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Acromegaly, genetic variants of the aryl hydrocarbon receptor pathway and environmental burden

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AHR: Aryl Hydrocarbon Receptor AIP: AHR-Interacting Protein

1 ACROMEGALY, GENETIC VARIANTS OF THE ARYL HYDROCARBON RECEPTOR

2 PATHWAY AND ENVIRONMENTAL BURDEN

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49 ABSTRACT

Increasing evidence suggests that environmental contaminants can exert endocrine disruptors activities and that pollution exposition can have a role in tumorigenic processes. Several environmental pollutants have been shown to affect pituitary cells biology and function. The aryl hydrocarbon receptor (AHR) pathway is involved in xenobiotics' metabolism and in tumorigenesis. A deregulation of the AHR pathway could have a role in pituitary tumours' pathophysiology, especially in the GH secreting ones. AHR-interacting protein (AIP) is one of the key partners of AHR and is implicated in pituitary tumours' pathogenesis. Moreover, an increased prevalence of acromegaly has been reported in a highly polluted area of the province of Messina (Sicily, Italy). Nevertheless, at present, few data are available about the potential role of environmental factors in the pathogenesis and clinical expression of GH secreting pituitary tumours. This review is aimed at discussing the evidences on the potential links among environmental pollutants, the AHR pathway and the pathophysiology of GH-secreting pituitary adenomas.

Key terms: acromegaly, AHR, AIP, pollution, endocrine disruptors

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73 Introduction

74 Acromegaly is a rare disease characterised by an abnormal growth of bone, soft tissues and organs, 75 as a consequence of a growth hormone (GH) excess due – in most of the cases – to a pituitary 76 adenoma (Broder, Chang, Cherepanov et al., 2016; Capatina and Wass, 2015; Dal, Feldt-77 Rasmussen, Andersen et al., 2016; Daly, Rixhon, Adam et al., 2006; Melmed, 2009; Mestron, 78 Webb, Astorga et al., 2004). The estimated prevalence of acromegaly has been variably reported 79 between 34-125 cases per million of persons (cpm), although it has been shown to be even higher in 80 some studies (Cannavo, Ferrau, Ragonese et al., 2010). Acromegaly is generally diagnosed after 81 several years from disease onset with considerable clinical, economical and social consequences 82 (Capatina and Wass, 2015). To date, the pathogenic mechanisms underlying the development, 83 progression and clinical impact of GH-secreting pituitary tumours are only partially disclosed.

84 Some studies have shown that environmental pollutants with endocrine disruptors activities can 85 impact on pituitary function and biology (Gore, 2010). It's known that the aryl hydrocarbon 86 receptor (AHR) is the main actor of the intracellular mechanisms of xenobiotics' metabolism and 87 detoxification, and can influence the major stages of tumorigenesis (Figure 1) (Dietrich and Kaina, 88 2010; Feng, Cao and Wang, 2013; Hao and Whitelaw, 2013). However, data on the role of 89 environmental pollutants and of the alterations of the AHR pathway in pituitary tumorigenesis are 90 scanty. On the other hand, the AHR cytosolic stabilization, function and signalling pathway are 91 strictly dependent on the AHR-interacting protein (AIP) that in turn has a key role in pituitary 92 tumour development (Beckers, Aaltonen, Daly et al., 2013; Beischlag, Luis Morales, Hollingshead 93 et al., 2008; Hao and Whitelaw, 2013; Petrulis and Perdew, 2002; Trivellin and Korbonits, 2011). 94 Indeed, AIP gene mutations have been found in about 20% of patients with familial isolated 95 pituitary adenoma syndrome (FIPA) and in 40% of patients with isolated familial 96 somatotropinomas, as well as in up to 4% of patients with apparently sporadic acromegaly (Beckers 97 et al., 2013; Ferrau, Romeo, Puglisi et al., 2016; Hernandez-Ramirez, Gabrovska, Denes et al., 98 2015; Lloyd and Grossman, 2014). Pituitary tumours associated with AIP gene mutations are

99 mainly GH and/or PRL secreting, show an aggressive clinical and biochemical phenotype, occur 100 more frequently in young patients and are more resistant to conventional treatments (Alband and 101 Korbonits, 2014; Beckers et al., 2013; Martucci, Trivellin and Korbonits, 2012). In this review, the 102 evidence on the potential links among environmental pollutants, the AHR pathway and the 103 pathophysiology of GH-secreting pituitary adenomas will be discussed.

104

105 The AHR-AIP pathway

106 The AHR is a transcription factor belonging to the basic helix-loop-helix/Per/ARNT/Sim (PAS) 107 family (Dietrich and Kaina, 2010). It is stimulated by several natural compounds which are present 108 in food, as indoles and flavonoids, or by tryptophan derivatives, arachidonic acid metabolites and 109 other endogenous products (Denison and Nagy, 2003; Denison, Pandini, Nagy et al., 2002; Dietrich 110 and Kaina, 2010). The most potent AHR ligand known so far is the 2,3,7,8-tetrachlorodibenzo-p-111 dioxin (TCDD) but more than 400 exogenous compounds act as AHR ligands, most of which are 112 environmental endocrine disruptors, including a variety of polycyclic aromatic hydrocarbons and 113 planar polychlorinated biphenyls (PCBs) (Denison and Nagy, 2003; Denison et al., 2002; Dietrich 114 and Kaina, 2010).

115 It is generally accepted that the metabolic responses to environmental pollutants can be the direct 116 consequence of AHR activation. In the cytosol, the unbound receptor forms a complex with AIP, 117 the co-chaperone protein p23 and two heat shock protein 90 (Hsp90) molecules. Ligand binding 118 results in nuclear translocation of AHR, dissociation from the chaperone proteins, 119 heterodimerization with a nuclear translocator (ARNT) and subsequent binding to xenobiotic-120 responsive elements (XREs) (Figure 1). This leads to transactivation of several genes encoding 121 phase I and II xenobiotic metabolizing enzymes, such as cytochrome P450s (CYP1A1, CYP1A2 122 and CYP1B1), as well as other genes coding for non-enzymatic molecules or proteins like the 123 cyclin-dipendent kinases inhibitor p27Kip1 (Denison and Nagy, 2003; Dietrich and Kaina, 2010; 124 Feng et al., 2013; Hao and Whitelaw, 2013). The functional relevance of the AHR pathway is not

restricted to the cellular response to the toxic insult, since others 'non-canonical' effects (on the cell-cycle, contact inhibition, cell adhesion, function and metabolism of the oestrogen receptors, and DNA repairing processes) can be the direct consequence of the activation of this transcriptional factor, further contributing to the disruption of cell homeostasis (Denison and Nagy, 2003; Dietrich and Kaina, 2010; Feng et al., 2013).

The AHR plays a relevant role in tissue-specific embryonic development, hematopoietic stem cell self-renewal, pluripotent stem cell and neural stem cell differentiation, and erythroid stem cell growth, as well as in development of cells with cancer stem cell-like qualities (Gasiewicz, Singh and Bennett, 2014; Mulero-Navarro and Fernandez-Salguero, 2016; Stanford, Wang, Novikov et al., 2016). The AHR interacts directly and affects the activity of Sox2, a master regulator of selfrenewal that is also required for pituitary progenitor proliferation (Goldsmith, Lovell-Badge and Rizzoti, 2016; Stanford et al., 2016).

137 The AHR is thought to be involved in tumorigenic processes, although the molecular mechanism 138 underlying the AHR-related carcinogenesis is largely unknown and probably species and tissue 139 specific (Feng et al., 2013; Murray, Patterson and Perdew, 2014). Abnormal AHR expression 140 and/or activity have been shown in a variety of sporadic human cancers (Dietrich and Kaina, 2010; 141 Feng et al., 2013; Harper, Riddick and Okey, 2006). Some studies suggested that AHR is also 142 implicated in the transition from a benign to a malignant tumour, and that the AHR pathway 143 activation associates with increased tumour invasiveness (Villano, Murphy, Akintobi et al., 2006; 144 Yang, Liu, Murray et al., 2005). Moreover, the pattern of regulation of the AHR gene has been 145 identified as one of the most altered in a recent study aimed to identify the driver genes in colorectal 146 cancer (Aziz, Periyasamy, Al Yousef et al., 2014). Among the endocrine tumours, AHR has been 147 found overexpressed in papillary thyroid carcinomas of patients with and without acromegaly, 148 especially in tumour samples carrying the BRAF V600E mutation (Mian, Ceccato, Barollo et al., 149 2014).

150 AIP is one of the main partners of AHR, being crucial for its cytoplasmic stabilization and function, 151 and both of them are expressed in the pituitary (Denison et al., 2002; Trivellin and Korbonits, 152 2011). AIP is a co-chaperone protein that interacts with several other proteins such as nuclear 153 receptors (peroxisome proliferator activated receptor alpha, estrogen receptor alpha, thyroid 154 receptor beta1 and glucocorticoid receptor), oncogenes and components of the cAMP-signalling 155 pathway (Trivellin and Korbonits, 2011). AIP gene mutations have been described in families with 156 cases of isolated pituitary adenomas, predominantly presenting as GH and/or PRL secreting 157 tumours (Beckers et al., 2013; Vierimaa, Georgitsi, Lehtonen et al., 2006). AIP gene mutations have 158 been also found in young subjects with sporadic pituitary adenomas presenting with aggressive 159 disease and less responsive to conventional medical treatment (Cazabat, Bouligand, Salenave et al., 160 2012; Daly, Tichomirowa, Petrossians et al., 2010; Lloyd and Grossman, 2014). AIP has been 161 suggested to act as a tumour suppressor in pituitary cells (Leontiou, Gueorguiev, van der Spuy et 162 al., 2008). The precise tumorigenic mechanism is not well understood but some recent data point to 163 a deregulation of the cAMP pathway. Indeed, it has been demonstrated that AIP deficiency leads to 164 elevated cAMP concentrations through defective inhibitory G protein α subunits (G α)i-2 and G α i-3 165 that normally inhibit cAMP synthesis (Tuominen, Heliovaara, Raitila et al., 2015). The same authors demonstrated, by immunostaining, that AIP deficiency is associated with a reduction in 166 167 Gai-2 protein expression in human and mouse GH-secreting pituitary adenomas, thus indicating 168 defective Gai signalling in these tumours (Tuominen et al., 2015). Moreover, a recent study showed 169 a reduced half-life of AIP mutants, due to enhanced proteosomal degradation, that would correlate 170 with the clinical phenotype (Hernandez-Ramirez, Martucci, Morgan et al., 2016).

Some studies suggested that AIP gene mutations could impact on AHR signalling pathway (Nukaya, Lin, Glover et al., 2010; Trivellin and Korbonits, 2011). Indeed, AIP gene mutations, known to be pathogenic and affecting the C-terminal region of the protein, have the potential to disrupt the AHR and phosphodiesterase (PDE)4A5 client-protein interaction (Morgan, Hernandez-Ramirez, Trivellin et al., 2012). Among more than 20 interactions of AIP, that one with the AHR-

176 cAMP-PDE signalling pathway seems to be the most promising in terms of a potential involvement 177 in pituitary tumorigenesis (Dietrich and Kaina, 2010). On the other hand, the cAMP pathway has 178 been shown to be relevant in GH-secreting pituitary cells' biology and is a target of the action of 179 somatostatin analogues (SSA) that are widely used to control acromegaly (Melmed, 2003). 180 Moreover, cAMP is a non-ligand activator of AHR and nucleocytoplasmic shuttling of AHR 181 induced by cAMP is inhibited by PDE2, which in turn is stabilised by AIP. Furthermore, AIP has 182 been demonstrated to reduce forskolin-induced cAMP signalling in GH3 cells (GH and PRL 183 secreting rat pituitary tumour cells) (de Oliveira, Hoffmeister, Gambaryan et al., 2007; Formosa, 184 Xuereb-Anastasi and Vassallo, 2013; Oesch-Bartlomowicz, Huelster, Wiss et al., 2005).

185 However, the AIP effects on AHR are still a matter of debate. Some authors showed that AIP could 186 increase the transcriptional activity and expression of AHR, while others suggested the opposite 187 (Kazlauskas, Poellinger and Pongratz, 2000). Some evidence suggests that AIP prevent AHR from ubiquitinous-dipendent proteosomal degradation (Kazlauskas et al., 2000; Morales and Perdew, 188 189 2007; Trivellin and Korbonits, 2011). Recently, Lecoq A. et al. showed that AIP protein level in 190 fibroblasts from AIP mutated patients was reduced as compared to healthy subjects, without any 191 significant change in terms of AHR expression (Lecoq, Viengchareun, Hage et al., 2016). However, 192 in the same study, gene expression analysis showed significant alterations in the expression of the 193 AHR target genes CYP1B1 and AHRR in AIP-mutated fibroblasts, both before and after 194 stimulation with the endogenous AHR ligand kynurenine. AHR activation increased CYP1B1 195 expression to a greater extent in GH3 cells overexpressing wild type AIP than in cells expressing 196 mutant AIP. Furthermore, AHR related CYP1B1 induction was reduced in AIP-knockdown GH3 197 cells as well as the reduced expression of AIP affected the kynurenine-dependent GH secretion of 198 GH3 cells. This data would suggest an impairment of AHR transcriptional activity in AIP-mutated 199 cells (Lecoq et al., 2016).

200 Moreover, there are several functional cross talks between the AHR-AIP pathway and other 201 intracellular signalling pathways that in turn could mediate the effects of environmental pollutants

202 and could be also implicated in the tumorigenic process. For instance, AHR is involved in androgen 203 and oestrogen receptors (ER) function, promoting the ubiquitination and proteosomal degradation 204 and modulating the actions of sex steroids. Moreover, the AHR/ARNT complex antagonizes the 205 estrogenic action by inhibiting competitively the binding of ERalfa with the oestrogen responsive 206 elements when close to or overlapping the XRE (Klinge, Bowers, Kulakosky et al., 1999; Shanle 207 and Xu, 2011). The interaction among AHR and ER and AR would modulate in a positive or 208 negative fashion the oestrogen/androgen signalling pathways according to the cellular context. In 209 this regard, it's noteworthy that several endocrine disruptors can interfere with the oestrogen 210 receptor signalling in GH3 cells (Kim, Jung, Choi et al., 2012; Vo, An, Yang et al., 2012; Wang, 211 Knosp, Tai et al., 2014).

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213 The AHR pathway in GH-secreting pituitary tumours

214 In GH-secreting pituitary adenomas, lower AIP expression, as detected by immunohistochemistry, 215 has been shown to associate with a more aggressive disease, since it correlated with higher pre-216 operative GH and IGF-1 levels, more invasive pituitary tumours and higher Ki67 index (Jaffrain-217 Rea, Angelini, Gargano et al., 2009; Kasuki, Colli, Elias et al., 2012; Kasuki, Vieira Neto, 218 Wildemberg et al., 2012). Nevertheless, the expression of AHR and its partners in pituitary tumours 219 is more controversial. It's worth of noting, however, that AHR expression and signalling could be 220 differently affected according to pituitary tumour phenotype, in line with the postulated cell and 221 tissue specific actions of AHR. In 2009, a study from Jaffrain-Rea et al. showed a lower expression 222 of AIP and AHR in invasive somatotropinomas compared to non-invasive ones, suggesting that the 223 down-regulation of both could be associated to disease aggressiveness (Jaffrain-Rea et al., 2009). 224 The same group in a following study found some AHR cytoplasmic (AHRc) immunostaining in 225 94% of cases and AHR nuclear (AHRn) staining in 49% of samples, with a positive correlation 226 between the AHRc and AHRn and between AIP and AHRc, which would suggest a role for AIP in 227 stabilising AHR. The GH-secreting adenomas with higher AHR expression (32% of the analysed

228 cases) were smaller than the other ones and included a higher percentage of pure GH-secreting 229 tumours as compared to those expressing AHR at low levels. Moreover, the AHR content or 230 localization was not statistically influenced by preoperative SSA treatment, excluding a relevant 231 role for AHR in the pharmacological response to SSA. However, SSA treatment was able to reduce 232 the correlation between AHRc and AIP content, suggesting different effects of these drugs on the 233 expression of the two proteins or a destabilization of their complex (Jaffrain-Rea, Rotondi, Turchi 234 et al., 2013). The same authors found that Gsp status could have some effects on AHR expression or localization. Indeed, AHR nuclear content was reduced in Gsp^{+ve} tumours, probably due to a 235 236 reduced cAMP concentration with consequent inhibited AHR shuttling, while pre-operative SSA 237 treatment was able to increase AHR expression in Gsp^{-ve} cells. However, octreotide treatment of 238 primary cell cultures of human somatotropinomas did not increase AHR expression, regardless of 239 Gsp status (Jaffrain-Rea et al., 2013).

240 On the other hand, Heliovaara et al. found ARNT to be underexpressed and AHR nuclear content to 241 be increased in AIP mutated pituitary adenomas compared to AIP wild type tumours. The authors 242 suggested that the down-regulation of ARNT could be connected to an imbalance in AHR/ARNT 243 complex formation arising from aberrant cAMP signalling (Heliovaara, Raitila, Launonen et al., 244 2009). It's worth to be mentioned, however, that in this study 3 out of 14 AIP mutated samples were 245 pure PRL-secreting adenomas. However, AIP silencing in HeLa, HEK293, or Aip-null mouse 246 embryonic fibroblast cells did not show reduced expression of ARNT, but in GH3 cells caused a 247 partial reduction of ARNT and a clear increase in cell proliferation (Heliovaara et al., 2009).

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249 Xenobiotics, the AHR and pituitary tumours: *in vitro* studies.

Some studies showed that in pituitary cell lines, the activation of AHR could result in endocrine effects. Indeed, in GH3 cells, the reversible AHR agonist β -naphthoflavone induced the expression of CYP1A1 and impaired PRL but not the GH expression, without any clear effect on cell proliferation, although the levels of the anti-proliferative signalling cytokine TGFbeta1 were

suppressed (Moran, Brannick and Raetzman, 2012). Similarly, in the rat pituitary gland cells,
TCDD was demonstrated to upregulate known AHR target genes, to upregulate ER1 expression and
to abrogate estradiol-induced PRL expression (Cao, Patisaul and Petersen, 2011).

On the other hand, several environmental contaminants have been shown to act as endocrinedisruptors in pituitary cells.

Perfluoroalkyl acids, perfluorinated compounds with endocrine disruption potential in humans and animals, have been shown to affect GH3 cells growth and the T3-induced cell proliferation, and to interfere with the thyroid hormone receptor (TR) and AHR function (Long, Ghisari and Bonefeld-Jorgensen, 2013).

263 PCBs, chemicals that can disrupt the endocrine function and promote the incidence of tumours, 264 could bind to AHR if they have a dioxin like structure (Giesy and Kannan, 1998; Ludewig and 265 Robertson, 2013). Some authors showed that non-dioxin like PCBs could be involved in the 266 regulation of rat pituitary cells apoptosis with a pro-apoptotic or an anti-apoptotic effect, depending 267 on their chemical structure. They also showed that the regulation of pituitary apoptosis by PCBs 268 involves multiple pathways, occurring through an AHR or a TR-dependent mechanism, and is 269 associated with changes in the expression level and activity of caspases (Raggi, Russo, Urbani et 270 al., 2016).

271 A very recent *in vitro* study showed that incubation with some endocrine disruptors, e.g. phenol and 272 bis-(2-ethylhexyl)-phthalate, increases cell viability, energy content and proliferation, as well as 273 Ccnd1, PTTG, AhR and AIP expression in normal rat pituitary cells (Tapella, Sesta, Cassarino et 274 al., 2016). In addition, the long-term benzene exposition increased GH synthesis in GH3 cells, and 275 this effect was associated with decreased AIP and increased AHR expression (Zunino, Catalano, 276 Guaraldi et al., 2014). In the same study, exposition to benzene also increased the expression of the 277 somatostatin receptor subtype 2 but decreased ZAC1 expression, effect in accordance with a 278 potential impairment of the sensitivity to SSA (Zunino et al., 2014).

279	Phytoestrogens are natural plant components that can interfere with the hormones' actions or
280	secretion. It has been shown that single isoflavonoid metabolites and their mixture and coumestrol
281	induced GH3 cell growth and AHR transactivity dose-dependently (Long, Kruger, Ghisari et al.,
282	2012).
283	Pesticides have been also proved to interfere with TR signalling and AHR function in vitro and
284	might have the potential to cause endocrine disruption in GH3 cells (Ghisari, Long, Tabbo et al.,

285 2015).

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288 Xenobiotics effects on GH, PRL and POMC: studies in animal models.

289 Several data suggest the pituitary gland to be a direct target for TCDD or other toxins and the AHR 290 system to play a physiological role in controlling neuroendocrine functions (Huang, Ceccatelli and 291 Rannug, 2002). In mice, maternal exposure to TCDD attenuates gonadotropin-regulated 292 steroidogenesis and GH expression leading to the impairment of pup development and sexual 293 immaturity via mechanisms that involve AHR activation (Takeda, Taura, Hattori et al., 2014). In 294 mice, loss of AHR leads to a reduction in PRL mRNA, while GH is unaffected (Moran et al., 2012). 295 In 129/SV/C57BL/6 mice, TCDD or beta-naphthoflavone treatment increased the levels of 296 CYP1A1 mRNA and protein, as well as the AHR repressor (AHRR) mRNA levels, in the pituitary 297 gland. Furthermore, a three-fold increase in POMC mRNA was observed in the pituitary of TCDD 298 treated mice. POMC mRNA level was also increased in the pituitary cell line AtT-20 after TCDD 299 treatment (Huang, Ceccatelli, Hoegberg et al., 2003). Exposure to PCBs can affect growth and 300 pituitary GH content in chicken embryos as well (Gould, Cooper and Scanes, 1997). In the rainbow 301 trout's pituitary gland, the treatment with o,p'-DDT (o,p'-dichlorodiphenyltrichloroethane), a 302 xenoestrogen, resulted in a significant induction of GH and PRL mRNA. Co-incubation of 303 pituitaries with TCDD and alpha-napthoflavone (ANF), which is an AHR inhibitor, caused an 304 inhibition of TCDD-induced PRL mRNA increase at the higher and lower concentrations, but these

305 effects were less consistent on GH mRNA levels. However, the responses of PRL and GH mRNA 306 to co-incubation with TCDD and ANF were bi-phasic, since stimulation was seen at the low 307 concentrations and inhibition at the high concentrations. The o,p'-DDT and TCDD effects on the 308 expression of GH and PRL genes in the rainbow trout pituitary were mediated by mechanisms 309 involving the ER and AHR (Elango, Shepherd and Chen, 2006).

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The relationship among environmental pollutants, the AHR pathway and acromegaly: clues
from *in vivo* studies.

Several evidences suggest that environmental contaminants with a carcinogenic potential, can disrupt the hypothalamic/pituitary function with neuroendocrine consequences. However, few studies have investigated in humans or animals the in vivo effects of environmental pollution exposition, or of the AHR pathway alterations on epidemiology, pathophysiology and clinical features of pituitary tumours.

In 2008, Pesatori et al. investigated the incidence of pituitary tumours in the Seveso population exposed to 2,3,7,8-tetrachlorodibenzo-para-dioxin following an industrial accident occurred in 1976. They found a higher risk, despite non statistically significant, of developing pituitary adenomas in subjects exposed to high-intermediate dioxin concentrations as compared with the non-exposed population, suggesting the need for an extended follow-up (Pesatori, Baccarelli, Consonni et al., 2008).

In 2010, we investigated the epidemiology of acromegaly in the province of Messina, one out of nine provinces in Sicily, focusing on the relationship between the geographical pattern of disease distribution and environmental context (Cannavo et al., 2010). On the basis of the degree of exposition to environmental pollutants mostly due to industrial emissions, we identified four distinct zones in the province (Figure 2, Table 1): i) area A (the Ionian area, 31 towns, 76338 inhabitants), low industrial density; ii) area B (the Tyrrhenian area, 71 towns, 287328 inhabitants),

331 middle-low industrial density; iii) area C (the city of Messina, 1 town, 243381 inhabitants), middle 332 industrial density; and iv) area D (5 towns, 47554 inhabitants), high industrial density. The area D 333 is officially identified as a high-risk zone for environmental crisis by the Department of the 334 Environment of the Italian Government, because of the presence of an oil refinery, a steel plant, a 335 thermoelectric power station, a lead recovery plant, and several small factories (Cannavo et al., 336 2010). In the air of area D, elevated atmospheric concentrations of non-methanic hydrocarbons 337 (NMHC) and volatile organic compounds (VOC) have been detected. In this highly polluted area 338 the prevalence of acromegaly was 210 cpm, dramatically higher than that one reported in literature 339 or found in the other less polluted areas of the province of Messina (Table 1). The Relative Risk 340 (RR) calculation in our province showed a significant increased risk to develop acromegaly in the 341 population residing in the highly polluted area D, assuming the population of low polluted area A as 342 a reference (Table 1). RR estimation demonstrated a sort of gradient of increasing prevalence of 343 acromegaly related to an increasing degree of pollution in the different zones (D>C>B>A), as 344 shown by the finding of an increased relative risk also in area C, at middle industrial density 345 (Cannavo et al., 2010). Recent unpublished investigations in the polluted area suggest that the 346 current prevalence is 330 cpm.

On the basis of the studies suggesting an involvement of the AHR pathway in pituitary tumours'
pathophysiology, we evaluated the role of the pollution exposition and of the genetic variants of
AHR and AIP in acromegaly patients.

Several AHR gene polymorphisms have been described, mostly in the exon 10. Some of these genetic variants have been associated with an altered induction of the activity of CYP1A1 and CYP1A2 in response to specific ligands (Harper, Wong, Lam et al., 2002). In one of our study published in 2014, we screened a population of acromegalic patients searching for AHR gene variants. In this cohort of patients, we found only two polymorphisms in the exon 10, the rs2066853 and rs4986836. The AHR rs2066853 variant, which consists in a G>A substitution causing the replacement of a arginine residue with a lisin within the transactivating domain, has been found in a

357 quarter of the patients and associated with increased IGF-1 levels at diagnosis, more invasive 358 pituitary tumours and increased likelihood of developing other neoplasms such as those of thyroid, 359 bladder and emolimphatic organs (Cannavo, Ferrau, Ragonese et al., 2014). This polymorphism has 360 been associated with an increased risk of developing cancer in some studies but not in others (Chen, 361 Tian, Wang et al., 2009; Luo, Zou, Ji et al., 2013; Perez-Morales, Mendez-Ramirez, Moreno-362 Macias et al., 2014). In our cohort of acromegalics the AHR rs4986826 variant was rare and always 363 associated with the rs2066853 (Cannavo et al., 2014). Interestingly, a recent study suggested that 364 the effect of the rs2066853 SNP could interfere with the AHR activity by affecting its secondary 365 structure and stability and, potentially, with ligands' interaction (Aftabi, Colagar and Mehrnejad, 366 2016).

367 Furthermore, we recently evaluated the impact of AHR and AIP gene variants on clinical phenotype 368 of acromegaly in patients living in highly polluted areas (Cannavo, Ragonese, Puglisi et al., 2016). 369 Two hundred and ten patients with acromegaly, from 5 Italian regions (Sicily, Calabria Region, 370 Apulia, Veneto Region and Marche Region), have been searched for AIP and AHR genetic variants. 371 We compared their clinical, biochemical and radiological data after stratification on the basis of the 372 area in which they lived for at least 20 years before diagnosis. The areas have been considered as 373 highly polluted or not polluted on the basis of the official classification of the Department of 374 Environment of the Italian Government that identified 57 districts of national interest (SIN) because 375 of environmental pollution. One of the SIN is located in Veneto Region, 1 in Calabria Region, 2 in 376 Marche Region, 4 in Apulia and 4 in Sicily. Twelve of our patients were from 2 SIN in Sicily, 7 377 from the Sin in Veneto Region, 2 from 2 SIN in Marche Region, 1 from a SIN in Apulia and 1 from 378 the SIN in Calabria Region. In these areas the Regional Agencies for Environmental Protection 379 have detected an increased concentration of non-methanic hydrocarbons, volatile organic 380 compounds (especially benzene), and heavy metals (especially cadmium, chromo, and lead). The 381 presence of asbestos has been reported only in the SIN in Apulia. We found that acromegaly is 382 more severe in patients living in polluted areas if they also carry AHR and/or AIP genetic variants.

Indeed, these patients with AHR/AIP gene variants and coming from polluted areas (HR/VAR^{+ve}) showed higher IGF-1 levels and larger pituitary tumours as compared to the other cases. Moreover, these patients were more resistant to the treatment with SSA administered as first line for six months. Indeed, SSA normalized IGF-1 levels only in 14% of HR/VAR^{+ve} patients, while in the 54-56% of the other patients. Similarly, SSA halved GH values in 14% of the HR/VAR^{+ve} patients while in 54-80% of the acromegalics of the other groups (Figure 3), and significantly reduced GH and IGF-1 levels in all the patients but in the HR/VAR^{+ve} ones (Cannavo et al., 2016).

Interestingly, some authors found an increased plasma concentration of organ-alogenated
compounds, like PCBs and other substances with oestrogen like activity, in acromegalic domestic
cats, thus suggesting an impairment of the xenobiotic metabolizing enzymatic system (Dirtu,
Niessen, Jorens et al., 2013).

394

395 Conclusions

396 The number of environmental contaminants and toxic agents activating the AHR system and with 397 endocrine effects is remarkable. On the other hand, increasing evidence suggests that environmental 398 pollution exposition is associated with human and animal tumorigenesis, but at present few data are 399 available about the potential role of environmental factors in GH secreting pituitary tumours. In 400 vitro and in vivo studies carried out so far showed that: i) cancer-related environmental 401 contaminants, acting also via AHR, can alter pituitary cells biology and function, affecting 402 proliferation and hormone secretion; ii) the AHR signalling pathway has significant functional 403 cross-talk with molecular pathways known to be involved in pituitary tumour pathogenesis; iii) a 404 deregulation of the AHR pathway occurs in somatotropinomas; iv) there is an increased prevalence 405 of acromegaly in a highly polluted area of the province of Messina (Sicily, Italy); v) the AHR gene 406 SNP rs2066853 is more frequent among acromegalics then in the general population and among 407 patients associate to a worse clinical, biochemical profile and more invasive pituitary tumour; vi) 408 patients with AHR or AIP gene variants and living in polluted areas have larger pituitary tumours,

409	higher IGF-1 levels at diagnosis and are more resistant to SSA treatment as compared to
410	acromegalics living in polluted areas but not carrying AHR/AIP gene variants or living in non
411	polluted areas; vii) acromegalic cats have been found with increased levels of environmental
412	contaminants. Nevertheless, the role of specific environmental pollutants and of the AHR pathway
413	in the development, biological features and therapeutic outcome of human GH-secreting pituitary
414	tumours is still far from being definitively cleared. Further efforts are needed to gain knowledge in
415	this complex and intriguing field.
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$\begin{array}{c} 417\\ 418\\ 419\\ 420\\ 421\\ 422\\ 423\\ 424\\ 425\\ 426\\ 427\\ 428\\ 429\\ 430\\ 431\\ 432\\ 433\\ 434\\ 435\\ 436\\ 437\\ 438\\ 439\\ 440\\ 441\\ 442\\ 443\\ 444\\ 445\\ 446\\ 447\\ 448\\ 449\end{array}$	CHRIER MAN
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453 FIGURE LEGENDS

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455 Figure 1. In the cytosol, the unbound aryl hydrocarbon receptor (AHR) forms a complex with 456 AHR-interacting protein (AIP), the co-chaperone protein p23 and two heat shock protein 90 457 (Hsp90) molecules. AIP is an oncosuppressor involved in the pathogenesis of pituitary tumours. 458 AHR recognises several exogenous and endogenous ligands, whose binding results in nuclear 459 translocation of AHR, dissociation from the chaperone proteins, heterodimerization with a nuclear 460 translocator (ARNT) and subsequent binding to xenobiotic-responsive elements (XREs) modulated 461 by other coregulators. This leads to transactivation of several genes encoding phase I and II 462 xenobiotic metabolizing enzymes, such as cytochrome P450s (CYP1A1, CYP1A2 and CYP1B1), 463 as well as other genes coding for non-enzymatic proteins. Other genomic and non-genomic 'non-464 canonical' effects (on the cell-cycle, contact inhibition, cell adhesion, function and metabolism of 465 the oestrogen receptors, and DNA repairing processes) can be the direct consequence of the 466 activation of this transcriptional factor, contributing to the disruption of cell homeostasis and 467 tumorigenic processes. AHRR: AHR repressor.

468

Figure 2. Map of the province of Messina, 108 towns distributed in4 zones characterized by
different environmental context: i) area A (the Ionian area), low industrial density; ii) area B
(theTyrrhenian area), middle-low industrial density; iii) area C (the city of Messina), middle
industrial density; and iv) area D, high industrial density.

473

Figure 3. Percentage of acromegaly patients who normalized IGF-1 levels or reduced GH
concentrations >50%. HR/VAR^{+ve}: patients with AIP/AHR variants and living in polluted areas;
Other cases: patients living in polluted areas but without AIP/AHR variants and patients living in non polluted areas, regardless of AIP/AHR gene status.

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485 **REFERENCES**

- Aftabi, Y., Colagar, A.H. and Mehrnejad, F., 2016. An in silico approach to investigate the source
 of the controversial interpretations about the phenotypic results of the human AhRgene G1661A polymorphism, J Theor Biol. 393, 1-15.
- Alband, N. and Korbonits, M., 2014. Familial pituitary tumors, Handb Clin Neurol. 124, 339-60.
- Aziz, M.A., Periyasamy, S., Al Yousef, Z., AlAbdulkarim, I., Al Otaibi, M., Alfahed, A. and Alasiri,
 G., 2014. Integrated exon level expression analysis of driver genes explain their role in
 colorectal cancer, PLoS One. 9, e110134.
- Beckers, A., Aaltonen, L.A., Daly, A.F. and Karhu, A., 2013. Familial isolated pituitary adenomas
 (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl
 hydrocarbon receptor interacting protein (AIP) gene, Endocr Rev. 34, 239-77.
- Beischlag, T.V., Luis Morales, J., Hollingshead, B.D. and Perdew, G.H., 2008. The aryl
 hydrocarbon receptor complex and the control of gene expression, Crit Rev Eukaryot
 Gene Expr. 18, 207-50.
- Broder, M.S., Chang, E., Cherepanov, D., Neary, M.P. and Ludlam, W.H., 2016. Incidence and
 Prevalence of Acromegaly in the United States: A Claims-Based Analysis, Endocr Pract.
- Cannavo, S., Ferrau, F., Ragonese, M., Curto, L., Torre, M.L., Magistri, M., Marchese, A.,
 Alibrandi, A. and Trimarchi, F., 2010. Increased prevalence of acromegaly in a highly
 polluted area, Eur J Endocrinol. 163, 509-13.
- Cannavo, S., Ferrau, F., Ragonese, M., Romeo, P.D., Torre, M.L., Puglisi, S., De Menis, E., Arnaldi,
 G., Salpietro, C., Cotta, O.R., Albani, A., Ruggeri, R.M. and Trimarchi, F., 2014. Increased
 frequency of the rs2066853 variant of aryl hydrocarbon receptor gene in patients with
 acromegaly, Clin Endocrinol (Oxf). 81, 249-53.
- Cannavo, S., Ragonese, M., Puglisi, S., Romeo, P.D., Torre, M.L., Alibrandi, A., Scaroni, C., Occhi,
 G., Ceccato, F., Regazzo, D., De Menis, E., Sartorato, P., Arnaldi, G., Trementino, L.,
 Trimarchi, F. and Ferrau, F., 2016. Acromegaly Is More Severe in Patients With AHR or
 AIP Gene Variants Living in Highly Polluted Areas, J Clin Endocrinol Metab. 101, 18729.
- Cao, J., Patisaul, H.B. and Petersen, S.L., 2011. Aryl hydrocarbon receptor activation in
 lactotropes and gonadotropes interferes with estradiol-dependent and -independent
 preprolactin, glycoprotein alpha and luteinizing hormone beta gene expression, Mol
 Cell Endocrinol. 333, 151-9.
- 517 Capatina, C. and Wass, J.A., 2015. 60 YEARS OF NEUROENDOCRINOLOGY: Acromegaly, J 518 Endocrinol. 226, T141-60.
- Cazabat, L., Bouligand, J., Salenave, S., Bernier, M., Gaillard, S., Parker, F., Young, J., GuiochonMantel, A. and Chanson, P., 2012. Germline AIP mutations in apparently sporadic
 pituitary adenomas: prevalence in a prospective single-center cohort of 443 patients, J
 Clin Endocrinol Metab. 97, E663-70.
- 523 Chen, D., Tian, T., Wang, H., Liu, H., Hu, Z., Wang, Y., Liu, Y., Ma, H., Fan, W., Miao, R., Sun, W.,
 524 Wang, Y., Qian, J., Jin, L., Wei, Q., Shen, H., Huang, W. and Lu, D., 2009. Association of
 525 human aryl hydrocarbon receptor gene polymorphisms with risk of lung cancer among
 526 cigarette smokers in a Chinese population, Pharmacogenet Genomics. 19, 25-34.
- 527 Dal, J., Feldt-Rasmussen, U., Andersen, M., Kristensen, L.O., Laurberg, P., Pedersen, L., Dekkers,
 528 O.M., Sorensen, H.T. and Jorgensen, J.O., 2016. Acromegaly incidence, prevalence,
 529 complications and long-term prognosis: a nationwide cohort study, Eur J Endocrinol.
 530 175, 181-90.
- Daly, A.F., Rixhon, M., Adam, C., Dempegioti, A., Tichomirowa, M.A. and Beckers, A., 2006. High
 prevalence of pituitary adenomas: a cross-sectional study in the province of Liege,
 Belgium, J Clin Endocrinol Metab. 91, 4769-75.

- 534 Daly, A.F., Tichomirowa, M.A., Petrossians, P., Heliovaara, E., Jaffrain-Rea, M.L., Barlier, A., 535 Naves, L.A., Ebeling, T., Karhu, A., Raappana, A., Cazabat, L., De Menis, E., Montanana, 536 C.F., Raverot, G., Weil, R.J., Sane, T., Maiter, D., Neggers, S., Yaneva, M., Tabarin, A., 537 Verrua, E., Eloranta, E., Murat, A., Vierimaa, O., Salmela, P.I., Emy, P., Toledo, R.A., 538 Sabate, M.I., Villa, C., Popelier, M., Salvatori, R., Jennings, J., Longas, A.F., Labarta Aizpun, 539 I.I., Georgitsi, M., Paschke, R., Ronchi, C., Valimaki, M., Saloranta, C., De Herder, W., 540 Cozzi, R., Guitelman, M., Magri, F., Lagonigro, M.S., Halaby, G., Corman, V., Hagelstein, 541 M.T., Vanbellinghen, J.F., Barra, G.B., Gimenez-Roqueplo, A.P., Cameron, F.J., Borson-Chazot, F., Holdaway, I., Toledo, S.P., Stalla, G.K., Spada, A., Zacharieva, S., Bertherat, J., 542 543 Brue, T., Bours, V., Chanson, P., Aaltonen, L.A. and Beckers, A., 2010. Clinical characteristics and therapeutic responses in patients with germ-line AIP mutations 544 545 and pituitary adenomas: an international collaborative study, J Clin Endocrinol Metab. 546 95, E373-83.
- de Oliveira, S.K., Hoffmeister, M., Gambaryan, S., Muller-Esterl, W., Guimaraes, J.A. and
 Smolenski, A.P., 2007. Phosphodiesterase 2A forms a complex with the co-chaperone
 XAP2 and regulates nuclear translocation of the aryl hydrocarbon receptor, J Biol
 Chem. 282, 13656-63.
- Denison, M.S. and Nagy, S.R., 2003. Activation of the aryl hydrocarbon receptor by structurally
 diverse exogenous and endogenous chemicals, Annu Rev Pharmacol Toxicol. 43, 309 34.
- 554 Denison, M.S., Pandini, A., Nagy, S.R., Baldwin, E.P. and Bonati, L., 2002. Ligand binding and 555 activation of the Ah receptor, Chem Biol Interact. 141, 3-24.
- Dietrich, C. and Kaina, B., 2010. The aryl hydrocarbon receptor (AhR) in the regulation of cell cell contact and tumor growth, Carcinogenesis. 31, 1319-28.
- Dirtu, A.C., Niessen, S.J., Jorens, P.G. and Covaci, A., 2013. Organohalogenated contaminants in
 domestic cats' plasma in relation to spontaneous acromegaly and type 2 diabetes
 mellitus: a clue for endocrine disruption in humans?, Environ Int. 57-58, 60-7.
- Elango, A., Shepherd, B. and Chen, T.T., 2006. Effects of endocrine disrupters on the
 expression of growth hormone and prolactin mRNA in the rainbow trout pituitary, Gen
 Comp Endocrinol. 145, 116-27.
- Feng, S., Cao, Z. and Wang, X., 2013. Role of aryl hydrocarbon receptor in cancer, Biochim
 Biophys Acta. 1836, 197-210.
- Ferrau, F., Romeo, P.D., Puglisi, S., Ragonese, M., Torre, M.L., Scaroni, C., Occhi, G., De Menis, E.,
 Arnaldi, G., Trimarchi, F. and Cannavo, S., 2016. Analysis of GPR101 and AIP genes
 mutations in acromegaly: a multicentric study, Endocrine.
- Formosa, R., Xuereb-Anastasi, A. and Vassallo, J., 2013. Aip regulates cAMP signalling and GH
 secretion in GH3 cells, Endocr Relat Cancer. 20, 495-505.
- Gasiewicz, T.A., Singh, K.P. and Bennett, J.A., 2014. The Ah receptor in stem cell cycling,
 regulation, and quiescence, Ann N Y Acad Sci. 1310, 44-50.
- Ghisari, M., Long, M., Tabbo, A. and Bonefeld-Jorgensen, E.C., 2015. Effects of currently used
 pesticides and their mixtures on the function of thyroid hormone and aryl
 hydrocarbon receptor in cell culture, Toxicol Appl Pharmacol. 284, 292-303.
- Giesy, J.P. and Kannan, K., 1998. Dioxin-like and non-dioxin-like toxic effects of
 polychlorinated biphenyls (PCBs): implications for risk assessment, Crit Rev Toxicol.
 28, 511-69.
- Goldsmith, S., Lovell-Badge, R. and Rizzoti, K., 2016. SOX2 is sequentially required for
 progenitor proliferation and lineage specification in the developing pituitary,
 Development. 143, 2376-88.
- Gore, A.C., 2010. Neuroendocrine targets of endocrine disruptors, Hormones (Athens). 9, 16 27.

- Gould, J.C., Cooper, K.R. and Scanes, C.G., 1997. Effects of polychlorinated biphenyl mixtures
 and three specific congeners on growth and circulating growth-related hormones, Gen
 Comp Endocrinol. 106, 221-30.
- Hao, N. and Whitelaw, M.L., 2013. The emerging roles of AhR in physiology and immunity,
 Biochem Pharmacol. 86, 561-70.
- Harper, P.A., Riddick, D.S. and Okey, A.B., 2006. Regulating the regulator: factors that control
 levels and activity of the aryl hydrocarbon receptor, Biochem Pharmacol. 72, 267-79.
- Harper, P.A., Wong, J., Lam, M.S. and Okey, A.B., 2002. Polymorphisms in the human AH
 receptor, Chem Biol Interact. 141, 161-87.
- Heliovaara, E., Raitila, A., Launonen, V., Paetau, A., Arola, J., Lehtonen, H., Sane, T., Weil, R.J.,
 Vierimaa, O., Salmela, P., Tuppurainen, K., Makinen, M., Aaltonen, L.A. and Karhu, A.,
 2009. The expression of AIP-related molecules in elucidation of cellular pathways in
 pituitary adenomas, Am J Pathol. 175, 2501-7.
- Hernandez-Ramirez, L.C., Gabrovska, P., Denes, J., Stals, K., Trivellin, G., Tilley, D., Ferrau, F.,
 Evanson, J., Ellard, S., Grossman, A.B., Roncaroli, F., Gadelha, M.R., Korbonits, M. and
 International, F.C., 2015. Landscape of Familial Isolated and Young-Onset Pituitary
 Adenomas: Prospective Diagnosis in AIP Mutation Carriers, J Clin Endocrinol Metab.
 100, E1242-54.
- Hernandez-Ramirez, L.C., Martucci, F., Morgan, R.M., Trivellin, G., Tilley, D., Ramos-Guajardo,
 N., Iacovazzo, D., D'Acquisto, F., Prodromou, C. and Korbonits, M., 2016. Rapid
 Proteasomal Degradation of Mutant Proteins Is the Primary Mechanism Leading to
 Tumorigenesis in Patients With Missense AIP Mutations, J Clin Endocrinol Metab. 101,
 3144-54.
- Huang, P., Ceccatelli, S., Hoegberg, P., Sten Shi, T.J., Hakansson, H. and Rannug, A., 2003. TCDDinduced expression of Ah receptor responsive genes in the pituitary and brain of
 cellular retinol-binding protein (CRBP-I) knockout mice, Toxicol Appl Pharmacol. 192,
 262-74.
- Huang, P., Ceccatelli, S. and Rannug, A., 2002. A study on diurnal mRNA expression of CYP1A1,
 AHR, ARNT, and PER2 in rat pituitary and liver, Environ Toxicol Pharmacol. 11, 11926.
- Jaffrain-Rea, M.L., Angelini, M., Gargano, D., Tichomirowa, M.A., Daly, A.F., Vanbellinghen, J.F.,
 D'Innocenzo, E., Barlier, A., Giangaspero, F., Esposito, V., Ventura, L., Arcella, A.,
 Theodoropoulou, M., Naves, L.A., Fajardo, C., Zacharieva, S., Rohmer, V., Brue, T., Gulino,
 A., Cantore, G., Alesse, E. and Beckers, A., 2009. Expression of aryl hydrocarbon
 receptor (AHR) and AHR-interacting protein in pituitary adenomas: pathological and
 clinical implications, Endocr Relat Cancer. 16, 1029-43.
- Jaffrain-Rea, M.L., Rotondi, S., Turchi, A., Occhi, G., Barlier, A., Peverelli, E., Rostomyan, L.,
 Defilles, C., Angelini, M., Oliva, M.A., Ceccato, F., Maiorani, O., Daly, A.F., Esposito, V.,
 Buttarelli, F., Figarella-Branger, D., Giangaspero, F., Spada, A., Scaroni, C., Alesse, E. and
 Beckers, A., 2013. Somatostatin analogues increase AIP expression in
 somatotropinomas, irrespective of Gsp mutations, Endocr Relat Cancer. 20, 753-66.
- Kasuki, L., Colli, L.M., Elias, P.C., Castro, M. and Gadelha, M.R., 2012. Resistance to octreotide
 LAR in acromegalic patients with high SSTR2 expression: analysis of AIP expression,
 Arq Bras Endocrinol Metabol. 56, 501-6.
- Kasuki, L., Vieira Neto, L., Wildemberg, L.E., Colli, L.M., de Castro, M., Takiya, C.M. and Gadelha,
 M.R., 2012. AIP expression in sporadic somatotropinomas is a predictor of the
 response to octreotide LAR therapy independent of SSTR2 expression, Endocr Relat
 Cancer. 19, L25-9.

- Kazlauskas, A., Poellinger, L. and Pongratz, I., 2000. The immunophilin-like protein XAP2
 regulates ubiquitination and subcellular localization of the dioxin receptor, J Biol
 Chem. 275, 41317-24.
- Kim, Y.R., Jung, E.M., Choi, K.C. and Jeung, E.B., 2012. Synergistic effects of octylphenol and
 isobutyl paraben on the expression of calbindin-D(9)k in GH3 rat pituitary cells, Int J
 Mol Med. 29, 294-302.
- Klinge, C.M., Bowers, J.L., Kulakosky, P.C., Kamboj, K.K. and Swanson, H.I., 1999. The aryl
 hydrocarbon receptor (AHR)/AHR nuclear translocator (ARNT) heterodimer interacts
 with naturally occurring estrogen response elements, Mol Cell Endocrinol. 157, 10519.
- Lecoq, A.L., Viengchareun, S., Hage, M., Bouligand, J., Young, J., Boutron, A., Zizzari, P., Lombes,
 M., Chanson, P. and Kamenicky, P., 2016. AIP mutations impair AhR signaling in
 pituitary adenoma patients fibroblasts and in GH3 cells, Endocr Relat Cancer. 23, 43343.
- 646 Leontiou, C.A., Gueorguiev, M., van der Spuy, J., Quinton, R., Lolli, F., Hassan, S., Chahal, H.S., 647 Igreja, S.C., Jordan, S., Rowe, J., Stolbrink, M., Christian, H.C., Wray, J., Bishop-Bailey, D., 648 Berney, D.M., Wass, J.A., Popovic, V., Ribeiro-Oliveira, A., Jr., Gadelha, M.R., Monson, J.P., 649 Akker, S.A., Davis, J.R., Clayton, R.N., Yoshimoto, K., Iwata, T., Matsuno, A., Eguchi, K., 650 Musat, M., Flanagan, D., Peters, G., Bolger, G.B., Chapple, J.P., Frohman, L.A., Grossman, 651 A.B. and Korbonits, M., 2008. The role of the aryl hydrocarbon receptor-interacting 652 protein gene in familial and sporadic pituitary adenomas, J Clin Endocrinol Metab. 93, 653 2390-401.
- Lloyd, C. and Grossman, A., 2014. The AIP (aryl hydrocarbon receptor-interacting protein)
 gene and its relation to the pathogenesis of pituitary adenomas, Endocrine. 46, 387-96.
- Long, M., Ghisari, M. and Bonefeld-Jorgensen, E.C., 2013. Effects of perfluoroalkyl acids on the
 function of the thyroid hormone and the aryl hydrocarbon receptor, Environ Sci Pollut
 Res Int. 20, 8045-56.
- Long, M., Kruger, T., Ghisari, M. and Bonefeld-Jorgensen, E.C., 2012. Effects of selected
 phytoestrogens and their mixtures on the function of the thyroid hormone and the aryl
 hydrocarbon receptor, Nutr Cancer. 64, 1008-19.
- Ludewig, G. and Robertson, L.W., 2013. Polychlorinated biphenyls (PCBs) as initiating agents
 in hepatocellular carcinoma, Cancer Lett. 334, 46-55.
- Luo, C., Zou, P., Ji, G., Gu, A., Zhao, P. and Zhao, C., 2013. The aryl hydrocarbon receptor (AhR)
 1661G>A polymorphism in human cancer: a meta-analysis, Gene. 513, 225-30.
- Martucci, F., Trivellin, G. and Korbonits, M., 2012. Familial isolated pituitary adenomas: an
 emerging clinical entity, J Endocrinol Invest. 35, 1003-14.
- Melmed, S., 2003. Mechanisms for pituitary tumorigenesis: the plastic pituitary, J Clin Invest.
 112, 1603-18.
- 670 Melmed, S., 2009. Acromegaly pathogenesis and treatment, J Clin Invest. 119, 3189-202.
- Mestron, A., Webb, S.M., Astorga, R., Benito, P., Catala, M., Gaztambide, S., Gomez, J.M.,
 Halperin, I., Lucas-Morante, T., Moreno, B., Obiols, G., de Pablos, P., Paramo, C., Pico, A.,
 Torres, E., Varela, C., Vazquez, J.A., Zamora, J., Albareda, M. and Gilabert, M., 2004.
 Epidemiology, clinical characteristics, outcome, morbidity and mortality in acromegaly
 based on the Spanish Acromegaly Registry (Registro Espanol de Acromegalia, REA),
 Eur J Endocrinol. 151, 439-46.
- Mian, C., Ceccato, F., Barollo, S., Watutantrige-Fernando, S., Albiger, N., Regazzo, D., de Lazzari,
 P., Pennelli, G., Rotondi, S., Nacamulli, D., Pelizzo, M.R., Jaffrain-Rea, M.L., Grimaldi, F.,
 Occhi, G. and Scaroni, C., 2014. AHR over-expression in papillary thyroid carcinoma:
 clinical and molecular assessments in a series of Italian acromegalic patients with a
 long-term follow-up, PLoS One. 9, e101560.

- Morales, J.L. and Perdew, G.H., 2007. Carboxyl terminus of hsc70-interacting protein (CHIP)
 can remodel mature aryl hydrocarbon receptor (AhR) complexes and mediate
 ubiquitination of both the AhR and the 90 kDa heat-shock protein (hsp90) in vitro,
 Biochemistry. 46, 610-21.
- Moran, T.B., Brannick, K.E. and Raetzman, L.T., 2012. Aryl-hydrocarbon receptor activity
 modulates prolactin expression in the pituitary, Toxicol Appl Pharmacol. 265, 139-45.
- Morgan, R.M., Hernandez-Ramirez, L.C., Trivellin, G., Zhou, L., Roe, S.M., Korbonits, M. and
 Prodromou, C., 2012. Structure of the TPR domain of AIP: lack of client protein
 interaction with the C-terminal alpha-7 helix of the TPR domain of AIP is sufficient for
 pituitary adenoma predisposition, PLoS One. 7, e53339.
- Mulero-Navarro, S. and Fernandez-Salguero, P.M., 2016. New Trends in Aryl Hydrocarbon
 Receptor Biology, Front Cell Dev Biol. 4, 45.
- Murray, I.A., Patterson, A.D. and Perdew, G.H., 2014. Aryl hydrocarbon receptor ligands in
 cancer: friend and foe, Nat Rev Cancer. 14, 801-14.
- Nukaya, M., Lin, B.C., Glover, E., Moran, S.M., Kennedy, G.D. and Bradfield, C.A., 2010. The aryl
 hydrocarbon receptor-interacting protein (AIP) is required for dioxin-induced
 hepatotoxicity but not for the induction of the Cyp1a1 and Cyp1a2 genes, J Biol Chem.
 285, 35599-605.
- Oesch-Bartlomowicz, B., Huelster, A., Wiss, O., Antoniou-Lipfert, P., Dietrich, C., Arand, M.,
 Weiss, C., Bockamp, E. and Oesch, F., 2005. Aryl hydrocarbon receptor activation by
 cAMP vs. dioxin: divergent signaling pathways, Proc Natl Acad Sci U S A. 102, 9218-23.
- Perez-Morales, R., Mendez-Ramirez, I., Moreno-Macias, H., Mendoza-Posadas, A.D., MartinezRamirez, O.C., Castro-Hernandez, C., Gonsebatt, M.E. and Rubio, J., 2014. Genetic
 susceptibility to lung cancer based on candidate genes in a sample from the Mexican
 Mestizo population: a case-control study, Lung. 192, 167-73.
- Pesatori, A.C., Baccarelli, A., Consonni, D., Lania, A., Beck-Peccoz, P., Bertazzi, P.A. and Spada,
 A., 2008. Aryl hydrocarbon receptor-interacting protein and pituitary adenomas: a
 population-based study on subjects exposed to dioxin after the Seveso, Italy, accident,
 Eur J Endocrinol. 159, 699-703.
- Petrulis, J.R. and Perdew, G.H., 2002. The role of chaperone proteins in the aryl hydrocarbon
 receptor core complex, Chem Biol Interact. 141, 25-40.
- Raggi, F., Russo, D., Urbani, C., Sardella, C., Manetti, L., Cappellani, D., Lupi, I., Tomisti, L.,
 Martino, E., Marcocci, C. and Bogazzi, F., 2016. Divergent Effects of Dioxin- or NonDioxin-Like Polychlorinated Biphenyls on the Apoptosis of Primary Cell Culture from
 the Mouse Pituitary Gland, PLoS One. 11, e0146729.
- Shanle, E.K. and Xu, W., 2011. Endocrine disrupting chemicals targeting estrogen receptor
 signaling: identification and mechanisms of action, Chem Res Toxicol. 24, 6-19.
- Stanford, E.A., Wang, Z., Novikov, O., Mulas, F., Landesman-Bollag, E., Monti, S., Smith, B.W.,
 Seldin, D.C., Murphy, G.J. and Sherr, D.H., 2016. The role of the aryl hydrocarbon
 receptor in the development of cells with the molecular and functional characteristics
 of cancer stem-like cells, BMC Biol. 14, 20.
- Takeda, T., Taura, J., Hattori, Y., Ishii, Y. and Yamada, H., 2014. Dioxin-induced retardation of
 development through a reduction in the expression of pituitary hormones and possible
 involvement of an aryl hydrocarbon receptor in this defect: a comparative study using
 two strains of mice with different sensitivities to dioxin, Toxicol Appl Pharmacol. 278,
 220-9.
- Tapella, L., Sesta, A., Cassarino, M.F., Zunino, V., Catalano, M.G. and Pecori Giraldi, F., 2016.
 Benzene and 2-ethyl-phthalate induce proliferation in normal rat pituitary cells,
 Pituitary.

- Trivellin, G. and Korbonits, M., 2011. AIP and its interacting partners, J Endocrinol. 210, 137-55.
- Tuominen, I., Heliovaara, E., Raitila, A., Rautiainen, M.R., Mehine, M., Katainen, R., Donner, I.,
 Aittomaki, V., Lehtonen, H.J., Ahlsten, M., Kivipelto, L., Schalin-Jantti, C., Arola, J.,
 Hautaniemi, S. and Karhu, A., 2015. AIP inactivation leads to pituitary tumorigenesis
 through defective Galphai-cAMP signaling, Oncogene. 34, 1174-84.
- Vierimaa, O., Georgitsi, M., Lehtonen, R., Vahteristo, P., Kokko, A., Raitila, A., Tuppurainen, K.,
 Ebeling, T.M., Salmela, P.I., Paschke, R., Gundogdu, S., De Menis, E., Makinen, M.J.,
 Launonen, V., Karhu, A. and Aaltonen, L.A., 2006. Pituitary adenoma predisposition
 caused by germline mutations in the AIP gene, Science. 312, 1228-30.
- Villano, C.M., Murphy, K.A., Akintobi, A. and White, L.A., 2006. 2,3,7,8-tetrachlorodibenzo-pdioxin (TCDD) induces matrix metalloproteinase (MMP) expression and invasion in
 A2058 melanoma cells, Toxicol Appl Pharmacol. 210, 212-24.
- Vo, T.T., An, B.S., Yang, H., Jung, E.M., Hwang, I. and Jeung, E.B., 2012. Calbindin-D9k as a
 sensitive molecular biomarker for evaluating the synergistic impact of estrogenic
 chemicals on GH3 rat pituitary cells, Int J Mol Med. 30, 1233-40.
- Wang, W., Knosp, E., Tai, G., Zhao, Y., Liang, Q. and Guo, Y., 2014. Differential effects of estrogen and estrogen receptor antagonist, ICI 182 780, on the expression of calbindin-D9k in rat pituitary prolactinoma GH(3) cells, Int J Clin Exp Pathol. 7, 8498-505.
- Yang, X., Liu, D., Murray, T.J., Mitchell, G.C., Hesterman, E.V., Karchner, S.I., Merson, R.R., Hahn,
 M.E. and Sherr, D.H., 2005. The aryl hydrocarbon receptor constitutively represses c myc transcription in human mammary tumor cells, Oncogene. 24, 7869-81.
- Zunino, V., Catalano, M.G., Guaraldi, F., D'Angelo, V., Arvat, E. and Fortunati, N., 2014. Exposure
 to benzene modifies SSTR2-ZAC1 signalling pathway in GH3 pituitary adenoma cells.,
 Abstract book of the 38th National Meeting of the Italian Society of Endocrinology.
 11(1), p 113, abstract PD17.

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			Prevalence of acromegaly	~
Area	Characteristics <mark>*</mark>	Inhabitants	(c.p.m.)	RR
A (Ionian)	Low industrial density	76338	26	
B (Tyrrhenian)	Middle-low industrial density	287328	84	3.19 (CI 0.75-13.49)
C (City of Messina)	Middle industrial density	243381	115	4.39 (CI 1.05-18.43)
D (High-risk for health zone)	High industrial density	47554	210	8.03 (CI 1.76-36.63)
Province of Messina		654601	97	_
(overall)				

Table 1: Demography, environmental characteristics and epidemiology of acromegaly in the Province of Messina

RR: relative risk; CI: 95% confidence intervals; c.p.m.: cases per million of inhabithants

*: on the basis of a qualitative (different degree of industrialization and urbanization) and quantitative (different concentrations of atmospheric contaminants) evaluation.



Figure 2



Figure 3



HR/VAR^{+ve}: patients with AIP/AHR variants and living in polluted areas;

Other cases: patients living in polluted areas but without AIP/AHR variants and patients living in non polluted areas, regardless of AIP/AHR gene status.

Highlights

- AHR is involved in cells' detoxification mechanisms and in tumorigenic processes.
- Endocrine-disrupting environmental compounds can affect pituitary function via AHR.
- The AHR key partner is AIP, which is involved in pituitary tumorigenesis.
- Pollutants exposition, along with AHR/AIP variants, impacts on acromegaly features.