

SHORT REPORT

Open Access



Detection of vector-borne pathogens in cats and their ectoparasites in southern Italy

Maria-Flaminia Persichetti^{1†}, Laia Solano-Gallego^{2*†}, Lorena Serrano³, Laura Altet³, Stefano Reale¹, Marisa Masucci⁴ and Maria-Grazia Pennisi^{4†}

Abstract

Background: Vector-borne pathogens are the subject of several investigations due to the zoonotic concern of some of them. However, limited data are available about the simultaneous presence of these pathogens in cats and their ectoparasites. The aim of the present study was to define the species of ectoparasites found on cats as well as to investigate vector-borne pathogens in cats and their ectoparasites in southern Italy.

Methods: Blood from 42 cats and fleas or flea pools ($n = 28$) and ticks ($n = 73$) collected from them were investigated by quantitative PCR for the detection of vector-borne pathogens. Feline serum samples were tested by IFAT to detect IgG antibodies against *Leishmania infantum*, *Bartonella henselae*, *Rickettsia conorii*, *Rickettsia felis*, *Rickettsia typhi*, *Babesia microti*, *Ehrlichia canis* and *Anaplasma phagocytophilum* antigens.

Results: Only one flea species (*Ctenocephalides felis*) and four tick species belonging to the genera *Rhipicephalus* and *Ixodes* were identified on cats from southern Italy. Molecular evidence of *Bartonella* spp., *Rickettsia* spp., hemoplasmas, *Babesia vogeli* and *L. infantum* was found in ectoparasites (fleas and/or ticks) while DNA from *Hepatozoon felis* and *Ehrlichia/Anaplasma* spp. was not detected. Likewise, DNAs from *Bartonella*, hemoplasma and *Leishmania* were the only pathogens amplified from feline blood samples. Cats had also antibodies against all the investigated pathogens with the exception of *Rickettsia typhi*. Agreement between serological and molecular results in individual cats and their ectoparasites was not found. The only exception was for *Bartonella* with a fair to moderate agreement between individual cats and their ectoparasites. *Bartonella clarridgeiae* was the species most frequently found in cats and their fleas followed by *B. henselae*.

Conclusions: In conclusion, cats harboring ticks and fleas are frequently exposed to vector-borne pathogens. Furthermore, ticks and fleas harbored by cats frequently carry pathogens of zoonotic concern therefore appropriate feline ectoparasiticide preventative treatments should be used in cats.

Keywords: Vector-borne pathogens, Flea, Tick, Cat

Background

Ticks, fleas and mosquitoes are globally distributed and their ability to transmit pathogens gives them important medical relevance. On the other hand, the growing success of pets in developed countries, especially the cat with its independent lifestyle, results in an increased risk for

humans of contact with feline ectoparasites [1]. It is well known that the most frequent flea species found on cats is *Ctenocephalides felis* [2]. Conversely, limited information is available about the species of ticks which infest cats and vector-borne pathogens (VBPs) harbored by them [2–10]. In addition, the comparison of vector-borne pathogens from cats and from their ectoparasites (fleas and ticks) has not been fully explored [11].

The aims of this investigation which was carried out in two regions (Calabria and Sicily) of southern Italy were: (i) to evaluate the flea and tick species collected from

* Correspondence: laia.solano@uab.cat
†Equal contributors

²Departament de Medicina i Cirurgia Animal, Facultat de Veterinària, Universitat Autònoma de Barcelona, 08193, Bellaterra, Barcelona, Spain
Full list of author information is available at the end of the article

outdoor domestic cats and determine if they harbor VBPs; (ii) to evaluate exposure of outdoor cats to VBPs by means of antibody and molecular testing; and (iii) to compare the VBPs DNA from feline blood and from the ectoparasites (fleas and ticks) collected from them.

Methods

The present study integrates data already published on 132 ticks collected from a large number of cats ($n = 308$) in Southern Italy and the pathogens that they harbor [4]. We included in this study a total of 42 cats from province of Reggio Calabria ($n = 27$) and from Messina city and Lipari Island in Sicily region ($n = 15$) enrolled between March 2012 and January 2013. These cats were selected based on the following criteria: the presence of at least one ectoparasite (tick or flea) on physical examination, residual ethylenediaminetetraacetic acid (EDTA) blood and serum samples available, signed owner informed consent and outdoor life style. Both sick (22/42 = 52.4 %) and apparently healthy cats (20/42 = 47.6 %) based on clinical history and physical examination were enrolled. Date of sampling, gender, age, breed, lifestyle, vaccination status, ongoing therapy, reason for consultation, physical examination, the number of collected ectoparasites, feeding status of collected ticks as well as antiparasitic treatments of cats were recorded.

Each cat was carefully combed for at least five minutes throughout the whole body surface and inspected for the presence of fleas or/and ticks. All ectoparasites detected were removed by a veterinarian and stored in alcohol 70 % as a preventative measure. Feline blood residual samples were employed in the present study. Therefore, ethical committee approval was not needed. Informed consent was obtained from all owners and from the legal representative of animal welfare groups in charge of the management of stray cats.

Serum from all cats was tested for the detection of immunoglobulin G (IgG) antibodies against *Bartonella henselae*, *Rickettsia conorii*, *Rickettsia felis*, *Rickettsia typhi*, *Ehrlichia canis*, *Babesia microti* and *Anaplasma phagocytophilum* antigens by the immunofluorescence antibody test (IFAT) using commercial kits (Fuller Laboratories Fullerton, California, USA). The manufacturer's protocol was followed for all serological tests using a cut-off dilution of 1:64 for *B. henselae*, *R. conorii*, *R. felis*, *R. typhi* and *B. microti*; and 1:50 for *E. canis* and *A. phagocytophilum*. The presence of *L. infantum* IgG antibodies was investigated using *L. infantum* (strain MHOM/IT/80/IPT1) antigen slides manufactured by the *National reference centre for Leishmaniosis*, (C.Re.Na.L., Palermo, Italy) and fluoresceinated rabbit anti-cat IgG (Anti-IgG-FITC, SIGMA) diluted in PBS 1:200 [12]. The cut-off value was established at 1:80 for *L. infantum* [12].

Morphometric identification of fleas and ticks was made through a stereomicroscope before DNA extraction for polymerase chain reaction (PCR) assays [13, 14]. Afterwards, fleas from each cat were extracted and processed and only for cats carrying more than one single flea, pools were done. Specifically, a number spanning from two to five fleas collected from each cat was pooled for molecular investigations. Conversely, ticks were in any case extracted and processed individually.

DNA extraction from 300 μ l of blood was performed using High Pure PCR Template preparation kit (Roche, Mannheim, Germany). DNA was eluted in 100 μ l of elution buffer and stored at -20 °C until used. DNA extraction from individual ticks, fleas and flea pools was carried out using High Pure PCR Template preparation kit (Roche, Mannheim, Germany) according to the manufacturer's tissue protocol with some modifications. Briefly, all ectoparasites were washed twice in sterile PBS solution for 5 min shaking it slowly, then overnight at 4 °C. Each flea was manually cut by a sterile lancet in four pieces and then suspended in 200 μ l of Tissue Lysis Buffer of the same kit. DNA was eluted in 50 μ l of elution buffer and stored at -20 °C for later analysis.

Real-time PCR technology was applied as described elsewhere [4], to identify specific DNA target for *Ehrlichia/Anaplasma* spp., piroplasmids (*Babesia* spp. and *Theileria* spp.), *Hepatozoon felis*, hemotropic *Mycoplasma* spp., *Rickettsia* spp., *Bartonella* spp. and *L. infantum* from ticks and feline blood samples while only the last four pathogens were investigated on fleas due to economical restrictions. All positive PCR results for each ectoparasite or cat were sequenced according to the Big-Dye Terminator Cycle Sequencing Ready reaction Kit (AB, Life Technologies) using the same primers. Sequences obtained were compared with GenBank database (www.ncbi.nlm.nih.gov/BLAST). All positive PCR results for hemoplasmas or *L. infantum* were not sequenced. Instead, species-specific real time PCRs were performed as described by Martinez et al. [15] to discriminate among feline hemoplasmas species (*Mycoplasma haemofelis* (Mhf), 'Candidatus Mycoplasma haemominutum' (CMhm) and 'Candidatus Mycoplasma turicensis' (CMT)) as well as for *L. infantum* real time PCR [16].

For each pathogen investigated, Kappa agreement test (GraphPad InStat) was used to establish agreement between serological and molecular results in cats, between molecular results in cats, ticks or fleas and between serological results in cats and molecular results in ticks or fleas. The Kappa values were evaluated as follows: no agreement ($k < 0$), slight agreement ($0 < k < 0.2$), fair agreement ($0.2 < k < 0.4$), moderate agreement ($0.41 < k < 0.6$), substantial agreement ($0.61 < k < 0.8$) and almost perfect agreement ($k > 0.81$).

Results

Clinical data and antibody detection in cats

Age of cats ranged from six months to ten years with a median of 1.5 years. Twenty-three cats were females and 19 were males. Only six were not neutered. Thirty-nine were mixed breed domestic short hair cats and six mixed breed domestic long hair cats. Information on ectoparasiticide treatment was available for 40 cats and most of them (35/40 = 87.5 %) were never treated with ectoparasiticide. One cat was only treated during the summer season but the other four cats were monthly treated. Ticks were also detected on these five treated cats.

Thirty-nine of the 42 cats examined (92.9 %) were antibody positive to at least one investigated antigen. Antibodies were detected against all the VBPs studied except for *R. typhi* antigens (Table 1).

Detection and morphological identification of ticks and fleas

Sixty-five cats were infested by both ticks and fleas and were collected from 28 out of the 42 cats and all were identified as *C. felis*. Seventy-three ticks were also removed from 15 cats and only one cat (from Calabria) was infested by both ticks and fleas. Ticks belonged to the genera *Rhipicephalus* ($n = 42$) and *Ixodes* ($n = 31$) and 25 specimens were engorged. In detail, 25 *Rhipicephalus sanguineus* (3 engorged), 17 *Rhipicephalus pusillus*, 19 *Ixodes ventralloii* (16 engorged), ten *Ixodes ricinus* (four engorged), two engorged *Ixodes* spp. were identified. The number of ectoparasites collected from individual cats ranged from one to five fleas with a median value of two fleas and one to 21 ticks with a median value of four ticks. Fleas were mostly (27/28 = 96 %) collected from cats in Calabria Province with the exception of one male flea that was removed from a cat living in Messina city. In contrast, all ticks were found on cats from Lipari island (Messina province) (14/15 = 93 %) with the exception of one tick (*Ixodes ricinus* engorged female) that was removed from a cat living in Calabria Province (1/15 = 7 %).

Table 1 Serological results of investigated pathogens in 42 cats infested by ectoparasites

Antigen	Number of seropositive cats (%)
<i>Bartonella henselae</i>	23 (54.8)
<i>Rickettsia conorii</i>	23 (54.8)
<i>Anaplasma phagocytophilum</i>	14 (33.3)
<i>Babesia microti</i>	10 (23.8)
<i>Ehrlichia canis</i>	6 (14.3)
<i>Leishmania infantum</i>	1 (2.4)
<i>Rickettsia felis</i>	1 (2.4)
<i>Rickettsia typhi</i>	0 (0)

Molecular results on ticks, fleas and feline blood samples

Almost all fleas (96.4 %), 19.2 % of ticks and 42.8 % of cats were found PCR-positive to at least one investigated pathogen. PCR results from ticks, fleas and cats are summarized in Table 2.

Bartonella clarridgeiae was confirmed by sequencing in seven cats (16.6 %), in 16 pools of fleas (57.1 %) and in two ticks (1.5 %). *Bartonella henselae* was confirmed by sequencing in nine cats (21.4 %) and in only four pools of fleas (14.3 %). DNA sequences were 99–100 % identical to both *Bartonella* species available in GenBank (*B. clarridgeiae* (GenBank ID: FN645454.1) and *B. henselae* (GenBank ID: KF466255.1). Similarly, *L. infantum* DNA was amplified from three cats, two fleas or pools and eight ticks.

Interestingly, DNA of hemoplasmas was not amplified from any of the ectoparasites but 11 feline blood samples were positive. Briefly, seven *Mhf*, seven *CMhm* and four *CMt* DNAs were detected in cats. *Babesia vogeli* was only amplified from one tick with a 100 % identity of GenBank sequences (GenBank ID: JX871885.1).

Rickettsia monacensis and *R. helvetica* were detected in five ticks and DNA sequences were 98–100 % identical to GenBank sequences (*R. monacensis* (GenBank ID: KF016136.1) and *R. helvetica* (GenBank ID: JQ796866.1). *Rickettsia felis* was found in 23 flea pools or single fleas with an identity of 100 % of GenBank sequences (GenBank ID: KF245441.1).

Six out of 25 engorged ticks were PCR positive to at least one investigated pathogen but they never harbored the same microorganisms of the host cat. Individual results of serology and PCR from cats and their ectoparasites are listed in Tables 3 and 4. In feline blood, the most frequent co-infection was among different hemoplasma species ($n = 6$). Moreover, co-infection was found between *Bartonella* spp. and hemoplasmas ($n = 3$). *Leishmania*

Table 2 Results of pathogens investigated by PCR in ticks, fleas and cats

Pathogens	Number of positive ticks, fleas or cats/ total number of ticks, fleas or cats (%)		
	Ticks	Fleas ^a	Feline blood
<i>Bartonella</i> spp.	2/73 (2.7)	20/28 (71.4)	16/42 (38.1)
Hemoplasmas	0 (0)	0 (0)	11/42 (26.2)
<i>Rickettsia</i> spp.	5/73 (6.8)	23/28 (82.1)	0 (0)
<i>Ehrlichia</i> spp./ <i>Anaplasma</i> spp.	0 (0)	NE	0 (0)
Piroplasmid	1/73 (1.4)	NE	0 (0)
<i>Hepatozoon felis</i>	0 (0)	NE	0 (0)
<i>Leishmania infantum</i>	8/73 (10.9)	2/28 (7.1)	3/42 (7.1)
Total (%) ^b	14/73 (19.2)	27/28 (96.4)	18/42 (42.8)

NE: not evaluated

^a10 single fleas and 18 flea pools (range 2–5 fleas/pool); ^bTotal number of specimens positive at least to one pathogen. Co-infections were counted only once

Table 3 Distribution of serological and PCR results in cats and in their ticks

Number of cats	Pathogens exposure confirmed by antibody detection in cats	Pathogens identified by real-time PCRs	
		Cat	Ticks
1	<i>R. conorii</i> , <i>B. microti</i>	None	None
2	<i>R. conorii</i> , <i>A. phagocytophilum</i>	None	None
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>E. canis</i> , <i>A. phagocytophilum</i>	<i>L. infantum</i>	None
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>A. phagocytophilum</i>	<i>B. henselae</i> , CMhm	None
1	<i>B. henselae</i> , <i>B. microti</i>	<i>B. henselae</i>	None
1	<i>R. conorii</i> , <i>A. phagocytophilum</i>	Mhf	None
1	<i>R. conorii</i> , <i>B. microti</i>	None	<i>L. infantum</i>
1	<i>B. henselae</i> , <i>B. microti</i>	<i>B. henselae</i>	<i>L. infantum</i>
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>B. microti</i>	CMhm, CMt	<i>B. clarridgeiae</i>
1 ^b	Negative	Mhf, CMhm	<i>R. monacensis</i>
1	<i>A. phagocytophilum</i>	<i>B. clarridgeiae</i>	<i>R. monacensis</i> , <i>L. infantum</i>
1	<i>B. microti</i>	None	<i>B. vogeli</i> , <i>L. infantum</i>
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>A. phagocytophilum</i>	<i>B. henselae</i> , Mhf, CMhm	<i>B. clarridgeiae</i> , <i>L. infantum</i>
1 ^a	<i>R. conorii</i> , <i>R. felis</i> , <i>E. canis</i> , <i>B. microti</i>	None	<i>R. helvetica</i> , <i>R. monacensis</i> , <i>L. infantum</i>

Abbreviations: Mhf *Mycoplasma haemofelis*, CMhm *Candidatus Mycoplasma haemominutum*, CMt *Candidatus Mycoplasma turicensis*
^aCat infested by 21 ticks; ^b Cat infested by both ticks and fleas

infantum DNA was also amplified in one cat positive to both *B. henselae* and *B. clarridgeiae*.

Molecular investigations detected DNA of different pathogens in single ticks. Briefly, *L. infantum* DNA was amplified in three ticks already positive to *B. clarridgeiae*, *R. monacensis* and *B. vogeli* and another tick was found positive to both *R. monacensis* and *R. helvetica*.

In fleas, the most frequent co-infection was between *R. felis* and *B. clarridgeiae* that was found in pools but also in one single flea. Only four fleas, collected from a PCR negative cat, were positive at the same time to three pathogens (*R. felis*, *B. clarridgeiae* and *L. infantum*).

The agreement between serological *Bartonella* results in cats and PCR data was moderate with ticks ($k = 0.461$; accuracy 0.79), followed by a fair agreement with feline blood ($k = 0.395$; accuracy 0.69) and with fleas ($k = 0.292$; accuracy 0.68). A fair or slight agreement was respectively observed between *B. henselae* PCR results in cats and in fleas ($k = 0.340$; accuracy 0.82) as well as for *B. clarridgeiae* ($k = 0.208$; accuracy 0.57). Agreement between serological and molecular results for other pathogens in individual cats and their ectoparasites was slight or not found.

Table 4 Distribution of serological and PCR results in cats and in their fleas

Number of cats	Pathogens exposure confirmed by antibody detection in cats	Pathogens identified by real-time PCRs	
		Cats	Fleas
1	<i>B. henselae</i>	None	None
1	<i>B. henselae</i>	None	<i>B. henselae</i>
1	Negative	Mhf, <i>B. clarridgeiae</i>	<i>B. clarridgeiae</i>
2	<i>B. henselae</i>	<i>B. henselae</i>	<i>B. clarridgeiae</i>
1	Negative	None	<i>R. felis</i>
1 ^a	Negative	Mhf, CMhm	<i>R. felis</i>
1	<i>R. conorii</i>	None	<i>R. felis</i>
1	<i>B. henselae</i>	CMhm, CMt	<i>R. felis</i>
1	<i>B. henselae</i> , <i>R. conorii</i>	CMhm, CMt	<i>R. felis</i>
1	<i>R. conorii</i> , <i>A. phagocytophilum</i>	None	<i>R. felis</i>
1	<i>B. henselae</i> , <i>R. conorii</i>	<i>B. henselae</i> , <i>L. infantum</i>	<i>B. henselae</i> , <i>R. felis</i>
1	<i>B. henselae</i>	<i>B. clarridgeiae</i>	<i>B. henselae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>A. phagocytophilum</i>	<i>B. henselae</i>	<i>B. henselae</i> , <i>R. felis</i>
2	<i>R. conorii</i>	None	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>A. phagocytophilum</i>	None	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>E. canis</i> , <i>B. microti</i> , <i>L. infantum</i>	None	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>B. microti</i>	Mhf, CMhm, CMt	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>R. conorii</i>	Mhf	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>R. conorii</i>	<i>B. henselae</i>	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>E. canis</i>	<i>B. clarridgeiae</i>	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>E. canis</i> , <i>A. phagocytophilum</i>	<i>B. clarridgeiae</i>	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i> , <i>B. microti</i>	<i>B. clarridgeiae</i>	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>B. henselae</i>	Mhf	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>A. phagocytophilum</i>	<i>B. clarridgeiae</i> , <i>L. infantum</i>	<i>B. clarridgeiae</i> , <i>R. felis</i>
1	<i>R. conorii</i> , <i>A. phagocytophilum</i>	None	<i>R. felis</i> , <i>L. Infantum</i>
1	<i>B. henselae</i> , <i>R. conorii</i> , <i>E. canis</i> , <i>A. phagocytophilum</i>	None	<i>B. clarridgeiae</i> , <i>R. felis</i> , <i>L. Infantum</i>

Abbreviations: Mhf *Mycoplasma hemofelis*, CMhm *Candidatus Mycoplasma haemominutum*, CMt *Candidatus Mycoplasma turicensis*
^aCat infested by both ticks and fleas

Discussion

In this study, we confirmed *C. felis* as the unique flea species found in cats from Southern Italy mainly in Calabria Province. In contrast, species of *Rhipicephalus* and *Ixodes* were found as the only tick species collected from cats mainly living in Lipari Island (Sicily) [4]. Ecological factors, season of sampling, climatic variations may be responsible for these findings. Moreover, outdoor cats from Lipari Island are free roaming in a wild habitat and therefore they are in close contact with wild rabbits and birds and their ectoparasites.

Almost all fleas (96.4 %), 19.2 % of ticks and 42.8 % of cats were found PCR positive to at least one investigated pathogen. The most common pathogens identified by molecular techniques were of zoonotic concern and include *Rickettsia*, *Bartonella* and *L. infantum*, however with different distribution and rate of infection in cats and their ectoparasites. For instance, *Rickettsia* spp. DNA was exclusively amplified from the ectoparasites (*R. helvetica* and *R. monacensis* from ticks and *R. felis* from fleas) supporting a possible role for cofeeding transmission in the maintenance of these pathogens within the vector population as already demonstrated for *R. conorii israelensis* in *R. sanguineus* ticks [17] and for *R. felis* in fleas (*C. felis* and *Xenopsilla cheopis*) [18]. These rickettsial species can cause febrile illness among other clinical manifestations in humans as well established for *R. felis* [19] and also described for *R. helvetica* and *R. monacensis* [20]. Therefore, it is important to highlight that cats will be carriers of ectoparasites and associated *Rickettsia* species to humans suggesting a zoonotic potential but they do not appear to be reservoirs of these infections. Conversely, *Bartonella* DNA (*B. clarridgeiae* and *B. henselae*) was the most frequent pathogen found in cats suggesting an important zoonotic risk to humans [21] as carriers of ectoparasites and apparent reservoirs for both infections [22]. Moreover, hemoplasma (*Mhf*, CMt, CMhm) DNA was detected only in cat blood confirming the potential limited role of vectors in their transmission [23, 24] despite DNA of some hemotropic mycoplasmas can be found in ectoparasites collected from cats [11, 25, 26].

It is noteworthy that *L. infantum* DNA was found in 7–10 % of ectoparasite or cat blood samples and it was the most common parasite found in ticks. Interestingly, this is the first *bona fide* report of *L. infantum* DNA from cat fleas. DNA from *Bartonella henselae*, *B. clarridgeiae*, *R. monacensis*, *R. helvetica*, *R. felis*, *M. haemofelis*, *Ca. M. haemominutum*, *Ca. M. turicensis*, *B. vogeli* and *L. infantum* was amplified in feline blood and/or in ectoparasites. Vector-borne pathogens found in this study partly confirm previous data reported in Italy from cats or their ectoparasites [8–10, 27, 28]. However, we obtained data at the same time from cats and the ticks and fleas they harbor.

Detection of antibodies against most of the investigated VBPs and/or of circulating pathogen DNA showed that cats harboring ticks or fleas are frequently exposed to VBPs in the geographic area under study. In fact as much as half of cats had antibodies against *R. conorii* and *B. henselae* and the percentage of detectable antibodies against *A. phagocytophilum*, *B. microti* and *E. canis* was not negligible confirming data previously reported in Italy [10] and throughout Europe [6, 29–31]. Interestingly, in the present study, a very low *R. felis* antibody rate was observed (2.4 %) as well as no detection of *R. felis* DNA in any feline blood samples as reported in other studies [26, 32, 33]. Our findings are in disagreement with other data that reported higher *R. felis* antibody rates (16.3 %) in Spain [33] as well as *R. felis* DNA detection (28 %) in feline blood from Bangladesh cats [34]. In the present study, high *R. conorii* antibody rates were found in the absence of rickettsiemia as previously reported in other studies in cats [29]. Similar findings are also observed in dogs with high *R. conorii* antibody rates and low rickettsiemia in endemic areas [35]. However, the present findings are in disagreement with a study performed in northeastern Spain where *Rickettsia* DNA similar to *R. conorii* or *R. massiliae* was found in 10 % of cats [36]. Obviously, we cannot exclude that infections caused by other *Rickettsia* species of the spotted fever group circulating in southern Italy such as *R. massiliae* among others, contributed to this high antibody prevalence [36, 37]. It is well known that cross-reactions are common among *Rickettsia* species and this is a limitation of antibody prevalence studies [38]. Sicily and Calabria are Italian regions with a high incidence of human rickettsial diseases [39] and it is of peculiar interest to clarify the role of cats in their eco-epidemiology. In contrast with Spanish data [40], no serological and molecular traces of *R. typhi* were observed in both cats and ectoparasites.

Agreement between the majority of pathogens based on molecular or antibody detection among cats and in their ectoparasites was not found. The only exception was for fleas which were DNA positive to *Bartonella* (four for *B. henselae* and five to *B. clarridgeiae*). These fleas were collected from cats which were antibody and/or PCR positive to the same pathogen. Lappin et al. reported that almost all cats (94.7 %) infected by *B. clarridgeiae* carried *B. clarridgeiae* infected fleas and they suggested that *C. felis* may be a vector for this pathogen [11]. Conversely, in Taiwan, researchers found a high prevalence of fleas PCR positive for *B. clarridgeiae* and a very low prevalence of this bacterium in cats. They raised the hypothesis that *B. clarridgeiae* is more adapted to the flea than to the vertebrate host [5]. Other studies found a higher prevalence of *Bartonella* and hemoplasma DNA in fleas than in cat blood, but no data were given about positivity of fleas and

their corresponding cat host [31, 41]. In this study, *B. clarridgeiae* was the microorganism most frequently found simultaneously in fleas and their feline host.

To the best of our knowledge, no studies compared the molecular detection of pathogens in ticks removed from cats and their host. Host molecular negativity for pathogens found in ectoparasites may depend on the vector competence. For tick-borne pathogens, the transmission occurs at different times after the beginning of the blood meal depending upon specific life cycle characteristics [24]. Moreover, the bacteremia of some VBPs is transient, lasting a few hours, as reported for *Rickettsia* spp. and *E. canis* in dogs. This makes it difficult to detect the pathogens in the blood stream [24]. The low level of circulating pathogens requires highly sensitive molecular tools.

Conclusions

In conclusion, cats harboring ticks and fleas are frequently exposed to many VBPs. However, the simultaneous detection of VBPs in the hosts and their ectoparasites is uncommon with the exception of *Bartonella*. *Bartonella clarridgeiae* followed by *B. henselae* were the species most frequently found at the same time in fleas and the cat host.

Ticks and fleas harbored by cats frequently carry pathogens of zoonotic concern. As a preventative measure, the appropriate use of ectoparasiticide treatments is strongly recommended for use in cats.

Abbreviations

°C: Celsius degree; Bp: base pairs; DNA: deoxyribonucleic acid; EDTA: ethylenediamine tetraacetic acid; IFAT: immunofluorescence antibody test; IgG: Immunoglobulins G; k: k agreement; min: minutes; n: number; OR: Odds Ratio; PBS: Phosphate buffered saline; PCR: Polymerase Chain Reaction; VBPs: Vector-Borne Pathogens; μ L: microliters.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MGP and LSG conceived the research study. MFP worked in the field. MFP contributed with data analysis and interpretation, wrote the first draft and revised the manuscript. MFP performed identification of ectoparasites and laboratory techniques. MFP, MGP and MM performed serological analysis. LA, LS and SR supervised the performance of molecular techniques. LSG and MGP contributed with data analysis and interpretation and wrote and revised the manuscript. All authors read and approved the final version of the manuscript.

Acknowledgements

The authors are grateful to Angela Burrascano for technical collaboration for serological techniques and to Tatiana Proboste for helping with morphological identification of ticks. The authors also thank Dr. Shazia Hosein for English revision of the manuscript.

This clinical study was funded by Bayer Animal Health Care-Animal Health Division (Monheim, Germany). Publication fees of this manuscript have been sponsored by Bayer HealthCare - Animal Health division, in the framework of the 11th CVBD World Forum Symposium.

Author details

¹Istituto Zooprofilattico Sperimentale della Sicilia Adelmo Mirri, Via G. Marinuzzi 3, 90129 Palermo, Italy. ²Departament de Medicina i Cirurgia

Animal, Facultat de Veterinària, Universitat Autònoma de Barcelona, 08193, Bellaterra, Barcelona, Spain. ³Vetgenomics, Edifici Eureka, PRUAB, 08193, Bellaterra, Barcelona, Spain. ⁴Dipartimento di Scienze Veterinarie, Università di Messina, Polo Universitario Annunziata, 98168 Messina, Italy.

Received: 14 March 2016 Accepted: 25 April 2016

Published online: 10 May 2016

References

- Dantas-Torres F, Otranto D. Dogs, cats, parasites, and humans in Brazil: opening the black box. *Parasit Vectors*. 2014;7:22.
- Otranto D, Dantas-Torres F. Canine and feline vector-borne diseases in Italy: current situation and perspectives. *Parasit Vectors*. 2010;3:2.
- Maggi RG, Ericson M, Mascarelli PE, Bradley JM, Breitschwerdt EB. *Bartonella henselae* bacteremia in a mother and son potentially associated with tick exposure. *Parasit Vectors*. 2013;6:101.
- Pennisi M-G, Persichetti M-F, Serrano L, Altet L, Reale S, Gulotta L, et al. Ticks and associated pathogens collected from cats in Sicily and Calabria (Italy). *Parasit Vectors*. 2015;8:512.
- Tsai Y-L, Lin C-C, Chomel BB, Chuang S-T, Tsai K-H, Wu W-J, et al. *Bartonella* infection in shelter cats and dogs and their ectoparasites. *Vector Borne Zoonotic Dis*. 2011;11:1023–30.
- Ayllón T, Diniz PPVP, Breitschwerdt EB, Villaescusa A, Rodríguez-Franco F, Sainz A. Vector-borne diseases in client-owned and stray cats from Madrid, Spain. *Vector Borne Zoonotic Dis*. 2012;12:143–50.
- Vilhena H, Martinez-Díaz VL, Cardoso L, Vieira L, Altet L, Francino O, et al. Feline vector-borne pathogens in the north and centre of Portugal. *Parasit Vectors*. 2013;6:99.
- Spada E, Proverbio D, Galluzzo P, Della Pepa A, Perego R, Bagnagatti De Giorgi G, et al. Molecular study on selected vector-borne infections in urban stray colony cats in northern Italy. *J Feline Med Surg*. 2014;16:684–8.
- Spada E, Proverbio D, Galluzzo P, Della Pepa A, Bagnagatti De Giorgi G, Perego R, et al. Prevalence of haemoplasma infections in stray cats in northern Italy. *ISRN Microbiol*. 2014;2014:298352.
- Spada E, Proverbio D, Galluzzo P, Perego R, Bagnagatti De Giorgi G, Roggero N, et al. Frequency of piroplasms *Babesia microti* and *Cytauxzoon felis* in stray cats from northern Italy. *BioMed Res Int*. 2014;2014:943754.
- Lappin MR, Griffin B, Brunt J, Riley A, Burney D, Hawley J, et al. Prevalence of *Bartonella* species, haemoplasma species, *Ehrlichia* species, *Anaplasma phagocytophilum*, and *Neorickettsia risticii* DNA in the blood of cats and their fleas in the United States. *J Feline Med Surg*. 2006;8:85–90.
- Pennisi MG, Lupo T, Malara D, Masucci M, Miglizzo A, Lombardo G. Serological and molecular prevalence of *Leishmania infantum* infection in cats from Southern Italy. *J Feline Med Surg*. 2012;14:656–7.
- Ménier K, Beaucournu JC. Taxonomic study of the genus *Ctenocephalides* Stiles & Collins, 1930 (Insecta: Siphonaptera: Pulicidae) by using aedeagus characters. *J Med Entomol*. 1998;35:883–90.
- Manilla G. Acari: Ixodida. Vol. XXXVI di Fauna D'Italia. Bologna: Ed. Calderini; 1998.
- Martinez-Díaz VL, Silvestre-Ferreira AC, Vilhena H, Pastor J, Francino O, Altet L. Prevalence and co-infection of haemotropic mycoplasmas in Portuguese cats by real-time polymerase chain reaction. *J Feline Med Surg*. 2013;15: 879–85.
- Vitale F, Caracappa S, Manna L, Gravino A, Reale S. *Leishmania infantum* minicircle DNA sequencing analysis and diagnosis by PCR. In: Proceedings of the Second World Congress on Leishmaniasis (WL2). Crete; 2001.
- Zemtsova G, Killmaster LF, Mumcuoglu KY, Levin ML. Co-feeding as a route for transmission of *Rickettsia conorii israelensis* between *Rhipicephalus sanguineus* ticks. *Exp Appl Acarol*. 2010;52:383–92.
- Brown LD, Christofferson RC, Banajee KH, Del Piero F, Foil LD, Macaluso KR. Cofeeding intra- and interspecific transmission of an emerging insect-borne rickettsial pathogen. *Mol Ecol*. 2015;24:5475–89.
- Parola P. *Rickettsia felis*: from a rare disease in the USA to a common cause of fever in sub-Saharan Africa. *Clin Microbiol Infect Off Publ Eur Soc Clin Microbiol Infect Dis*. 2011;17:996–1000.
- Oteo JA, Portillo A. Tick-borne rickettsioses in Europe. *Ticks Tick-Borne Dis*. 2012;3:271–8.
- Vayssier-Taussat M, Moutailler S, Féménia F, Raymond P, Croce O, La Scola B, et al. Identification of Novel Zoonotic Activity of *Bartonella* spp., France. *Emerg Infect Dis*. 2016;22:457–62.

22. Chomel BB, Kasten RW. Bartonellosis, an increasingly recognized zoonosis. *J Appl Microbiol.* 2010;109:743–50.
23. Barker E, Tasker S. Haemoplasmas: lessons learnt from cats. *N Z Vet J.* 2013; 61:184–92.
24. Greene CE. Infectious diseases of the dog and cat. Bologna: Elsevier Health Sciences; 2013. p.1383.
25. Willi B, Boretti FS, Meli ML, Bernasconi MV, Casati S, Hegglin D, et al. Real-time PCR investigation of potential vectors, reservoirs, and shedding patterns of feline hemotropic mycoplasmas. *Appl Environ Microbiol.* 2007;73:3798–802.
26. Assarasakorn S, Veir JK, Hawley JR, Brewer MM, Morris AK, Hill AE, et al. Prevalence of *Bartonella* species, hemoplasmas, and *Rickettsia felis* DNA in blood and fleas of cats in Bangkok. *Thailand Res Vet Sci.* 2012;93:1213–6.
27. Salvatore D, Aureli S, Baldelli R, Di Francesco A, Tampieri MP, Galuppi R. Molecular evidence of *Leishmania infantum* in *Ixodes ricinus* ticks from dogs and cats, in Italy. *Vet Ital.* 2014;50:307–12.
28. Giudice E, Di Pietro S, Alaimo A, Blanda V, Lelli R, Francaviglia F, et al. A molecular survey of *Rickettsia felis* in fleas from cats and dogs in Sicily (Southern Italy). *PLoS One.* 2014;9, e106820.
29. Solano-Gallego L, Hegarty B, Espada Y, Llull J, Breitschwerdt E. Serological and molecular evidence of exposure to arthropod-borne organisms in cats from northeastern Spain. *Vet Microbiol.* 2006;118:274–7.
30. Alves AS, Milhano N, Santos-Silva M, Santos AS, Vilhena M, de Sousa R. Evidence of *Bartonella* spp., *Rickettsia* spp. and *Anaplasma phagocytophilum* in domestic, shelter and stray cat blood and fleas, Portugal. *Clin Microbiol Infect Off Publ Eur Soc Clin Microbiol Infect Dis.* 2009;15(2):1–3.
31. Barrs VR, Beatty JA, Wilson BJ, Evans N, Gowan R, Baral RM, et al. Prevalence of *Bartonella* species, *Rickettsia felis*, haemoplasmas and the *Ehrlichia* group in the blood of cats and fleas in eastern Australia. *Aust Vet J.* 2010;88:160–5.
32. Millán J, Proboste T, Fernández de Mera IG, Chirife AD, de la Fuente J, Altet L. Molecular detection of vector-borne pathogens in wild and domestic carnivores and their ticks at the human-wildlife interface. *Ticks Tick-Borne Dis.* 2016;7:284–90.
33. Gracia MJ, Marcén JM, Pinal R, Calvete C, Rodes D. Prevalence of *Rickettsia* and *Bartonella* species in Spanish cats and their fleas. *J Vector Ecol J Soc Vector Ecol.* 2015;40:233–9.
34. Ahmed R, Paul SK, Hossain MA, Ahmed S, Mahmud MC, Nasreen SA, et al. Molecular detection of *Rickettsia felis* in humans, cats, and cat fleas in Bangladesh, 2013–2014. *Vector Borne Zoonotic Dis.* 2016;16:356–8.
35. Solano-Gallego L, Capri A, Pennisi MG, Caldin M, Furlanello T, Trotta M. Acute febrile illness is associated with *Rickettsia* spp. infection in dogs. *Parasit Vectors.* 2015;8:216.
36. Segura F, Pons I, Miret J, Pla J, Ortuño A, Nogueras M-M. The role of cats in the eco-epidemiology of spotted fever group diseases. *Parasit Vectors.* 2014; 7:353.
37. Otranto D, Dantas-Torres F, Giannelli A, Latrofa MS, Cascio A, Cazzin S, et al. Ticks infesting humans in Italy and associated pathogens. *Parasit Vectors.* 2014;7:328.
38. Wachter M, Wolfel S, Pfeffer M, Dobler G, Kohn B, Moritz A, et al. Serological differentiation of antibodies against *Rickettsia helvetica*, *R. raoultii*, *R. slovaca*, *R. monacensis* and *R. felis* in dogs from Germany by a micro-immunofluorescent antibody test. *Parasit Vectors.* 2015;8:126.
39. Ciceroni L, Pinto A, Ciarrocchi S, Ciervo A. Current knowledge of rickettsial diseases in Italy. *Ann N Y Acad Sci.* 2006;1078:143–9.
40. Nogueras MM, Pons I, Ortuño A, Miret J, Pla J, Castellà J, et al. Molecular detection of *Rickettsia typhi* in cats and fleas. *PLoS One.* 2013;8:e71386.
41. Kamrani A, Parreira VR, Greenwood J, Prescott JF. The prevalence of *Bartonella*, hemoplasma, and *Rickettsia felis* infections in domestic cats and in cat fleas in Ontario. *Can J Vet Res Rev Can Rech Vét.* 2008;72:411–9.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

