer Inst.* **2015**, *107*, djv210. [\[CrossRef\]](https://doi.org/10.1093/jnci/djv210)
- 45. Chlebowski, R.T.; Wactawski-Wende, J.; Ritenbaugh, C.; Hubbell, F.A.; Ascensao, J.; Rodabough, R.J.; Rosenberg, C.A.; Taylor, V.M.; Harris, R.; Chen, C.; et al. Estrogen plus progestin and colorectal cancer in postmenopausal women. *N. Engl. J. Med.* **2004**, *350*, 991–1004. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa032071)
- 46. Avgerinos, K.I.; Spyrou, N.; Mantzoros, C.S.; Dalamaga, M. Obesity and cancer risk: Emerging biological mechanisms and perspectives. *Metab. Clin. Exp.* **2019**, *92*, 121–135. [\[CrossRef\]](https://doi.org/10.1016/j.metabol.2018.11.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30445141)
- 47. Boutari, C.; Mantzoros, C.S. Inflammation: A key player linking obesity with malignancies. *Metabolism* **2018**, *81*, A3–A6. [\[CrossRef\]](https://doi.org/10.1016/j.metabol.2017.12.015)
- 48. Deng, T.; Lyon, C.J.; Bergin, S.; Caligiuri, M.A.; Hsueh, W.A. Obesity, inflammation, and cancer. *Annu. Rev. Pathol.* **2016**, *11*, 421–449. [\[CrossRef\]](https://doi.org/10.1146/annurev-pathol-012615-044359)
- 49. Moore, L.L.; Chadid, S.; Singer, M.R.; Kreger, B.E.; Denis, G.V. Metabolic health reduces risk of obesity-related cancer in framingham study adults. *Cancer Epidemiol. Biomarks Prev.* **2014**, *23*, 2057–2065. [\[CrossRef\]](https://doi.org/10.1158/1055-9965.EPI-14-0240)
- 50. Sabharwal, S.S.; Schumacker, P.T. Mitochondrial ROS in cancer: Initiators, amplifiers or an Achilles' heel? *Nat. Rev. Cancer* **2014**, *14*, 709–721. [\[CrossRef\]](https://doi.org/10.1038/nrc3803)
- 51. Dikalov, S.I.; Nazarewicz, R.R. Angiotensin II-induced production of mitochondrial reactive oxygen species: Potential mechanisms and relevance for cardiovascular disease. *Antioxid. Redox Signal.* **2013**, *19*, 1085–1094. [\[CrossRef\]](https://doi.org/10.1089/ars.2012.4604)
- 52. Balkwill, F.; Charles, K.A.; Mantovani, A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* **2005**, *7*, 211–217. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2005.02.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15766659)
- 53. Iyengar, N.M.; Gucalp, A.; Dannenberg, A.J.; Hudis, C.A. Obesity and cancer mechanisms: Tumor microenvironment and inflammation. *J. Clin. Oncol.* **2016**, *34*, 4270–4276. [\[CrossRef\]](https://doi.org/10.1200/JCO.2016.67.4283) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27903155)
- 54. Yehuda-Shnaidman, E.; Schwartz, B. Mechanisms linking obesity, inflammation and altered metabolism to colon carcinogenesis. *Obes. Rev.* **2012**, *13*, 1083–1095. [\[CrossRef\]](https://doi.org/10.1111/j.1467-789X.2012.01024.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22937964)
- 55. Fischer, M. Census and evaluation of p53 target genes. *Oncogene* **2017**, *36*, 3943–3956. [\[CrossRef\]](https://doi.org/10.1038/onc.2016.502)
- 56. Engeland, K. Cell cycle arrest through indirect transcriptional repression by p53: I have a DREAM. *Cell Death Differ.* **2018**, *25*, 114–132. [\[CrossRef\]](https://doi.org/10.1038/cdd.2017.172)
- 57. Freudenstein, D.; Litchfield, C.; Caramia, F.; Wright, G.; Solomon, B.J.; Ball, D.; Keam, S.P.; Neeson, P.; Haupt, Y.; Haupt, S. TP53 Status, patient sex, and the immune response as determinants of lung cancer patient survival. *Cancers* **2020**, *12*, 1535. [\[CrossRef\]](https://doi.org/10.3390/cancers12061535)
- 58. Beyfuss, K.; Hood, D.A. A systematic review of p53 regulation of oxidative stress in skeletal muscle. *Redox Rep.* **2018**, *23*, 100–117. [\[CrossRef\]](https://doi.org/10.1080/13510002.2017.1416773)
- 59. Lebedeva, M.A.; Eaton, J.S.; Shadel, G.S. Loss of p53 causes mitochondrial DNA depletion and altered mitochondrial reactive oxygen species homeostasis. *Biochim. Biophys. Acta* **2009**, *1787*, 328–334. [\[CrossRef\]](https://doi.org/10.1016/j.bbabio.2009.01.004)
- 60. Liu, J.; Zhang, C.; Hu, W.; Feng, Z. Tumor suppressor p53 and metabolism. *J. Mol. Cell Biol.* **2019**, *11*, 284–292. [\[CrossRef\]](https://doi.org/10.1093/jmcb/mjy070)
- 61. Beekman, M.; Dowling, D.K.; Aanen, D.K. The costs of being male: Are there sex-specific effects of uniparental mitochondrial inheritance? *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2014**, *369*, 20130440. [\[CrossRef\]](https://doi.org/10.1098/rstb.2013.0440) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24864311)
- 62. Eriksson, S.E.; Ceder, S.; Bykov, V.J.N.; Wiman, K.G. p53 as a hub in cellular redox regulation and therapeutic target in cancer. *J. Mol. Cell Biol.* **2019**, *11*, 330–341. [\[CrossRef\]](https://doi.org/10.1093/jmcb/mjz005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30892598)
- 63. Perillo, B.; Di Donato, M.; Pezone, A.; Di Zazzo, E.; Giovannelli, P.; Galasso, G.; Castoria, G.; Migliaccio, A. ROS in cancer therapy: The bright side of the moon. *Exp. Mol. Med.* **2020**, *52*, 192–203. [\[CrossRef\]](https://doi.org/10.1038/s12276-020-0384-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32060354)
- 64. Haupt, S.; Haupt, Y. Cancer and Tumour Suppressor p53 Encounters at the Juncture of Sex Disparity. *Front. Genet.* **2021**, *12*, 632719. [\[CrossRef\]](https://doi.org/10.3389/fgene.2021.632719)
- 65. Choi, J.; Lee, S.J.; Lee, Y.A.; Maeng, H.G.; Lee, J.K.; Kang, Y.W. Reference values for peripheral blood lymphocyte subsets in a healthy korean population. *Immune Netw.* **2014**, *14*, 289–295. [\[CrossRef\]](https://doi.org/10.4110/in.2014.14.6.289)
- 66. Valiathan, R.; Deeb, K.; Diamante, M.; Ashman, M.; Sachdeva, N.; Asthana, D. Reference ranges of lymphocyte subsets in healthy adults and adolescents with special mention of T cell maturation subsets in adults of South Florida. *Immunobiology* **2014**, *219*, 487–496. [\[CrossRef\]](https://doi.org/10.1016/j.imbio.2014.02.010)
- 67. Milan-Mattos, J.C.; Anibal, F.F.; Perseguini, N.M.; Minatel, V.; Rehder-Santos, P.; Castro, C.A.; Vasilceac, F.A.; Mattiello, S.M.; Faccioli, L.H.; Catai, A.M. Effects of natural aging and gender on pro-inflammatory markers. *Braz. J. Med. Biol. Res.* **2019**, *52*, e8392. [\[CrossRef\]](https://doi.org/10.1590/1414-431x20198392)
- 68. Bernardi, S.; Toffoli, B.; Tonon, F.; Francica, M.; Campagnolo, E.; Ferretti, T.; Comar, S.; Giudici, F.; Stenner, E.; Fabris, B. Sex differences in proatherogenic cytokine levels. *Int. J. Mol. Sci.* **2020**, *21*, 3861. [\[CrossRef\]](https://doi.org/10.3390/ijms21113861)
- 69. Meier, A.; Chang, J.J.; Chan, E.S.; Pollard, R.B.; Sidhu, H.K.; Kulkarni, S.; Wen, T.F.; Lindsay, R.J.; Orellana, L.; Mildvan, D.; et al. Sex differences in the Toll-like receptor-mediated response of plasmacytoid dendritic cells to HIV-1. *Nat. Med.* **2009**, *15*, 955–959. [\[CrossRef\]](https://doi.org/10.1038/nm.2004)
- 70. Ortona, E.; Pierdominici, M.; Rider, V. Editorial: Sex Hormones and Gender Differences in Immune Responses. *Front. Immunol.* **2019**, *10*, 1076. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2019.01076)
- 71. Vida, C.; Gonzalez, E.M.; De la Fuente, M. Increase of oxidation and inflammation in nervous and immune systems with aging and anxiety. *Curr. Pharm. Des.* **2014**, *20*, 4656–4678. [\[CrossRef\]](https://doi.org/10.2174/1381612820666140130201734)
- 72. Aomatsu, M.; Kato, T.; Kasahara, E.; Kitagawa, S. Gender difference in tumor necrosis factor-α production in human neutrophils stimulated by lipopolysaccharide and interferon-γ. *Biochem. Biophys. Res. Commun.* **2013**, *441*, 220–225. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2013.10.042)
- 73. Klein, S.L.; Flanagan, K.L. Sex differences in immune responses. *Nat. Rev. Immunol.* **2016**, *16*, 626–638. [\[CrossRef\]](https://doi.org/10.1038/nri.2016.90)
- 74. Rettew, J.A.; Huet, Y.M.; Marriott, I. Estrogens augment cell surface TLR4 expression on murine macrophages and regulate sepsis susceptibility in vivo. *Endocrinology* **2009**, *150*, 3877–3884. [\[CrossRef\]](https://doi.org/10.1210/en.2009-0098)
- 75. Torre, L.A.; Siegel, R.L.; Ward, E.M.; Jemal, A. Global cancer incidence and mortality rates and trends—An update. *Cancer Epidemiol. Biomark. Prev.* **2016**, *25*, 16–27. [\[CrossRef\]](https://doi.org/10.1158/1055-9965.EPI-15-0578)
- 76. Sun, T.; Plutynski, A.; Ward, S.; Rubin, J.B. An integrative view on sex differences in brain tumors. *Cell. Mol. Life Sci.* **2015**, *72*, 3323–3342. [\[CrossRef\]](https://doi.org/10.1007/s00018-015-1930-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25985759)
- 77. Yang, W.; Warrington, N.M.; Taylor, S.J.; Whitmire, P.; Carrasco, E.; Singleton, K.W.; Wu, N.; Lathia, J.D.; Berens, M.E.; Kim, A.H.; et al. Sex differences in GBM revealed by analysis of patient imaging, transcriptome, and survival data. *Sci. Transl. Med.* **2019**, *11*, eaao5253. [\[CrossRef\]](https://doi.org/10.1126/scitranslmed.aao5253) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30602536)
- 78. Tedeschi-Blok, N.; Lee, M.; Sison, J.D.; Miike, R.; Wrensch, M. Inverse association of antioxidant and phytoestrogen nutrient intake with adult glioma in the San Francisco Bay Area: A case-control study. *BMC Cancer* **2006**, *6*, 148. [\[CrossRef\]](https://doi.org/10.1186/1471-2407-6-148)
- 79. Desai, V.; Jain, A.; Shaghaghi, H.; Summer, R.; Lai, J.C.K.; Bhushan, A. Combination of Biochanin A and Temozolomide Impairs Tumor Growth by Modulating Cell Metabolism in Glioblastoma Multiforme. *Anticancer Res.* **2019**, *39*, 57–66. [\[CrossRef\]](https://doi.org/10.21873/anticanres.13079) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30591440)
- 80. Zhang, X.; Ni, Q.; Wang, Y.; Fan, H.; Li, Y. Synergistic anticancer effects of formononetin and temozolomide on glioma C6 cells. *Biol. Pharm. Bull.* **2018**, *41*, 1194–1202. [\[CrossRef\]](https://doi.org/10.1248/bpb.b18-00002)
- 81. Ruszkiewicz, J.A.; Miranda-Vizuete, A.; Tinkov, A.A.; Skalnaya, M.G.; Skalny, A.V.; Tsatsakis, A.; Aschner, M. Sex-Specific Differences in Redox Homeostasis in Brain Norm and Disease. *J. Mol. Neurosci.* **2019**, *67*, 312–342. [\[CrossRef\]](https://doi.org/10.1007/s12031-018-1241-9)
- 82. Candeias, E.; Duarte, A.I.; Sebastião, I.; Fernandes, M.A.; Plácido, A.I.; Carvalho, C.; Correia, S.; Santos, R.X.; Seiça, R.; Santos, M.S.; et al. Middle-Aged Diabetic Females and Males Present Distinct Susceptibility to Alzheimer Diseaselike Pathology. *Mol. Neurobiol.* **2017**, *54*, 6471–6489. [\[CrossRef\]](https://doi.org/10.1007/s12035-016-0155-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27730513)
- 83. Chakraborti, A.; Gulati, K.; Banerjee, B.D.; Ray, A. Possible involvement of free radicals in the differential neurobehavioral responses to stress in male and female rats. *Behav. Brain Res.* **2007**, *179*, 321–325. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2007.02.018)
- 84. Cole, T.B.; Coburn, J.; Dao, K.; Roqué, P.; Chang, Y.C.; Kalia, V.; Guilarte, T.R.; Dziedzic, J.; Costa, L.G. Sex and genetic differences in the effects of acute diesel exhaust exposure on inflammation and oxidative stress in mouse brain. *Toxicology* **2016**, *374*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.tox.2016.11.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27865893)
- 85. Guevara, R.; Santandreu, F.M.; Valle, A.; Gianotti, M.; Oliver, J.; Roca, P. Sex-dependent differences in aged rat brain mitochondrial function and oxidative stress. *Free Radic. Biol. Med.* **2009**, *46*, 169–175. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2008.09.035)
- 86. Katalinic, V.; Modun, D.; Music, I.; Boban, M. Gender differences in antioxidant capacity of rat tissues determined by 2,2'-azinobis (3-ethylbenzothiazoline 6-sulfonate; ABTS) and ferric reducing antioxidant power (FRAP) assays. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **2005**, *140*, 47–52. [\[CrossRef\]](https://doi.org/10.1016/j.cca.2005.01.005)
- 87. Silva, T.L.A.; Braz, G.R.F.; Silva, S.C.A.; Pedroza, A.A.D.S.; Freitas, C.M.; Ferreira, D.J.S.; da Silva, A.I.; Lagranha, C.J. Serotonin transporter inhibition during neonatal period induces sex-dependent effects on mitochondrial bioenergetics in the rat brainstem. *Eur. J. Neurosci.* **2018**, *48*, 1620–1634. [\[CrossRef\]](https://doi.org/10.1111/ejn.13971)
- 88. Sobočanec, S.; Balog, T.; Kušić, B.; Šverko, V.; Šarić, A.; Marotti, T. Differential response to lipid peroxidation in male and female mice with age: Correlation of antioxidant enzymes matters. *Biogerontology* **2008**, *9*, 335–343. [\[CrossRef\]](https://doi.org/10.1007/s10522-008-9145-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18473185)
- 89. Guevara, R.; Gianotti, M.; Roca, P.; Oliver, J. Age and sex-related changes in rat brain mitochondrial function. *Cell. Physiol. Biochem.* **2011**, *27*, 201–206. [\[CrossRef\]](https://doi.org/10.1159/000327945)
- 90. Jung, M.E.; Metzger, D.B. A sex difference in oxidative stress and behavioral suppression induced by ethanol withdrawal in rats. *Behav. Brain Res.* **2016**, *314*, 199–214. [\[CrossRef\]](https://doi.org/10.1016/j.bbr.2016.07.054)
- 91. Uzun, H.; Kayali, R.; Çakatay, U. The chance of gender dependency of oxidation of brain proteins in aged rats. *Arch. Gerontol. Geriatr.* **2010**, *50*, 16–19. [\[CrossRef\]](https://doi.org/10.1016/j.archger.2009.01.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19230989)
- 92. Khalifa, A.R.; Abdel-Rahman, E.A.; Mahmoud, A.M.; Ali, M.H.; Noureldin, M.; Saber, S.H.; Mohsen, M.; Ali, S.S. Sex-specific differences in mitochondria biogenesis, morphology, respiratory function, and ROS homeostasis in young mouse heart and brain. *Physiol. Rep.* **2017**, *5*, e13125. [\[CrossRef\]](https://doi.org/10.14814/phy2.13125) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28325789)
- 93. Lloret, A.; Badía, M.C.; Mora, N.J.; Ortega, A.; Pallardó, F.V.; Alonso, M.D.; Atamna, H.; Viña, J. Gender and age-dependent differences in the mitochondrial apoptogenic pathway in Alzheimer's disease. *Free Radic. Biol. Med.* **2008**, *44*, 2019–2025. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2008.02.017)
- 94. Dkhil, M.A.; Al-Shaebi, E.M.; Lubbad, M.Y.; Al-Quraishy, S. Impact of sex differences in brain response to infection with Plasmodium berghei. *Parasitol. Res.* **2016**, *115*, 415–422. [\[CrossRef\]](https://doi.org/10.1007/s00436-015-4803-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26499384)
- 95. Ehrenbrink, G.; Hakenhaar, F.S.; Salomon, T.B.; Petrucci, A.P.; Sandri, M.R.; Benfato, M.S. Antioxidant enzymes activities and protein damage in rat brain of both sexes. *Exp. Gerontol.* **2006**, *41*, 368–371. [\[CrossRef\]](https://doi.org/10.1016/j.exger.2006.02.007)
- 96. Krolow, R.; Noschang, C.G.; Arcego, D.; Andreazza, A.C.; Peres, W.; Gonçalves, C.A.; Dalmaz, C. Consumption of a palatable diet by chronically stressed rats prevents effects on anxiety-like behavior but increases oxidative stress in a sex-specific manner. *Appetite* **2010**, *55*, 108–116. [\[CrossRef\]](https://doi.org/10.1016/j.appet.2010.03.013)
- 97. Mármol, F.; Rodríguez, C.A.; Sánchez, J.; Chamizo, V.D. Anti-oxidative effects produced by environmental enrichment in the hippocampus and cerebral cortex of male and female rats. *Brain Res.* **2015**, *1613*, 120–129. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2015.04.007)
- 98. Noschang, C.; Krolow, R.; Arcego, D.M.; Toniazzo, A.P.; Huffell, A.P.; Dalmaz, C. Neonatal handling affects learning, reversal learning and antioxidant enzymes activities in a sex-specific manner in rats. *Int. J. Dev. Neurosci.* **2012**, *30*, 285–291. [\[CrossRef\]](https://doi.org/10.1016/j.ijdevneu.2012.01.010)
- 99. Brocardo, P.S.; Boehme, F.; Patten, A.; Cox, A.; Gil-Mohapel, J.; Christie, B.R. Anxiety and depression-like behaviors are accompanied by an increase in oxidative stress in a rat model of fetal alcohol spectrum disorders: Protective effects of voluntary physical exercise. *Neuropharmacology* **2012**, *62*, 1607–1618. [\[CrossRef\]](https://doi.org/10.1016/j.neuropharm.2011.10.006)
- 100. Charradi, K.; Mahmoudi, M.; Bedhiafi, T.; Kadri, S.; Elkahoui, S.; Limam, F.; Aouani, E. Dietary supplementation of grape seed and skin flour mitigates brain oxidative damage induced by a high-fat diet in rat: Gender dependency. *Biomed. Pharmacother.* **2017**, *87*, 519–526. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2017.01.015)
- 101. Giménez-Llort, L.; García, Y.; Buccieri, K.; Revilla, S.; Suñol, C.; Cristofol, R.; Sanfeliu, C. Gender-specific neuroimmunoendocrine response to treadmill exercise in 3xTg-AD mice. *Int. J. Alzheimer Dis.* **2010**, *2010*, 128354. [\[CrossRef\]](https://doi.org/10.4061/2010/128354) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20981262)
- 102. Harish, G.; Venkateshappa, C.; Mahadevan, A.; Pruthi, N.; Srinivas Bharath, M.M.; Shankar, S.K. Effect of premortem and postmortem factors on the distribution and preservation of antioxidant activities in the cytosol and synaptosomes of human brains. *Biopreserv. Biobanking* **2012**, *10*, 253–265. [\[CrossRef\]](https://doi.org/10.1089/bio.2012.0001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24835064)
- 103. Engler-Chiurazzi, E.B.; Brown, C.M.; Povroznik, J.M.; Simpkins, J.W. Estrogens as neuroprotectants: Estrogenic actions in the context of cognitive aging and brain injury. *Prog. Neurobiol.* **2017**, *157*, 188–211. [\[CrossRef\]](https://doi.org/10.1016/j.pneurobio.2015.12.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26891883)
- 104. Liu, M.; Kelley, M.H.; Herson, P.S.; Hurn, P.D. Neuroprotection of sex steroids. *Minerva Endocrinol.* **2010**, *35*, 127–143. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20595940)
- 105. Siddiqui, A.N.; Siddiqui, N.; Khan, R.A.; Kalam, A.; Jabir, N.R.; Kamal, M.A.; Firoz, C.K.; Tabrez, S. Neuroprotective Role of Steroidal Sex Hormones: An Overview. *CNS Neurosci. Ther.* **2016**, *22*, 342–350. [\[CrossRef\]](https://doi.org/10.1111/cns.12538)
- 106. Spychala, M.S.; Honarpisheh, P.; McCullough, L.D. Sex differences in neuroinflammation and neuroprotection in ischemic stroke. *J. Neurosci. Res.* **2017**, *95*, 462–471. [\[CrossRef\]](https://doi.org/10.1002/jnr.23962)
- 107. Quillinan, N.; Deng, G.; Grewal, H.; Herson, P.S. Androgens and stroke: Good, bad or indifferent? *Exp. Neurol.* **2014**, *259*, 10–15. [\[CrossRef\]](https://doi.org/10.1016/j.expneurol.2014.02.004)
- 108. Reddy, D.S.; Bakshi, K. Neurosteroids: Biosynthesis, molecular mechanisms, and neurophysiological functions in the human brain. *Horm. Signal. Biol. Med.* **2020**, 69–82. [\[CrossRef\]](https://doi.org/10.1016/b978-0-12-813814-4.00004-3)
- 109. Yilmaz, C.; Karali, K.; Fodelianaki, G.; Gravanis, A.; Chavakis, T.; Charalampopoulos, I.; Alexaki, V.I. Neurosteroids as regulators of neuroinflammation. *Front. Neuroendocrinol.* **2019**, *55*, 100788. [\[CrossRef\]](https://doi.org/10.1016/j.yfrne.2019.100788)
- 110. Illan-Cabeza, N.A.; Garcia-Garcia, A.R.; Martinez-Martos, J.M.; Ramirez-Exposito, M.J.; Pena-Ruiz, T.; Moreno-Carretero, M.N. A potential antitumor agent, (6-amino-1-methyl-5-nitrosouracilato-N3)-triphenylphosphinegold(I): Structural studies and in vivo biological effects against experimental glioma. *Eur. J. Med. Chem.* **2013**, *64*, 260–272. [\[CrossRef\]](https://doi.org/10.1016/j.ejmech.2013.03.067)
- 111. Ramirez-Exposito, M.J.; Martinez-Martos, J.M. The Delicate Equilibrium between Oxidants and Antioxidants in Brain Glioma. *Curr. Neuropharmacol.* **2019**, *17*, 342–351. [\[CrossRef\]](https://doi.org/10.2174/1570159X16666180302120925)
- 112. Martínez-Martos, J.M.; Mayas, M.D.; Carrera, P.; Arias de Saavedra, J.M.; Sánchez-Agesta, R.; Marcela Arrazola, M.; Ramírez-Expósito, M.J. Phenolic compounds oleuropein and hydroxytyrosol exert differential effects on glioma development via antioxidant defense systems. *J. Funct. Food* **2014**, *11*, 221–234. [\[CrossRef\]](https://doi.org/10.1016/j.jff.2014.09.006)
- 113. Ramírez-Expósito, M.J.; Carrera-González, M.P.; Mayas, M.D.; Martínez-Martos, J.M. Gender differences in the antioxidant response of oral administration of hydroxytyrosol and oleuropein against N-ethyl-N-nitrosourea (ENU)-induced glioma. *Food Res. Int.* **2021**, *140*, 110023. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2020.110023)
- 114. Ramírez-Expósito, M.J.; Mayas, M.D.; Carrera-González, M.P.; Martínez-Martos, J.M. Gender Differences in the Antioxidant Response to Oxidative Stress in Experimental Brain Tumors. *Curr. Cancer Drug Targets* **2019**, *19*, 641–654. [\[CrossRef\]](https://doi.org/10.2174/1568009618666181018162549)
- 115. Ferlay, J.; Soerjomataram, I.; Dikshit, R.; Eser, S.; Mathers, C.; Rebelo, M.; Parkin, D.M.; Forman, D.; Bray, F. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int. J. Cancer* **2015**, *136*, E359–E386. [\[CrossRef\]](https://doi.org/10.1002/ijc.29210)
- 116. Villanueva, A.; Llovet, J.M. Liver cancer in 2013: Mutational landscape of HCC-the end of the beginning. *Nat. Rev. Clin. Oncol.* **2014**, *11*, 73–74. [\[CrossRef\]](https://doi.org/10.1038/nrclinonc.2013.243)
- 117. El-Serag, H.B. Hepatocellular carcinoma. *N. Engl. J. Med.* **2011**, *365*, 1118–1127. [\[CrossRef\]](https://doi.org/10.1056/NEJMra1001683)
- 118. Guy, J.; Peters, M.G. Liver disease in women: The influence of gender on epidemiology, natural history, and patient outcomes. *Gastroenterol. Hepatol.* **2013**, *9*, 633–639.
- 119. Vesselinovitch, S.D.; Mihailovich, N.; Wogan, G.N.; Lombard, L.S.; Rao, K.V.; Sugamori, K.S.; Brenneman, D.; Sanchez, O.; Doll, M.A.; Hein, D.W.; et al. Reduced 4-aminobiphenyl-induced liver tumorigenicity but not DNA damage in arylamine N-acetyltransferase null mice. *Cancer Lett.* **2012**, *318*, 206–213. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2011.12.022)
- 120. Gramenzi, A.; Caputo, F.; Biselli, M.; Kuria, F.; Loggi, E.; Andreone, P.; Bernardi, M. Review article: Alcoholic liver disease— Pathophysiological aspects and risk factors. *Aliment. Pharmacol. Ther.* **2006**, *24*, 1151–1161. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2036.2006.03110.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17014574)
- 121. Harada, S. Classification of alcohol metabolizing enzymes and polymorphisms–specificity in Japanese. *Nihon Arukoru Yakubutsu Igakkai Zasshi* **2001**, *36*, 85–106. (In Japanese)
- 122. Brandon-Warner, E.; Walling, T.L.; Schrum, L.W.; McKillop, I.H. Chronic ethanol feeding accelerates hepatocellular carcinoma progression in a sex-dependent manner in a mouse model of hepatocarcinogenesis. *Alcohol. Clin. Exp. Res.* **2012**, *36*, 641–653. [\[CrossRef\]](https://doi.org/10.1111/j.1530-0277.2011.01660.x)
- 123. Yoshino, K.; Komura, S.; Watanabe, I.; Nakagawa, Y.; Yagi, K. Effect of estrogens on serum and liver lipid peroxide levels in mice. *J. Clin. Biochem. Nutr.* **1987**, *3*, 233–239. [\[CrossRef\]](https://doi.org/10.3164/jcbn.3.233)
- 124. Lacort, M.; Leal, A.M.; Liza, M.; Martín, C.; Martínez, R.; Ruiz-Larrea, M.B. Protective effect of estrogens and catecholestrogens against peroxidative membrane damage in vitro. *Lipids* **1995**, *30*, 141–146. [\[CrossRef\]](https://doi.org/10.1007/BF02538267)
- 125. Omoya, T.; Shimizu, I.; Zhou, Y.; Okamura, Y.; Inoue, H.; Lu, G.; Itonaga, M.; Honda, H.; Nomura, M.; Ito, S. Effects of idoxifene and estradiol on NF-kappaB activation in cultured rat hepatocytes undergoing oxidative stress. *Liver* **2001**, *21*, 183–191. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0676.2001.021003183.x)
- 126. Inoue, H.; Shimizu, I.; Lu, G.; Itonaga, M.; Cui, X.; Okamura, Y.; Shono, M.; Honda, H.; Inoue, S.; Muramatsu, M.; et al. Idoxifene and estradiol enhance antiapoptotic activity through estrogen receptor-beta in cultured rat hepatocytes. *Dig. Dis. Sci.* **2003**, *48*, 570–580. [\[CrossRef\]](https://doi.org/10.1023/A:1022553119715)
- 127. Tanaka, Y.; Mukaide, M.; Orito, E.; Yuen, M.F.; Ito, K.; Kurbanov, F.; Sugauchi, F.; Asahina, Y.; Izumi, N.; Kato, M.; et al. Specific mutations in enhancer II/core promoter of hepatitis B virus subgenotypes C1/C2 increase the risk of hepatocellular carcinoma. *J. Hepatol.* **2006**, *45*, 646–653. [\[CrossRef\]](https://doi.org/10.1016/j.jhep.2006.06.018) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16935384)
- 128. Slocum, S.L.; Kensler, T.W. Nrf2: Control of sensitivity to carcinogens. *Arch. Toxicol.* **2011**, *85*, 273–284. [\[CrossRef\]](https://doi.org/10.1007/s00204-011-0675-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21369766)
- 129. Kitamura, Y.; Umemura, T.; Kanki, K.; Kodama, Y.; Kitamoto, S.; Saito, K.; Itoh, K.; Yamamoto, M.; Masegi, T.; Nishikawa, A.; et al. Increased susceptibility to hepatocarcinogenicity of Nrf2-deficient mice exposed to 2-amino-3-methylimidazo [4,5-f]quinoline. *Cancer Sci.* **2007**, *98*, 19–24. [\[CrossRef\]](https://doi.org/10.1111/j.1349-7006.2006.00352.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17083568)
- 130. Thimmulappa, R.K.; Lee, H.; Rangasamy, T.; Reddy, S.P.; Yamamoto, M.; Kensler, T.W.; Biswal, S. Nrf2 is a critical regulator of the innate immune response and survival during experimental sepsis. *J. Clin. Investig.* **2006**, *116*, 984–995. [\[CrossRef\]](https://doi.org/10.1172/JCI25790)
- 131. Wruck, C.J.; Streetz, K.; Pavic, G.; Götz, M.E.; Tohidnezhad, M.; Brandenburg, L.O.; Varoga, D.; Eickelberg, O.; Herdegen, T.; Trautwein, C.; et al. Nrf2 induces interleukin-6 (IL-6) expression via an antioxidant response element within the IL-6 promoter. *J. Biol. Chem.* **2011**, *286*, 4493–4499. [\[CrossRef\]](https://doi.org/10.1074/jbc.M110.162008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21127061)
- 132. Hanna, D.; Riedmaier, A.E.; Sugamori, K.S.; Grant, D.M. Influence of sex and developmental stage on acute hepatotoxic and inflammatory responses to liver procarcinogens in the mouse. *Toxicology* **2016**, *373*, 30–40. [\[CrossRef\]](https://doi.org/10.1016/j.tox.2016.10.006)
- 133. Shupe, T.; Sell, S. Low hepatic glutathione S-transferase and increased hepatic DNA adduction contribute to increased tumorigenicity of aflatoxin B1 in newborn and partially hepatectomized mice. *Toxicol. Lett.* **2004**, *148*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.toxlet.2003.11.008)
- 134. Eaton, D.L.; Gallagher, E.P. Mechanisms of aflatoxin carcinogenesis. *Annu. Rev. Pharmacol. Toxicol.* **1994**, *34*, 135–172. [\[CrossRef\]](https://doi.org/10.1146/annurev.pa.34.040194.001031) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8042848)
- 135. Egner, P.A.; Groopman, J.D.; Wang, J.S.; Kensler, T.W.; Friesen, M.D. Quantification of aflatoxin-B1-N7-Guanine in human urine by high-performance liquid chromatography and isotope dilution tandem mass spectrometry. *Chem. Res. Toxicol.* **2006**, *19*, 1191–1195. [\[CrossRef\]](https://doi.org/10.1021/tx060108d)
- 136. Crawford, D.R.; Ilic, Z.; Guest, I.; Milne, G.L.; Hayes, J.D.; Sell, S. Characterization of liver injury, oval cell proliferation and cholangiocarcinogenesis in glutathione S-transferase A3 knockout mice. *Carcinogenesis* **2017**, *38*, 717–727. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgx048) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28535182)
- 137. Fu, D.; Hornick, C.A. Modulation of lipid metabolism at rat hepatic subcellular sites by female sex hormones. *Biochim. Biophys. Acta* **1995**, *1254*, 267–273. [\[CrossRef\]](https://doi.org/10.1016/0005-2760(94)00187-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7857966)
- 138. Chen, J.; Li, Y.; Lavigne, J.A.; Trush, M.A.; Yager, J.D. Increased mitochondrial superoxide production in rat liver mitochondria, rat hepatocytes, and HepG2 cells following ethinyl estradiol treatment. *Toxicol. Sci.* **1999**, *52*, 224–235. [\[CrossRef\]](https://doi.org/10.1093/toxsci/51.2.224)
- 139. Gerber, M.; Astre, C.; Segala, C.; Saintot, M.; Scali, J.; Simony Lafontaine, J.; Grenier, J.; Pujol, H. Tumor progression and oxidant-antioxidant status. *Cancer Lett.* **1997**, *114*, 211–214. [\[CrossRef\]](https://doi.org/10.1016/S0304-3835(97)04665-X)
- 140. Sverko, V.; Sobocanec, S.; Balog, T.; Marotti, T. Age and gender differences in antioxidant enzyme activity: Potential relationship to liver carcinogenesis in male mice. *Biogerontology* **2004**, *5*, 235–242. [\[CrossRef\]](https://doi.org/10.1023/B:BGEN.0000038024.58911.6e)
- 141. Coto-Montes, A.; Boga, J.A.; Tomas-Zapico, C.; Rodriguez Colunga, M.J.; Martinez-Fraga, J.; Tolivia-Cadrecha, D.; Manendez, G.; Herdeband, R.; Tolivia, D. Physiological oxidative stress model: Syrian hamster harderian gland-sex differences in antioxidant enzymes. *Free Radic. Biol. Med.* **2001**, *30*, 785–792. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(01)00468-3)
- 142. Oberley, L.W.; Oberley, T.D. Role of antioxidant enzymes in cell immortalization and transformation. *Mol. Cell. Biochem.* **1988**, *84*, 147–153. [\[CrossRef\]](https://doi.org/10.1007/BF00421049)
- 143. Janssen, A.M.; Bosman, C.B.; Sier, C.F.; Griffioen, G.; Kubben, F.J.; Lamers, C.B.; van Krieken, J.H.; van de Velde, C.J.; Verspaget, H.W. Superoxide dismutase in relation to the overall survival of colorectal cancer patients. *Br. J. Cancer* **1998**, *78*, 1051–1057. [\[CrossRef\]](https://doi.org/10.1038/bjc.1998.626)
- 144. Manna, S.K.; Zhang, H.J.; Yan, T.; Oberley, L.W.; Aggarwal, B.B. Overexpression of manganese superoxide dismutase suppresses tumor necrosis factor-induced apoptosis and activation of nuclear transcription factor-jB and activated protein-1. *J. Biol. Chem.* **1998**, *273*, 13245–13254. [\[CrossRef\]](https://doi.org/10.1074/jbc.273.21.13245)
- 145. Szatrowski, T.P.; Nathan, C.F. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res.* **1991**, *51*, 794–798. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1846317)
- 146. Gardner, R.; Salvador, A.; Moradas-Ferreira, P. Why does SOD overexpression sometimes enhance, sometimes decrease, hydrogen peroxide production? A minimalist explanation. *Free Radic. Biol. Med.* **2022**, *32*, 1352–1357. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(02)00861-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12057773)
- 147. Pigeolet, E.; Corbisier, P.; Houbion, A.; Lambert, D.; Michiels, C.; Raes, M.; Zachary, M.D.; Remacle, J. Glutathione peroxidase, superoxide dismutase and catalase inactivation by peroxides and oxygen-derived free radicals. *Mech. Ageing Dev.* **1990**, *51*, 283–297. [\[CrossRef\]](https://doi.org/10.1016/0047-6374(90)90078-T) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2308398)
- 148. Gonzales, M.J. Lipid peroxidation and tumor growth: An inverse relationship. *Med. Hypotheses* **1992**, *38*, 106–110. [\[CrossRef\]](https://doi.org/10.1016/0306-9877(92)90081-M) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1528153)
- 149. Wang, S.; Sugamori, K.S.; Tung, A.; McPherson, J.P.; Grant, D.M. N-hydroxylation of 4-aminobiphenyl by CYP2E1 produces oxidative stress in a mouse model of chemically induced liver cancer. *Toxicol. Sci.* **2015**, *144*, 393–405. [\[CrossRef\]](https://doi.org/10.1093/toxsci/kfv006)
- 150. Calle, E.E.; Rodriguez, C.; Walker-Thurmond, K.; Thun, M.J. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N. Engl. J. Med.* **2003**, *348*, 1625–1638. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa021423) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12711737)
- 151. Bhaskaran, K.; Douglas, I.; Forbes, H.; dos-Santos-Silva, I.; Leon, D.A.; Smeeth, L. Body-mass index and risk of 22 specific cancers: A population-based cohort study of 5.24 million UK adults. *Lancet* **2014**, *384*, 755–765. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(14)60892-8)
- 152. Świątkiewicz, I.; Wróblewski, M.; Nuszkiewicz, J.; Sutkowy, P.; Wróblewska, J.; Woźniak, A. The Role of Oxidative Stress Enhanced by Adiposity in Cardiometabolic Diseases. *Int. J. Mol. Sci.* **2023**, *24*, 6382. [\[CrossRef\]](https://doi.org/10.3390/ijms24076382)
- 153. Ibrahim, M.M. Subcutaneous and visceral adipose tissue: Structural and functional differences. *Obes. Rev.* **2010**, *11*, 11–18. [\[CrossRef\]](https://doi.org/10.1111/j.1467-789X.2009.00623.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19656312)
- 154. Setiawan, V.W.; Lim, U.; Lipworth, L.; Lu, S.C.; Shepherd, J.; Ernst, T.; Wilkens, L.R.; Henderson, B.E.; Le Marchand, L. Sex and ethnic differences in the association of obesity with risk of hepatocellular carcinoma. *Clin. Gastroenterol. Hepatol.* **2016**, *14*, 309–316. [\[CrossRef\]](https://doi.org/10.1016/j.cgh.2015.09.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26404865)
- 155. Chang, Y.; Jung, H.S.; Cho, J.; Zhang, Y.; Yun, K.E.; Lazo, M.; Pastor-Barriuso, R.; Ahn, J.; Kim, C.W.; Rampal, S.; et al. Metabolically healthy obesity and the development of nonalcoholic fatty liver disease. *Am. J. Gastroenterol.* **2016**, *111*, 1133–1140. [\[CrossRef\]](https://doi.org/10.1038/ajg.2016.178)
- 156. Freedland, E.S. Roles of critical visceral adipose tissue threshold in metabolic syndrome: Implications for controlling dietary carbohydrates: A review. *Nutr. Metab.* **2004**, *1*, 12. [\[CrossRef\]](https://doi.org/10.1186/1743-7075-1-12)
- 157. Björntorp, P. Endocrine abnormalities in obesity. *Metabolism.* **1995**, *44* (Suppl. 3), 21–23. [\[CrossRef\]](https://doi.org/10.1016/0026-0495(95)90315-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7674912)
- 158. He, J.; Gu, D.; Wu, X.; Reynolds, K.; Duan, X.; Yao, C.; Wang, J.; Chen, C.S.; Chen, J.; Wildman, R.P.; et al. Major causes of death among men and women in China. *N. Engl. J. Med.* **2005**, *353*, 1124–1134. [\[CrossRef\]](https://doi.org/10.1056/NEJMsa050467) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16162883)
- 159. Riley, G.F.; Potosky, A.L.; Lubitz, J.D.; Kessler, L.G. Medicare payments from diagnosis to death for elderly cancer patients by stage at diagnosis. *Med. Care* **1995**, *33*, 828–841. [\[CrossRef\]](https://doi.org/10.1097/00005650-199508000-00007)
- 160. Ohnishi, S.; Murata, M.; Kawanishi, S. DNA damage induced by hypochlorite and hypobromite with reference to inflammationassociated carcinogenesis. *Cancer Lett.* **2002**, *178*, 37–42. [\[CrossRef\]](https://doi.org/10.1016/S0304-3835(01)00812-6)
- 161. Kiyohara, C.; Yoshimasu, K.; Takayama, K.; Nakanishi, Y. NQO1, MPO, and the risk of lung cancer: A HuGE review. *Genet. Med.* **2005**, *7*, 463–478. [\[CrossRef\]](https://doi.org/10.1097/01.gim.0000177530.55043.c1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16170238)
- 162. Yang, M.; Choi, Y.; Hwangbo, B.; Lee, J.S. Combined effects of genetic polymorphisms in six selected genes on lung cancer susceptibility. *Lung Cancer* **2007**, *57*, 135–142. [\[CrossRef\]](https://doi.org/10.1016/j.lungcan.2007.03.005)
- 163. Yang, J.; Ambrosone, C.B.; Hong, C.C.; Ahn, J.; Rodriguez, C.; Thun, M.J.; Calle, E.E. Relationships between polymorphisms in NOS3 and MPO genes, cigarette smoking and risk of post-menopausal breast cancer. *Carcinogenesis* **2007**, *28*, 1247–1253. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgm016)
- 164. Steenport, M.; Eom, H.; Uezu, M.; Schneller, J.; Gupta, R.; Mustafa, Y.; Villanueva, R.; Straus, E.W.; Raffaniello, R.D. Association of polymorphisms in myeloperoxidase and catalase genes with precancerous changes in the gastric mucosa of patients at inner-city hospitals in New York. *Oncol. Rep.* **2007**, *18*, 235–240. [\[CrossRef\]](https://doi.org/10.3892/or.18.1.235) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17549373)
- 165. Cascorbi, I.; Henning, S.; Brockmöller, J.; Gephart, J.; Meisel, C.; Müller, J.M.; Loddenkemper, R.; Roots, I. Substantially reduced risk of cancer of the aerodigestive tract in subjects with variant--463A of the myeloperoxidase gene. *Cancer Res.* **2000**, *60*, 644–649.
- 166. Li, Y.; Qin, Y.; Wang, M.L.; Zhu, H.F.; Huang, X.E. The myeloperoxidase-463 G>A polymorphism influences risk of colorectal cancer in southern China: A case-control study. *Asian Pac. J. Cancer Prev.* **2011**, *12*, 1789–1793.
- 167. Otero Regino, W.; Velasco, H.; Sandoval, H. The protective role of bilirubin in human beings. *Rev. Colomb. Gastroenterol.* **2009**, *24*, 293–301.
- 168. Wagner, K.H.; Wallner, M.; Molzer, C.; Gazzin, S.; Bulmer, A.C.; Tiribelli, C.; Vitek, L. Looking to the horizon: The role of bilirubin in the development and prevention of age-related chronic diseases. *Clin. Sci.* **2015**, *129*, 1–25. [\[CrossRef\]](https://doi.org/10.1042/CS20140566) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25881719)
- 169. Sedlak, T.W.; Saleh, M.; Higginson, D.S.; Paul, B.D.; Juluri, K.R.; Snyder, S.H. Bilirubin and glutathione have complementary antioxidant and cytoprotective roles. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 5171–5176. [\[CrossRef\]](https://doi.org/10.1073/pnas.0813132106)
- 170. Rodrigues, C.M.; Sola, S.; Brito, M.A.; Brites, D.; Moura, J.J. Bilirubin directly disrupts membrane lipid polarity and fluidity, protein order, and redox status in rat mitochondria. *J. Hepatol.* **2002**, *36*, 335–341. [\[CrossRef\]](https://doi.org/10.1016/S0168-8278(01)00279-3)
- 171. Hansen, T.W.; Mathiesen, S.B.; Walaas, S.I. Bilirubin has widespread inhibitory effects on protein phosphorylation. *Pediatr. Res.* **1996**, *39*, 1072–1077. [\[CrossRef\]](https://doi.org/10.1203/00006450-199606000-00023) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8725272)
- 172. Fevery, J. Bilirubin in clinical practice: A review. *Liver Int.* **2008**, *28*, 592–605. [\[CrossRef\]](https://doi.org/10.1111/j.1478-3231.2008.01716.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18433389)
- 173. Bulmer, A.C.; Ried, K.; Coombes, J.S.; Blanchfield, J.T.; Toth, I.; Wagner, K.H. The anti-mutagenic and antioxidant effects of bile pigments in the Ames Salmonella test. *Mutat. Res.* **2007**, *629*, 122–132. [\[CrossRef\]](https://doi.org/10.1016/j.mrgentox.2007.01.008)
- 174. Molzer, C.; Huber, H.; Diem, K.; Wallner, M.; Bulmer, A.C.; Wagner, K.H. Extracellular and intracellular anti-mutagenic effects of bile pigments in the Salmonella typhimurium reverse mutation assay. *Toxicol. Int. J. Public Assoc. BIBRA* **2013**, *27*, 433–437. [\[CrossRef\]](https://doi.org/10.1016/j.tiv.2012.08.004)
- 175. Molzer, C.; Huber, H.; Steyrer, A.; Ziesel, G.; Ertl, A.; Plavotic, A.; Wallner, M.; Bulmer, A.C.; Wagner, K.H. In vitro antioxidant capacity and antigenotoxic properties of protoporphyrin and structurally related tetrapyrroles. *Free Radic. Res.* **2012**, *46*, 1369–1377. [\[CrossRef\]](https://doi.org/10.3109/10715762.2012.715371) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22861140)
- 176. Molzer, C.; Huber, H.; Steyrer, A.; Ziesel, G.V.; Wallner, M.; Hong, H.T.; Blanchfield, J.T.; Bulmer, A.C.; Wagner, K.H. Bilirubin and related tetrapyrroles inhibit food-borne mutagenesis: A mechanism for antigenotoxic action against a model epoxide. *J. Nat. Prod.* **2013**, *76*, 1958–1965. [\[CrossRef\]](https://doi.org/10.1021/np4005807)
- 177. Seyed Khoei, N.; Jenab, M.; Murphy, N.; Banbury, B.L.; Carreras-Torres, R.; Viallon, V.; Kühn, T.; Bueno-de-Mesquita, B.; Aleksandrova, K.; Cross, A.J.; et al. Circulating bilirubin levels and risk of colorectal cancer: Serological and Mendelian randomization analyses. *BMC Med.* **2020**, *18*, 229. [\[CrossRef\]](https://doi.org/10.1186/s12916-020-01703-w)
- 178. Seyed Khoei, N.; Anton, G.; Peters, A.; Freisling, H.; Wagner, K.H. The Association between Serum Bilirubin Levels and Colorectal Cancer Risk: Results from the Prospective Cooperative Health Research in the Region of Augsburg (KORA) Study in Germany. *Antioxidants* **2020**, *9*, 908. [\[CrossRef\]](https://doi.org/10.3390/antiox9100908)
- 179. Ollinger, R.; Kogler, P.; Troppmair, J.; Hermann, M.; Wurm, M.; Drasche, A.; Konigsrainer, I.; Amberger, A.; Weiss, H.; Ofner, D.; et al. Bilirubin inhibits tumor cell growth via activation of ERK. *Cell Cycle* **2007**, *6*, 3078–3085. [\[CrossRef\]](https://doi.org/10.4161/cc.6.24.5022)
- 180. Keshavan, P.; Schwemberger, S.J.; Smith, D.L.; Babcock, G.F.; Zucker, S.D. Unconjugated bilirubin induces apoptosis in colon cancer cells by triggering mitochondrial depolarization. *Int. J. Cancer* **2004**, *112*, 433–445. [\[CrossRef\]](https://doi.org/10.1002/ijc.20418)
- 181. Grant, D.J.; Bell, D.A. Bilirubin UDP-glucuronosyltransferase 1A1 gene polymorphisms: Susceptibility to oxidative damage and cancer? *Mol. Carcinogen.* **2000**, *29*, 198–204. [\[CrossRef\]](https://doi.org/10.1002/1098-2744(200012)29:4<198::AID-MC1001>3.0.CO;2-K)
- 182. Zhang, T.; Joubert, P.; Ansari-Pour, N.; Zhao, W.; Hoang, P.H.; Lokanga, R.; Moye, A.L.; Rosenbaum, J.; Gonzalez-Perez, A.; Martínez-Jiménez, F.; et al. Genomic and Evolutionary Classification of Lung Cancer in Never Smokers. *Nat. Genet.* **2021**, *53*, 1348–1359. [\[CrossRef\]](https://doi.org/10.1038/s41588-021-00920-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34493867)
- 183. Adib, E.; Nassar, A.H.; Abou Alaiwi, S.; Groha, S.; Akl, E.W.; Sholl, L.M.; Michael, K.S.; Awad, M.M.; Jänne, P.A.; Gusev, A.; et al. Variation in Targetable Genomic Alterations in Non-Small Cell Lung Cancer by Genetic Ancestry, Sex, Smoking History, and Histology. *Genome Med.* **2022**, *14*, 39. [\[CrossRef\]](https://doi.org/10.1186/s13073-022-01041-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35428358)
- 184. Gasperino, J.; Rom, W.N. Gender and lung cancer. *Clin Lung Cancer* **2004**, *5*, 353–359. [\[CrossRef\]](https://doi.org/10.3816/CLC.2004.n.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15217534)
- 185. Thomas, L.; Doyle, L.A.; Edelman, M.J. Lung cancer in women: Emerging differences in epidemiology, biology, and therapy. *Chest* **2005**, *128*, 370–381. [\[CrossRef\]](https://doi.org/10.1378/chest.128.1.370) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16002959)
- 186. Kiyohara, C.; Ohno, Y. Sex differences in lung cancer susceptibility: A review. *Gend Med.* **2010**, *7*, 381–401. [\[CrossRef\]](https://doi.org/10.1016/j.genm.2010.10.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21056866)
- 187. Henschke, C.I.; Yip, R.; Miettinen, O.S. Women's susceptibility to tobacco carcinogens and survival after diagnosis of lung cancer. *J. Am. Med. Assoc.* **2006**, *296*, 180–184.
- 188. Parajuli, R.; Bjerkaas, E.; Tverdal, A.; Selmer, R.; Le Marchand, L.; Weiderpass, E.; Gram, I.T. The increased risk of colon cancer due to cigarette smoking may be greater in women than men. *Cancer Epidemiol. Biomarkers Prev.* **2013**, *22*, 862–871. [\[CrossRef\]](https://doi.org/10.1158/1055-9965.EPI-12-1351)
- 189. Forder, A.; Zhuang, R.; Souza, V.G.P.; Brockley, L.J.; Pewarchuk, M.E.; Telkar, N.; Stewart, G.L.; Benard, K.; Marshall, E.A.; Reis, P.P.; et al. Mechanisms Contributing to the Comorbidity of COPD and Lung Cancer. *Int. J. Mol. Sci.* **2023**, *24*, 2859. [\[CrossRef\]](https://doi.org/10.3390/ijms24032859)
- 190. Dewhirst, M.W.; Cao, Y.; Moeller, B. Cycling Hypoxia and Free Radicals Regulate Angiogenesis and Radiotherapy Response. *Nat. Rev. Cancer* **2008**, *8*, 425–437. [\[CrossRef\]](https://doi.org/10.1038/nrc2397)
- 191. Covey, T.M.; Edes, K.; Coombs, G.S.; Virshup, D.M.; Fitzpatrick, F.A. Alkylation of the Tumor Suppressor PTEN Activates Akt and β-Catenin Signaling: A Mechanism Linking Inflammation and Oxidative Stress with Cancer. *PLoS ONE* **2010**, *5*, e13545. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0013545) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20975834)
- 192. Cai, B.; Liu, M.; Li, J.; Xu, D.; Li, J. Cigarette Smoke Extract Amplifies NADPH Oxidase-Dependent ROS Production to Inactivate PTEN by Oxidation in BEAS-2B Cells. *Food Chem. Toxicol. Int. J. Publ. Br. Ind. Biol. Res. Assoc.* **2021**, *150*, 112050. [\[CrossRef\]](https://doi.org/10.1016/j.fct.2021.112050)
- 193. Shukla, S.D.; Shastri, M.D.; Jha, N.K.; Gupta, G.; Chellappan, D.K.; Bagade, T.; Dua, K. Female Gender as a Risk Factor for Developing COPD. *EXCLI J.* **2021**, *20*, 1290–1293.
- 194. Ben-Zaken Cohen, S.; Paré, P.D.; Man, S.F.P.; Sin, D.D. The Growing Burden of Chronic Obstructive Pulmonary Disease and Lung Cancer in Women: Examining Sex Differences in Cigarette Smoke Metabolism. *Am. J. Respir. Crit. Care Med.* **2007**, *176*, 113–120. [\[CrossRef\]](https://doi.org/10.1164/rccm.200611-1655PP) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17413125)
- 195. Meireles, S.I.; Esteves, G.H.; Hirata, R.J.; Peri, S.; Devarajan, K.; Slifker, M.; Mosier, S.L.; Peng, J.; Vadhanam, M.V.; Hurst, H.E.; et al. Early Changes in Gene Expression Induced by Tobacco Smoke: Evidence for the Importance of Estrogen within Lung Tissue. *Cancer Prev. Res.* **2010**, *3*, 707–717. [\[CrossRef\]](https://doi.org/10.1158/1940-6207.CAPR-09-0162)
- 196. Mollerup, S.; Ryberg, D.; Hewer, A.; Phillips, D.H.; Haugen, A. Sex Differences in Lung CYP1A1 Expression and DNA Adduct Levels among Lung Cancer Patients. *Cancer Res.* **1999**, *59*, 3317–3320.
- 197. Van Winkle, L.S.; Gunderson, A.D.; Shimizu, J.A.; Baker, G.L.; Brown, C.D. Gender Differences in Naphthalene Metabolism and Naphthalene-Induced Acute Lung Injury. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2002**, *282*, L1122–L1134. [\[CrossRef\]](https://doi.org/10.1152/ajplung.00309.2001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11943679)
- 198. Soriano, J.B.; Kendrick, P.J.; Paulson, K.R.; Gupta, V.; Abrams, E.M.; Adedoyin, R.A.; Adhikari, T.B.; Advani, S.M.; Agrawal, A.; Ahmadian, E.; et al. Prevalence and Attributable Health Burden of Chronic Respiratory Diseases, 1990-2017: A Systematic Analysis for the Global Burden of Disease Study 2017. *Lancet Respir. Med.* **2020**, *8*, 585–596. [\[CrossRef\]](https://doi.org/10.1016/S2213-2600(20)30105-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32526187)
- 199. Ramírez-Venegas, A.; Sansores, R.H.; Pérez-Padilla, R.; Regalado, J.; Velázquez, A.; Sánchez, C.; Mayar, M.E. Survival of Patients with Chronic Obstructive Pulmonary Disease Due to Biomass Smoke and Tobacco. *Am. J. Respir. Crit. Care Med.* **2006**, *173*, 393–397. [\[CrossRef\]](https://doi.org/10.1164/rccm.200504-568OC)
- 200. Armstrong, B.G.; Gibbs, G. Exposure-response relationship between lung cancer and polycyclic aromatic hydrocarbons (PAHs). *Occup. Environ. Med.* **2009**, *66*, 740–746. [\[CrossRef\]](https://doi.org/10.1136/oem.2008.043711)
- 201. Veglia, F.; Matullo, G.; Vineis, P. Bulky DNA adducts and risk of cancer: A meta-analysis. *Cancer Epidemiol. Biomarkers Prev.* **2003**, *12*, 157–160. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12582026)
- 202. Bostrom, C.E.; Gerde, P.; Hanberg, A.; Jernstrom, B.; Johansson, C.; Kyrklund, T.; Rannug, A.; Tornqvist, M.; Victorin, K.; Westerholm, R. Cancer risk assessment, indicators, and guidelines for polycyclic aromatic hydrocarbons in the ambient air. *Environ. Health Perspect.* **2002**, *110* (Suppl. 3), 451–488. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12060843)
- 203. Guo, H.; Huang, K.; Zhang, X.; Zhang, W.; Guan, L.; Kuang, D.; Deng, Q.; Deng, H.; Zhang, X.; He, M.; et al. Women are more susceptible than men to oxidative stress and chromosome damage caused by polycyclic aromatic hydrocarbons exposure. *Environ. Mol. Mutagen.* **2014**, *55*, 472–481. [\[CrossRef\]](https://doi.org/10.1002/em.21866)
- 204. Siegel, R.; Naishadham, D.; Jemal, A. Cancer statistics. *CA Cancer J. Clin.* **2013**, *63*, 11–30. [\[CrossRef\]](https://doi.org/10.3322/caac.21166)
- 205. Balch, C.M.; Gershenwald, J.E.; Soong, S.J.; Thompson, J.F.; Atkins, M.B.; Byrd, D.R.; Buzaid, A.C.; Cochran, A.J.; Coit, D.G.; Ding, S.; et al. Final version of 2009 AJCC melanoma staging and classification. *J. Clin. Oncol.* **2009**, *27*, 6199–6206. [\[CrossRef\]](https://doi.org/10.1200/JCO.2009.23.4799)
- 206. De Vries, E.; Nijsten, T.E.; Visser, O.; Bastiaannet, E.; Van Hattem, S.; Janssen-Heijnen, M.L.; Coebergh, J.W. Superior survival of females among 10,538 Dutch melanoma patients is independent of Breslow thickness, histologic type and tumor site. *Ann. Oncol.* **2008**, *19*, 583–589. [\[CrossRef\]](https://doi.org/10.1093/annonc/mdm498)
- 207. Lasithiotakis, K.; Leiter, U.; Meier, F.; Eigentler, T.; Metzler, G.; Moehrle, M.; Breuninger, H.; Garbe, C. Age and gender are significant independent predictors of survival in primary cutaneous melanoma. *Cancer* **2008**, *112*, 1795–1804. [\[CrossRef\]](https://doi.org/10.1002/cncr.23359)
- 208. Richardson, B.; Price, A.; Wagner, M.; Williams, V.; Lorigan, P.; Browne, S.; Miller, J.G.; Mac Neil, S. Investigation of female survival benefit in metastatic melanoma. *Br. J. Cancer* **1999**, *80*, 2025–2033. [\[CrossRef\]](https://doi.org/10.1038/sj.bjc.6690637)
- 209. Kemeny, M.M.; Busch, E.; Stewart, A.K.; Menck, H.R. Superior survival of young women with malignant melanoma. *Am. J. Surg.* **1998**, *175*, 437–444; discussion 444–445. [\[CrossRef\]](https://doi.org/10.1016/S0002-9610(98)00070-1)
- 210. Daryanani, D.; Plukker, J.T.; De Jong, M.A.; Haaxma-Reiche, H.; Nap, R.; Kuiper, H.; Hoekstra, H.J. Increased incidence of brain metastases in cutaneous head and neck melanoma. *Melanoma Res.* **2005**, *15*, 119–124. [\[CrossRef\]](https://doi.org/10.1097/00008390-200504000-00006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15846145)
- 211. Scoggins, C.R.; Ross, M.I.; Reintgen, D.S.; Noyes, R.D.; Goydos, J.S.; Beitsch, P.D.; Urist, M.M.; Ariyan, S.; Sussman, J.J.; Edwards, M.J.; et al. Gender-related differences in outcome for melanoma patients. *Ann. Surg.* **2006**, *243*, 693–698; discussion 698–700. [\[CrossRef\]](https://doi.org/10.1097/01.sla.0000216771.81362.6b)
- 212. Joosse, A.; Collette, S.; Suciu, S.; Nijsten, T.; Lejeune, F.; Kleeberg, U.R.; Coebergh, J.W.; Eggermont, A.M.; de Vries, E. Superior outcome of women with stage I/II cutaneous melanoma: Pooled analysis of four European Organisation for Research and Treatment of Cancer phase III trials. *J. Clin. Oncol.* **2012**, *30*, 2240–2247. [\[CrossRef\]](https://doi.org/10.1200/JCO.2011.38.0584) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22547594)
- 213. Joosse, A.; de Vries, E.; Eckel, R.; Nijsten, T.; Eggermont, A.M.; Hölzel, D.; Coebergh, J.W.; Engel, J.; Munich Melanoma Group. Gender differences in melanoma survival: Female patients have a decreased risk of metastasis. *J. Investig. Dermatol.* **2011**, *131*, 719–726. [\[CrossRef\]](https://doi.org/10.1038/jid.2010.354) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21150923)
- 214. Sondak, V.K.; Swetter, S.M.; Berwick, M.A. Gender disparities in patients with melanoma: Breaking the glass ceiling. *J. Clin. Oncol.* **2012**, *30*, 2177–2178. [\[CrossRef\]](https://doi.org/10.1200/JCO.2011.41.3849)
- 215. Gamba, C.S.; Clarke, C.A.; Keegan, T.H.; Tao, L.; Swetter, S.M. Melanoma survival disadvantage in young, non-Hispanic white males compared with females. *JAMA Dermatol.* **2013**, *149*, 912–920. [\[CrossRef\]](https://doi.org/10.1001/jamadermatol.2013.4408)
- 216. Fruehauf, J.P.; Trapp, V. Reactive oxygen species: An Achilles' heel of melanoma? *Expert Rev. Anticancer Ther.* **2008**, *8*, 1751–1757. [\[CrossRef\]](https://doi.org/10.1586/14737140.8.11.1751)
- 217. Liu, J.; Zheng, R.; Zhang, Y.; Jia, S.; He, Y.; Liu, J. The Cross Talk between Cellular Senescence and Melanoma: From Molecular Pathogenesis to Target Therapies. *Cancers* **2023**, *15*, 2640. [\[CrossRef\]](https://doi.org/10.3390/cancers15092640)
- 218. Bittinger, F.; Gonzalez-Garcia, J.L.; Klein, C.L.; Brochhausen, C.; Offner, F.; Kirkpatrick, C.J. Production of superoxide by human malignant melanoma cells. *Melanoma Res.* **1998**, *8*, 381–387. [\[CrossRef\]](https://doi.org/10.1097/00008390-199810000-00001)
- 219. Meyskens, F.L., Jr.; Chau, H.V.; Tohidian, N.; Buckmeier, J. Luminol-enhanced chemiluminescent response of human melanocytes and melanoma cells to hydrogen peroxide stress. *Pigment Cell Res.* **1997**, *10*, 184–189. [\[CrossRef\]](https://doi.org/10.1111/j.1600-0749.1997.tb00482.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9266607)
- 220. Joosse, A.; De Vries, E.; van Eijck, C.H.; Eggermont, A.M.; Nijsten, T.; Coebergh, J.W. Reactive oxygen species and melanoma: An explanation for gender differences in survival? *Pigment Cell Melanoma Res.* **2010**, *23*, 352–364. [\[CrossRef\]](https://doi.org/10.1111/j.1755-148X.2010.00694.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20218981)
- 221. Sander, C.S.; Hamm, F.; Elsner, P.; Thiele, J.J. Oxidative stress in malignant melanoma and non-melanoma skin cancer. *Br. J. Dermatol.* **2003**, *148*, 913–922. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2133.2003.05303.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12786821)
- 222. Meyskens, F.L., Jr.; McNulty, S.E.; Buckmeier, J.A.; Tohidian, N.B.; Spillane, T.J.; Kahlon, R.S.; Gonzalez, R.I. Aberrant redox regulation in human metastatic melanoma cells compared to normal melanocytes. *Free Radic. Biol. Med.* **2001**, *31*, 799–808. [\[CrossRef\]](https://doi.org/10.1016/S0891-5849(01)00650-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11557318)
- 223. Nishikawa, M. Reactive oxygen species in tumor metastasis. *Cancer Lett.* **2008**, *266*, 53–59. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2008.02.031) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18362051)
- 224. Trouba, K.J.; Hamadeh, H.K.; Amin, R.P.; Germolec, D.R. Oxidative stress and its role in skin disease. *Antioxid. Redox Signal.* **2002**, *4*, 665–673. [\[CrossRef\]](https://doi.org/10.1089/15230860260220175)
- 225. Thomas-Ahner, J.M.; Wulff, B.C.; Tober, K.L.; Kusewitt, D.F.; Riggenbach, J.A.; Oberyszyn, T.M. Gender differences in UVB-induced skin carcinogenesis, inflammation, and DNA damage. *Cancer Res.* **2007**, *67*, 3468–3474. [\[CrossRef\]](https://doi.org/10.1158/0008-5472.CAN-06-3798)
- 226. Zhong, Q.Y.; Lin, B.; Chen, Y.T.; Huang, Y.P.; Feng, W.P.; Wu, Y.; Long, G.H.; Zou, Y.N.; Liu, Y.; Lin, B.Q.; et al. Gender differences in UV-induced skin inflammation, skin carcinogenesis and systemic damage. *Environ. Toxicol. Pharmacol.* **2021**, *81*, 103512. [\[CrossRef\]](https://doi.org/10.1016/j.etap.2020.103512)
- 227. Masback, A.; Olsson, H.; Westerdahl, J.; Ingvar, C.; Jonsson, N. Prognostic factors in invasive cutaneous malignant melanoma: A population-based study and review. *Melanoma Res.* **2001**, *11*, 435–445. [\[CrossRef\]](https://doi.org/10.1097/00008390-200110000-00001)
- 228. Borrás, C.; Gambini, J.; López-Grueso, R.; Pallardó, F.V.; Viña, J. Direct antioxidant and protective effect of estradiol on isolated mitochondria. *Biochim. Biophys. Acta* **2010**, *1802*, 205–211. [\[CrossRef\]](https://doi.org/10.1016/j.bbadis.2009.09.007)
- 229. Duffield-Lillico, A.J.; Reid, M.E.; Turnbull, B.W.; Combs, G.F., Jr.; Slate, E.H.; Fischbach, L.A.; Marshall, J.R.; Clark, L.C. Baseline characteristics and the effect of selenium supplementation on cancer incidence in a randomized clinical trial: A summary report of the Nutritional Prevention of Cancer Trial. *Cancer Epidemiol. Biomarkers Prev.* **2002**, *11*, 630–639.
- 230. Hercberg, S.; Ezzedine, K.; Guinot, C.; Preziosi, P.; Galan, P.; Bertrais, S.; Estaquio, C.; Briançon, S.; Favier, A.; Latreille, J.; et al. Antioxidant supplementation increases the risk of skin cancers in women but not in men. *J. Nutr.* **2007**, *137*, 2098–2105. [\[CrossRef\]](https://doi.org/10.1093/jn/137.9.2098)
- 231. Hercberg, S.; Galan, P.; Preziosi, P.; Bertrais, S.; Mennen, L.; Malvy, D.; Roussel, A.M.; Favier, A.; Briancon, S. The SU. VI. MAX Study: A randomized, placebo-controlled trial of the health effects of antioxidant vitamins and minerals. *Arch. Intern. Med.* **2004**, *164*, 2335–2342. [\[CrossRef\]](https://doi.org/10.1001/archinte.164.21.2335) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15557412)
- 232. Radkiewicz, C.; Bruchfeld, J.B.; Weibull, C.E.; Jeppesen, M.L.; Frederiksen, H.; Lambe, M.; Jakobsen, L.; El-Galaly, T.C.; Smedby, K.E.; Wästerlid, T. Sex differences in lymphoma incidence and mortality by subtype: A population-based study. *Am. J. Hematol.* **2023**, *98*, 23–30. [\[CrossRef\]](https://doi.org/10.1002/ajh.26744)
- 233. Jacobus, J.A.; Duda, C.G.; Coleman, M.C.; Martin, S.M.; Mapuskar, K.; Mao, G.; Smith, B.J.; Aykin-Burns, N.; Guida, P.; Gius, D.; et al. Low-dose radiation-induced enhancement of thymic lymphomagenesis in Lck-Bax mice is dependent on LET and gender. *Radiat. Res.* **2013**, *180*, 156–165. [\[CrossRef\]](https://doi.org/10.1667/RR3293.1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23819597)
- 234. Štěrba, M.; Popelová, O.; Vávrová, A.; Jirkovský, E.; Kovaříková, P.; Geršl, V.; Šimůnek, T. Oxidative stress, redox signaling, and metal chelation in anthracycline cardiotoxicity and pharmacological cardioprotection. *Antioxid. Redox Signal.* **2013**, *18*, 899–929. [\[CrossRef\]](https://doi.org/10.1089/ars.2012.4795)
- 235. Malorni, W.; Campesi, I.; Straface, E.; Vella, S.; Franconi, F. Redox features of the cell: A gender perspective. *Antioxid. Redox Signal.* **2007**, *9*, 1779–1801. [\[CrossRef\]](https://doi.org/10.1089/ars.2007.1596) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17822369)
- 236. Vijay, V.; Han, T.; Moland, C.L.; Kwekel, J.C.; Fuscoe, J.C.; Desai, V.G. Sexual dimorphism in the expression of mitochondriarelated genes in rat heart at different ages. *PLoS ONE* **2015**, *10*, e0117047. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0117047)
- 237. Dobbs, N.A.; Twelves, C.J.; Gillies, H.; James, C.A.; Harper, P.G.; Rubens, R.D. Gender affects the doxorubicin pharmacokinetics in patients with normal liver biochemistry. *Cancer Chemother. Pharmacol.* **1995**, *36*, 473–476. [\[CrossRef\]](https://doi.org/10.1007/BF00685796)
- 238. Wade, J.R.; Kelman, A.W.; Kerr, D.J.; Robert, J.; Whiting, B. Variability in the pharmacokinetics of epirubicin: A population analysis. *Cancer Chemother. Pharmacol.* **1992**, *29*, 391–395. [\[CrossRef\]](https://doi.org/10.1007/BF00686009) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1551178)
- 239. Varricchi, G.; Ameri, P.; Cadeddu, C.; Ghigo, A.; Madonna, R.; Marone, G.; Mercurio, V.; Monte, I.; Novo, G.; Parrella, P.; et al. Antineoplastic drug-induced cardiotoxicity: A redox perspective. *Front. Physiol.* **2018**, *9*, 167. [\[CrossRef\]](https://doi.org/10.3389/fphys.2018.00167)
- 240. Tocchetti, C.G.; Cadeddu, C.; Di Lisi, D.; Femminò, S.; Madonna, R.; Mele, D.; Monte, I.; Novo, G.; Penna, C.; Pepe, A.; et al. From molecular mechanisms to clinical management of antineoplastic drug-induced cardiovascular toxicity: A translational overview. *Antioxid. Redox Signal.* **2019**, *30*, 2110–2153. [\[CrossRef\]](https://doi.org/10.1089/ars.2016.6930)
- 241. Pupo, M.; Pisano, A.; Abonante, S.; Maggiolini, M.; Musti, A.M. GPER activates Notch signaling in breast cancer cells and cancerassociated fibroblasts (CAFs). *Int. J. Biochem. Cell Biol.* **2014**, *46*, 56–67. [\[CrossRef\]](https://doi.org/10.1016/j.biocel.2013.11.011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24275097)
- 242. Lubecka, K.; Kurzava, L.; Flower, K.; Buvala, H.; Zhang, H.; Teegarden, D.; Camarillo, I.; Suderman, M.; Kuang, S.; Andrisani, O.; et al. Stilbenoids remodel the DNA methylation patterns in breast cancer cells and inhibit oncogenic NOTCH signaling through epigenetic regulation of MAML2 transcriptional activity. *Carcinogenesis* **2016**, *37*, 656–668. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgw048) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27207652)
- 243. Gonzalez, Y.; Pokrzywinski, K.L.; Rosen, E.T.; Mog, S.; Aryal, B.; Chehab, L.M.; Vijay, V.; Moland, C.L.; Desai, V.G.; Dickey, J.S.; et al. Reproductive hormone levels and differential mitochondria-related oxidative gene expression as potential mechanisms for gender differences in cardiosensitivity to doxorubicin in tumor-bearing spontaneously hypertensive rats. *Cancer Chemother. Pharmacol.* **2015**, *76*, 447–459. [\[CrossRef\]](https://doi.org/10.1007/s00280-015-2786-8)
- 244. Octavia, Y.T.C.; Gabrielson, K.L.; Janssens, S.; Crijns, H.J.; Moens, A.L. Doxorubicin-induced cardiomyopathy. *J. Mol. Cell. Cardiol.* **2012**, *52*, 1213–1225. [\[CrossRef\]](https://doi.org/10.1016/j.yjmcc.2012.03.006)
- 245. Carvalho, F.S.; Burgeiro, A.; Garcia, R.; Moreno, A.J.; Carvalho, R.A.; Oliveira, P.J. Doxorubicin-induced cardiotoxicity: From bioenergetic failure and cell death to cardiomyopathy. *Med. Res. Rev.* **2014**, *34*, 106–135. [\[CrossRef\]](https://doi.org/10.1002/med.21280) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23494977)
- 246. Christiansen, S.; Autschbach, R. Doxorubicin in experimental and clinical heart failure. *Eur. J. Cardiothorac. Surg.* **2006**, *30*, 611–616. [\[CrossRef\]](https://doi.org/10.1016/j.ejcts.2006.06.024)
- 247. Kumar, S.; Marfatia, R.; Tannenbaum, S.; Yang, C.; Avelar, E. Doxorubicin-induced cardiomyopathy 17 years after chemotherapy. *Texas Heart Inst. J.* **2012**, *39*, 424–427.
- 248. Dimitrova, K.R.; DeGroot, K.; Myers, A.K.; Kim, Y.D. Estrogen and homocysteine. *Cardiovasc. Res.* **2002**, *53*, 577–588. [\[CrossRef\]](https://doi.org/10.1016/S0008-6363(01)00462-X)
- 249. Ogita, H.; Node, K.; Kitakaze, M. The role of estrogen and estrogen-related drugs in cardiovascular diseases. *Curr. Drug Metab.* **2003**, *4*, 497–504. [\[CrossRef\]](https://doi.org/10.2174/1389200033489271)
- 250. Hershman, D.L.; McBride, R.B.; Eisenberger, A.; Tsai, W.Y.; Grann, V.R.; Jacobson, J.S. Doxorubicin, cardiac risk factors, and cardiac toxicity in elderly patients with diffuse B-cell non-Hodgkin's lymphoma. *J. Clin. Oncol.* **2008**, *26*, 3159–3165. [\[CrossRef\]](https://doi.org/10.1200/JCO.2007.14.1242)
- 251. Biancaniello, T.; Meyer, R.A.; Wong, K.Y.; Sager, C.; Kaplan, S. Doxorubicin cardiotoxicity in children. *J. Pediatr.* **1980**, *97*, 45–50. [\[CrossRef\]](https://doi.org/10.1016/S0022-3476(80)80128-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7381647)
- 252. Bell, J.R.; Bernasochi, G.B.; Varma, U.; Raaijmakers, A.J.; Delbridge, L.M. Sex and sex hormones in cardiac stress—Mechanistic insights. *J. Steroid Biochem. Mol. Biol.* **2012**, *137*, 124–135. [\[CrossRef\]](https://doi.org/10.1016/j.jsbmb.2013.05.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23770428)
- 253. Zhang, J.; Knapton, A.; Lipshultz, S.E.; Cochran, T.R.; Hiraragi, H.; Herman, E.H. Sex-related differences in mast cell activity and doxorubicin toxicity: A study in spontaneously hypertensive rats. *Toxicol. Pathol.* **2014**, *42*, 361–375. [\[CrossRef\]](https://doi.org/10.1177/0192623313482778) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23531790)
- 254. Belham, M.; Kruger, A.; Mepham, S.; Faganello, G.; Pritchard, C. Monitoring left ventricular function in adults receiving anthracycline-containing chemotherapy. *Eur. J. Heart Fail.* **2007**, *9*, 409–414. [\[CrossRef\]](https://doi.org/10.1016/j.ejheart.2006.09.007)
- 255. Lin, D.Y.T.F.; Tsai, C.H.; Huang, C.Y. Mechanisms governing the protective effect of 17B-estradiol and estrogen receptors against cardiomyocyte injury. *BioMedicine* **2011**, *1*, 21–28. [\[CrossRef\]](https://doi.org/10.1016/j.biomed.2011.10.004)
- 256. International Commission on Radiological Protection (ICRP). *Assessing Dose of the Representative Person for the Purpose of Radiation Protection of the Public and the Optimisation of Radiological Protection: Broadening the Process*; ICRP Publication 101; ICRP: Ottawa, ON, Canada, 2006.
- 257. International Commission on Radiological Protection (ICRP). *Adult Reference Computational Phantoms*; ICRP Publication 110; ICRP: Ottawa, ON, Canada, 2009.
- 258. Langen, B.; Vorontsov, E.; Spetz, J.; Swanpalmer, J.; Sihlbom, C.; Helou, K.; Forssell-Aronsson, E. Age and sex effects across the blood proteome after ionizing radiation exposure can bias biomarker screening and risk assessment. *Sci. Rep.* **2022**, *12*, 7000. [\[CrossRef\]](https://doi.org/10.1038/s41598-022-10271-3)
- 259. Yahyapour, R.; Motevaseli, E.; Rezaeyan, A.; Abdollahi, H.; Farhood, B.; Cheki, M.; Najafi, M.; Villa, V. Mechanisms of Radiation Bystander and Non-Targeted Effects: Implications to Radiation Carcinogenesis and Radiotherapy. *Curr. Radiopharm.* **2018**, *11*, 34–45. [\[CrossRef\]](https://doi.org/10.2174/1874471011666171229123130)
- 260. Guizard, A.; Boutou, O.; Pottier, D.; Troussard, X.; Pheby, D.; Launoy, G.; Slama, R.; Spira, A. The incidence of childhood leukaemia around the La Hague nuclear waste reprocessing plant (France): A survey for the years 1978–1998. *J. Epidemiol. Comm. Health* **2001**, *55*, 469–474. [\[CrossRef\]](https://doi.org/10.1136/jech.55.7.469)
- 261. Yoshida, K.; Nemoto, K.; Nishimura, M.; Seki, M. Exacerbating factors of radiation-induced myeloid leukemogenesis. *Leukemia Res.* **1993**, *17*, 437–440. [\[CrossRef\]](https://doi.org/10.1016/0145-2126(93)90099-7)
- 262. Kovalchuk, O.; Burke, P.; Besplug, J.; Slovack, M.; Filkowski, J.; Pogribny, I. Methylation changes in muscle and liver tissues of male and female mice exposed to acute and chronic low-dose Xray-irradiation. *Mutat. Res.* **2004**, *548*, 75–84. [\[CrossRef\]](https://doi.org/10.1016/j.mrfmmm.2003.12.016)
- 263. Koturbash, I.; Zemp, F.; Kolb, B.; Kovalchuk, O. Sex-specific radiation-induced microRNAome responses in the hippocampus, cerebellum and frontal cortex in a mouse model. *Mutat. Res.* **2011**, *722*, 114–118. [\[CrossRef\]](https://doi.org/10.1016/j.mrgentox.2010.05.007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20478395)
- 264. Koturbash, I.; Kutanzi, K.; Hendrickson, K.; Rodriguez-Juarez, R.; Kogosov, D.; Kovalchuk, O. Radiation-induced bystander effects in vivo are sex specific. *Mutat. Res.* **2008**, *642*, 28–36. [\[CrossRef\]](https://doi.org/10.1016/j.mrfmmm.2008.04.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18508093)
- 265. Koturbash, I.; Zemp, F.J.; Kutanzi, K.; Luzhna, L.; Loree, J.; Kolb, B.; Kovalchuk, O. Sex-specific microRNAome deregulation in the shielded bystander spleen of cranially exposed mice. *Cell Cycle* **2008**, *7*, 1658–1667. [\[CrossRef\]](https://doi.org/10.4161/cc.7.11.5981) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18560276)
- 266. Chai, Y.; Calaf, G.M.; Zhou, H.; Ghandhi, S.A.; Elliston, C.D.; Wen, G.; Nohmi, T.; Amundson, S.A.; Hei, T.K. Radiation induced COX-2 expression and mutagenesis at non-targeted lung tissues of gpt delta transgenic mice. *Br. J. Cancer* **2013**, *108*, 91–98. [\[CrossRef\]](https://doi.org/10.1038/bjc.2012.498)
- 267. Cheng, C.; Omura Minamisawa, M.; Kang, Y.; Hara, T.; Koike, I.; Inoue, T. Quantification of circulating cellfree DNA in the plasma of cancer patients during radiation therapy. *Cancer Sci.* **2009**, *100*, 303–309. [\[CrossRef\]](https://doi.org/10.1111/j.1349-7006.2008.01021.x)
- 268. Ma, Y.; Zhang, L.; Rong, S.; Qu, H.; Zhang, Y.; Chang, D.; Pan, H.; Wang, W. Relation between gastric cancer and protein oxidation, DNA damage, and lipid peroxidation. *Oxid. Med. Cell Longev.* **2013**, *2013*, 543760. [\[CrossRef\]](https://doi.org/10.1155/2013/543760)
- 269. Deligezer, U.; Eralp, Y.; Akisik, E.Z.; Akisik, E.E.; Saip, P.; Topuz, E.; Dalay, N. Effect of adjuvant chemotherapy on integrity of free serum DNA in patients with breast cancer. *Ann. N. Y. Acad Sci.* **2008**, *1137*, 175–179. [\[CrossRef\]](https://doi.org/10.1196/annals.1448.010)
- 270. He, K.; Zhang, S.; Shao, L.L.; Yin, J.C.; Wu, X.; Shao, Y.W.; Yuan, S.; Yu, J. Developing more sensitive genomic approaches to detect radioresponse in precision radiation oncology: From tissue DNA analysis to circulating tumor DNA. *Cancer Lett.* **2020**, *472*, 108–118. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2019.12.004)
- 271. Mbugua, S.N. Targeting Tumor Microenvironment by Metal Peroxide Nanoparticles in Cancer Therapy. *Bioinorg. Chem. Appl.* **2022**, *2022*, 5041399. [\[CrossRef\]](https://doi.org/10.1155/2022/5041399)
- 272. Allegra, A.; Tonacci, A.; Pioggia, G.; Musolino, C.; Gangemi, S. Anticancer Activity of *Rosmarinus officinalis* L.: Mechanisms of Action and Therapeutic Potentials. *Nutrients* **2020**, *12*, 1739. [\[CrossRef\]](https://doi.org/10.3390/nu12061739)
- 273. Allegra, A.; Speciale, A.; Molonia, M.S.; Guglielmo, L.; Musolino, C.; Ferlazzo, G.; Costa, G.; Saija, A.; Cimino, F. Curcumin ameliorates the in vitro efficacy of carfilzomib in human multiple myeloma U266 cells targeting p53 and NF-κB pathways. *Toxicol. In Vitro* **2018**, *47*, 186–194. [\[CrossRef\]](https://doi.org/10.1016/j.tiv.2017.12.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29223572)
- 274. Cristani, M.; Speciale, A.; Saija, A.; Gangemi, S.; Minciullo, P.L.; Cimino, F. Circulating Advanced Oxidation Protein Products as Oxidative Stress Biomarkers and Progression Mediators in Pathological Conditions Related to Inflammation and Immune Dysregulation. *Curr. Med. Chem.* **2016**, *23*, 3862–3882. [\[CrossRef\]](https://doi.org/10.2174/0929867323666160902154748) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27593960)
- 275. Kim, S.Y. Oxidative stress and gender disparity in cancer. *Free Radic. Res.* **2022**, *56*, 90–105. [\[CrossRef\]](https://doi.org/10.1080/10715762.2022.2038789) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35118928)
- 276. May, R.C. Gender, immunity and the regulation of longevity. *Bioessays* **2007**, *29*, 795–802. [\[CrossRef\]](https://doi.org/10.1002/bies.20614) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17621669)
- 277. Malorni, W.; Straface, E.; Matarrese, P.; Ascione, B.; Coinu, R.; Canu, S.; Galluzzo, P.; Marino, M.; Franconi, F. Redox state and gender differences in vascular smooth muscle cells. *FEBS Lett.* **2008**, *582*, 635–642. [\[CrossRef\]](https://doi.org/10.1016/j.febslet.2008.01.034) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18242172)
- 278. Vina, J.; Sastre, J.; Pallardo, F.; Borras, C. Mitochondrial theory of aging: Importance to explain why females live longer than males. *Antioxid. Redox Signal.* **2003**, *5*, 549–556. [\[CrossRef\]](https://doi.org/10.1089/152308603770310194)
- 279. Loft, S.; Vistisen, K.; Ewertz, M.; Tjonneland, A.; Overvad, K.; Poulsen, H.E. Oxidative DNA damage estimated by 8-hydroxydeoxyguanosine excretion in humans: Influence of smoking, gender and body mass index. *Carcinogenesis* **1992**, *13*, 2241–2247. [\[CrossRef\]](https://doi.org/10.1093/carcin/13.12.2241)
- 280. Marnett, L.J. Oxyradicals and DNA damage. *Carcinogenesis* **2000**, *21*, 361–370. [\[CrossRef\]](https://doi.org/10.1093/carcin/21.3.361)
- 281. Bokov, A.F.; Ko, D.; Richardson, A. The effect of gonadectomy and estradiol on sensitivity to oxidative stress. *Endocr. Res.* **2009**, *34*, 43–58. [\[CrossRef\]](https://doi.org/10.1080/07435800902913600)
- 282. Luo, S.; Valencia, C.A.; Zhang, J.; Lee, N.C.; Slone, J.; Gui, B.; Wang, X.; Li, Z.; Dell, S.; Brown, J.; et al. Biparental inheritance of mitochondrial DNA in humans. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 13039–13044. [\[CrossRef\]](https://doi.org/10.1073/pnas.1810946115)
- 283. Rius, R.; Cowley, M.J.; Riley, L.; Puttick, C.; Thorburn, D.R.; Christodoulou, J. Biparental inheritance of mitochondrial DNA in humans is not a common phenomenon. *Genet. Med.* **2019**, *21*, 2823–2826. [\[CrossRef\]](https://doi.org/10.1038/s41436-019-0568-0)
- 284. Lutz-Bonengel, S.; Parson, W. No further evidence for paternal leakage of mitochondrial DNA in humans yet. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 1821–1822. [\[CrossRef\]](https://doi.org/10.1073/pnas.1820533116)
- 285. Gaignard, P.; Liere, P.; Thérond, P.; Schumacher, M.; Slama, A.; Guennoun, R. Role of sex hormones on brain mitochondrial function, with special reference to aging and neurodegenerative diseases. *Front. Aging Neurosci.* **2017**, *9*, 2017. [\[CrossRef\]](https://doi.org/10.3389/fnagi.2017.00406) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29270123)
- 286. Ventura-Clapier, R.; Moulin, M.; Piquereau, J.; Lemaire, C.; Mericskay, M.; Veksler, V.; Garnier, A. Mitochondria: A central target for sex differences in pathologies. *Clin. Sci.* **2017**, *131*, 803–822. [\[CrossRef\]](https://doi.org/10.1042/CS20160485) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28424375)
- 287. Nishigaki, Y.; Marti, R.; Copeland, W.C.; Hirano, M. Site-specific somatic mitochondrial DNA point mutations in patients with thymidine phosphorylase deficiency. *J. Clin. Investig.* **2003**, *111*, 1913–1921. [\[CrossRef\]](https://doi.org/10.1172/JCI17828)
- 288. Elamin, Y.Y.; Rafee, S.; Osman, N.; O'Byrne, K.J.; Gately, K. Thymidine phosphorylase in cancer; enemy or friend? *Cancer Microenviron.* **2016**, *9*, 33–43. [\[CrossRef\]](https://doi.org/10.1007/s12307-015-0173-y)
- 289. Cetrone, M.; Mele, A.; Tricarico, D. Effects of the antidiabetic drugs on the age-related atrophy and sarcopenia associated with diabetes type II. *Curr. Diabetes Rev.* **2014**, *10*, 231–237. [\[CrossRef\]](https://doi.org/10.2174/1573399810666140918121022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25245021)
- 290. Hasegawa, K.; Okamoto, H.; Kawamura, K.; Kato, R.; Kobayashi, Y.; Sekiya, T.; Udagawa, Y. The effect of chemotherapy or radiotherapy on thymidine phosphorylase and dihydropyrimidine dehydrogenase expression in cancer of the uterine cervix. *Eur. J. Obstet. Gynecol. Reprod Biol.* **2012**, *163*, 67–70. [\[CrossRef\]](https://doi.org/10.1016/j.ejogrb.2012.03.014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22480411)
- 291. Bijnsdorp, I.V.; de Bruin, M.; Laan, A.C.; Fukushima, M.; Peters, G.J. The role of platelet-derived endothelial cell growth factor/thymidine phosphorylase in tumor behavior. *Nucleosides Nucleotides Nucleic Acids* **2008**, *27*, 681–691. [\[CrossRef\]](https://doi.org/10.1080/15257770802143988) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18600526)
- 292. Ioachim, E. Thymidine phoshorylase expression in breast cancer: The prognostic significance and its association with other angiogenesis related proteins and extracellular matrix components. *Histol. Histopathol.* **2008**, *23*, 187–196. [\[CrossRef\]](https://doi.org/10.14670/HH-23.187) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17999375)
- 293. Zhang, L.; Mackenzie, I.Z.; Rees, M.C.; Bicknell, R. Regulation of the expression of the angiogenic enzyme platelet-derived endothelial cell growth factor/thymidine phosphorylase in endometrial isolates by ovarian steroids and cytokines. *Endocrinology* **1997**, *138*, 4921–4930. [\[CrossRef\]](https://doi.org/10.1210/endo.138.11.5517)
- 294. Zizzo, N.; Passantino, G.; D'alessio, R.M.; Tinelli, A.; Lopresti, G.; Patruno, R.; Tricarico, D.; Maqoud, F.; Scala, R.; Zito, F.A.; et al. Thymidine Phosphorylase Expression and Microvascular Density Correlation Analysis in Canine Mammary Tumor: Possible Prognostic Factor in Breast Cancer. *Front. Vet. Sci.* **2019**, *6*, 368. [\[CrossRef\]](https://doi.org/10.3389/fvets.2019.00368) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31709268)
- 295. Canady, K.S.; Ali-Osman, F.; Rubel, E.W. Extracellular potassium influences DNA and protein syntheses and glial fibrillary acidic protein expression in cultured glial cells. *Glia* **1990**, *3*, 368–374. [\[CrossRef\]](https://doi.org/10.1002/glia.440030508) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2146224)
- 296. March, H.N.; Rust, A.G.; Wright, N.A.; ten Hoeve, J.; de Ridder, J.; Eldridge, M.; van der Weyden, L.; Berns, A.; Gadiot, J.; Uren, A.; et al. Insertional mutagenesis identifies multiple networks of cooperating genes driving intestinal tumorigenesis. *Nat. Genet.* **2011**, *43*, 1202–1209. [\[CrossRef\]](https://doi.org/10.1038/ng.990)
- 297. Morch, L.S.; Lidegaard, O.; Keiding, N.; Lokkegaard, E.; Kjaer, S.K. The influence of hormone therapies on colon and rectal cancer. *Eur. J. Epidemiol.* **2016**, *31*, 481–489. [\[CrossRef\]](https://doi.org/10.1007/s10654-016-0116-z)
- 298. Than, B.L.; Goos, J.A.; Sarver, A.L.; O'Sullivan, M.G.; Rod, A.; Starr, T.K.; Fijneman, R.J.; Meijer, G.A.; Zhao, L.; Zhang, Y.; et al. The role of KCNQ1 in mouse and human gastrointestinal cancers. *Oncogene* **2014**, *33*, 3861–3868. [\[CrossRef\]](https://doi.org/10.1038/onc.2013.350) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23975432)
- 299. Rapetti-Mauss, R.; O'Mahony, F.; Sepulveda, F.V.; Urbach, V.; Harvey, B. J Oestrogen promotes KCNQ1 potassium channel endocytosis and postendocytic trafficking in colonic epithelium. *J. Physiol.* **2013**, *591*, 2813–2831. [\[CrossRef\]](https://doi.org/10.1113/jphysiol.2013.251678)
- 300. Rapetti-Mauss, R.; Bustos, V.; Thomas, W.; McBryan, J.; Harvey, H.; Lajczak, N.; Madden, S.F.; Pellissier, B.; Borgese, F.; Soriani, O.; et al. Bidirectional KCNQ1: Beta-catenin interaction drives colorectal cancer cell differentiation. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 4159–4164. [\[CrossRef\]](https://doi.org/10.1073/pnas.1702913114)
- 301. Haziman, A.A.; Ravinderan, S.; Thangavelu, T.; Thomas, W. A novel role for estrogen-induced signaling in the colorectal cancer gender bias. *Ir. J. Med. Sci.* **2019**, *188*, 389–395. [\[CrossRef\]](https://doi.org/10.1007/s11845-018-1867-1)
- 302. Tricarico, D.; Barbieri, M.; Antonio, L.; Tortorella, P.; Loiodice, F.; Camerino, D.C. Dualistic actions of cromakalim and new potent 2H-1,4-benzoxazine derivatives on the native skeletal muscle K ATP channel. *Br. J. Pharmacol.* **2003**, *139*, 255–262. [\[CrossRef\]](https://doi.org/10.1038/sj.bjp.0705233)
- 303. Niu, K.; Saloman, J.L.; Zhang, Y.; Ro, J.Y. Sex differences in the contribution of ATP-sensitive K⁺ channels in trigeminal ganglia under an acute muscle pain condition. *Neuroscience* **2011**, *180*, 344–352. [\[CrossRef\]](https://doi.org/10.1016/j.neuroscience.2011.01.045) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21296645)
- 304. Malhi, H.; Irani, A.N.; Rajvanshi, P.; Suadicani, S.O.; Spray, D.C.; McDonald, T.V.; Gupta, S. KATP channels regulate mitogenically induced proliferation in primary rat hepatocytes and human liver cell lines. Implications for liver growth control and potential therapeutic targeting. *J. Biol. Chem.* **2000**, *275*, 26050–26057. [\[CrossRef\]](https://doi.org/10.1074/jbc.M001576200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10862612)
- 305. Huang, L.; Li, B.; Li, W.; Guo, H.; Zou, F. ATP-sensitive potassium channels control glioma cells proliferation by regulating ERK activity. *Carcinogenesis* **2009**, *30*, 737–744. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgp034)
- 306. Maqoud, F.; Zizzo, N.; Attimonelli, M.; Tinelli, A.; Passantino, G.; Antonacci, M.; Ranieri, G.; Tricarico, D. Immunohistochemical, pharmacovigilance, and omics analyses reveal the involvement of ATP-sensitive K⁺ channel subunits in cancers: Role in drug-disease interactions. *Front. Pharmacol.* **2023**, *14*, 1115543. [\[CrossRef\]](https://doi.org/10.3389/fphar.2023.1115543) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37180726)

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