Vitamin D in human reproduction: the more, the better? An evidence-based critical appraisal

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Abstract. – OBJECTIVE: Vitamin D is a fat-soluble secosteroid hormone that regulates calcium, magnesium, and phosphate homeostasis and plays a pivotal role as antiproliferative and immunomodulatory mediator. Considering the different sources of synthesis and dietary intake as well as the pleiotropic actions in extremely diverse (micro)environments of the body, the supplementation of this Vitamin should be carefully evaluated taking into account the several pathways that it regulates. In the current brief review, we aimed to summarize the available evidence about the topic, in order to suggest the best evidence-based supplementation strategy for human reproduction, avoiding the unuseful (and sometimes hazardous) empiric supplementation.

MATERIALS AND METHODS: Narrative overview, synthesizing the findings of literature retrieved from searches of computerized databases.

RESULTS: Accumulating evidence from in vitro fertilization (IVF) trials suggests that fertilization rate decreases significantly with increasing levels of 250H-D in follicular fluid; in addition, Vitamin D levels in the follicular fluid are negatively correlated to the quality of embryos and the higher values of Vitamin D are associated with lower possibility to achieve pregnancy. Both low and high Vitamin D serum concentrations decrease not only spermatozoa count, but their progressive motility as well as increase morphological abnormalities. Finally, studies in animal models found that severe hypervitaminosis D can reduce the total skeletal calcium store in embryos and may compromise the postnatal survival.

CONCLUSIONS: Based on the retrieved data, we solicit to be extremely selective in deciding for Vitamin D supplementation, since its excess may play a detrimental role in fertility.

Key Words:

Vitamin D, *In vitro* fertilization, Ovary, Infertility, Sex steroid hormone receptors.

Introduction

Vitamin D is a fat-soluble secosteroid hormone that regulates calcium, magnesium, and phosphate homeostasis¹, and plays a pivotal role as antiproliferative and immunomodulatory mediator². Despite the endogenous synthesis, it is possible to supplement Vitamin D with diet: in particular, Vitamin D3 (cholecalciferol) is mainly contained in sea fish fat and cod liver oil, while D2 (ergocalciferol) in plants and mushrooms.

Vitamin D3 is synthesized in the skin from 7-dehydrocholesterol, following the activity of ultraviolet B radiation³. It is metabolized within the body to the hormonally-active form known as 1,25(OH)₂D₃ through two steps: cholecalciferol is hydroxylated to 25-hydroxycholecalciferol by 25-hydroxylase within the liver; after this first hydroxylation, Vitamin D is transported by vitamin D-binding protein (VDBP) into the bloodstream; subsequently, 25-hydroxycholecalciferol serves as a substrate for 1 α-hydroxylase (CY-P27B1), forming the active 1,25(OH)₂D₃ within the kidneys⁴.

The blood level of 25-hydroxycholecalciferol reflects the amount of Vitamin D ingested with diet or synthetized in the skin, since hepatic 25-hydroxylase regulation by these parameters is negligible⁵. On the contrary, the activity of $1-\alpha$ -hydroxylase in the kidney is tightly regulated: both parathyroid hormone and low blood levels of phosphate induce its synthesis and modulate the production of the active hormone⁶.

Despite the impressive number of published studies, data are not still so robust to clearly elucidate all the pathways and effects of Vitamin D in human reproduction. Considering the different

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sources of synthesis and dietary intake as well as the pleiotropic actions in extremely diverse (micro)environments of the body, the supplementation of this Vitamin should be carefully evaluated taking into account the several pathways that it regulates. Based on these elements, in the current brief review we aimed to summarize the available evidence about the topic, in order to suggest the best evidence-based supplementation strategy for human reproduction, avoiding the unuseful (and sometimes hazardous) empiric supplementations.

Effects of Vitamin D on Hormonal and Metabolic Regulation: Master and Minions

Form the chemical point of view, Vitamin D is similar to steroid hormones, and both these compounds act at nuclear level. In particular, Vitamin D binds and activates the nuclear Vitamin D receptor (VDR), which is also a member of the steroid/thyroid/retinoid receptor superfamily? Subsequently, VDR forms a heterodimer with retinoid-X receptor, binds to hormone response elements on DNA and finally drives a pleiotropic cascade of events⁸⁻¹². Among these events, it is widely accepted that 1,25(OH)₂D₃ has a key role in bone metabolism and calcium homeostasis¹³, although it is also known to modulate cell proliferation, differentiation, cancer invasion, and angiogenesis^{3,14-17}.

In this view, since early 80's it was demonstrated that both decidua and placenta produce some active Vitamin D metabolites, including 1,25(OH)₂D₂¹⁸. In addition, the presence of 1- α -hydroxylase and VDR receptors was shown in the ovary (particularly in granulosa cells)¹⁹, endometrium²⁰, pituitary gland²¹ and placenta²², suggesting that Vitamin D can orchestrate several regulatory pathways in human reproduction^{23,24}. 1-α-hydroxylase also seems to play a pivotal role for acrosome reaction in spermatozoa, in a paracrine/autocrine fashion, allowing the increase of intracellular concentration of Ca+2 ions driven by 1,25(OH)₂D₃²⁵. Interestingly, the influence of Vitamin D on human reproduction seems to be tightly connected not only to calcium homeostasis, but also to its paramount importance as direct regulator of the aromatase gene expression: indeed, accumulating evidence suggests that VDR-bound 1,25(OH)₂D₃ acts as a transcription factor to regulate the expression of the CYP19 gene, which is widely known to encode aromatase, the key enzyme for estrogen production²⁶. In addition, 1,25(OH)₂D₃ induces the production of Vitamin D-24-hydroxylase (CYP24A1), which catalyzes its conversion to in-

active metabolites^{27,28}. Underlining these elements, it was recently shown that VDR expression in both human myometrium and endometrium significantly changes throughout the menstrual cycle, suggesting a hormonal-dependent regulation²⁹. Furthermore, recent data showed that Vitamin D is able to stimulate the production of progesterone, estrogen, estrone and insulin-like growth factor binding protein 1 (IGFBP-1) in cultured human ovarian cells; interestingly, Vitamin D and insulin synergistically inhibit IGFBP-1 production, although Vitamin D alone stimulates IGFBP-1 production in the same cells¹⁹. These events clearly suggest that a cross-talk between hormonal and metabolic pathways occurs. In partial agreement with these data, Vitamin D deficiency was also found to be associated with insulin resistance through the modulation of insulin receptor expression³⁰; furthermore, Vitamin D seems to increase insulin sensitivity³¹. Probably, the effects on estrogen homeostasis may be caused by Vitamin D regulatory mechanism on aromatase expression. Nevertheless, scholars showed that Vitamin D causes only poor changes on steroidogenic acute regulatory protein (StAR), 3-β-hydroxysteroid dehydrogenase (3-βHSD) and aromatase mRNA expression¹⁹, so the scenario seems very far to reach a final shape. 1,25(OH),D, seems to increase estrogen and progesterone production also in human placenta^{22,32}. In addition, it has been shown to regulate human chorionic gonadotropin (hCG) expression and secretion in cultured syncytiotrophoblasts, and to stimulate estradiol and progesterone secretion from trophoblasts in a dose-dependent fashion³³. Last but not least, Vitamin D can induce the transcription of HOXA10³⁴ in the endometrium, decidua and placenta. HOXA10 is known to play a pivotal role in orchestrating embryo implantation³⁵ and the development of female tract organogenesis³⁶, together with WNT/β-catenin³⁷. Interestingly, treatment with Vitamin D increases mRNA and protein expression of HOXA10³⁴. In the placental microenvironment, VDR modulates the transfer of calcium between trophoblast and the endometrial decidua, helping to avoid uterine contraction and, subsequently, reduces the risk of preterm delivery³⁸. The presence of VDR was also demonstrated in testicles and spermatozoa³⁹. In particular, 1,25(OH)₂D₃ is able to increase intracellular Ca+2 concentration and the activity of acrosin in spermatozoa in an autocrine/ paracrine fashion²⁵, accounting for the essential acrosome reactions during fertilization of the oocyte. Corroborating the hypothesis of autocrine/paracrine activity, it was found that CYP27B1, which is necessary to form active 1,25(OH)₂D₃, is directly expressed in human male reproductive tract^{40,41}. In addition, Vitamin D has been shown to stimulate calcium uptake through a nuclear receptor activity in Sertoli cells⁴², whose secretory activities are ion channel-dependent. Despite accumulating evidence already suggested a clear role for 1,25(OH)₂D₂ and VDR in the abovementioned pathways, the risk of adverse effects of Vitamin D supplementation is still surrounded by lights and shadows. In this regards, new data tried to shed light on this "grey zone". In particular, it was shown that transcriptional and translational regulation of progesterone biosynthesis-related genes in porcine granulosa cells is significantly altered by 1,25(OH)₂D₃; in addition, the same study⁴³ found that progesterone concentration was decreased in response to 1,25(OH)₂D₃. Progesterone is an essential hormone in reproduction 44,45, and it has been successfully used as prophylaxis in the prevention of spontaneous miscarriage⁴⁶⁻⁴⁹ and preterm labour⁵⁰ thanks to its immunomodulatory properties at the maternal-fetal interface⁵¹. Considering these elements, it is possible to hypothesize that Vitamin D excess may cause a reduction of this important hormone and consequently play a severe detrimental role during early pregnancy.

It was found that in vitro calcitriol treatment in human prostate cancer cell line up-regulated Anti-Müllerian Hormone (AMH) mRNA expression levels⁵² and that functional Vitamin D response element (VDRE) was found in the promoter region of human AMH gene⁵³. Nevertheless, recent evidence from large and well-designed human-based cross-sectional analysis clearly demonstrates that Vitamin D deficiency is highly unlikely to have a detrimental effect on ovarian reserve⁵⁴. Last but not least, the expression of the parathyroid hormone-related protein (PTH-rP) gene is repressed by $1,25(OH)_2D_3^{55-62}$; considering that PTH-rP has a potent vasorelaxant activity in human endometrium^{63,64}, its reduction caused by Vitamin D supplementation may consequently alter the necessary remodeling of spiral artery and the correct placental vascular framework development during early pregnancy. In the following section we try to analyze the reflection of these strong pieces of evidence on the clinical practice.

Clinical Evidence about Vitamin D in Reproduction: a Critical Appraisal

The most important data about Vitamin D levels and human reproduction came from *in vitro* fertilization (IVF) trials. In particular, on one hand it was shown that fertilization rate decreases

significantly with increasing levels of 25OH-D in the follicular fluid; on the other hand, the implantation rate did not significantly increase in the same cohort⁶⁵. Considering that level of 25OH-D in follicular fluid is reflective of body stores of Vitamin D, this evidence suggests that Vitamin D deficiency does not play a pivotal role in the outcome of IVF and, furthermore, that 25OH-D level in follicular fluid could not be used as an independent predictor of clinical pregnancy.

In partial confirmation of these results, it was found that Vitamin D levels in the follicular fluid are negatively correlated to the quality of embryos and that higher values of Vitamin D are associated with lower possibility to achieve pregnancy; in addition, women with overt hypervitaminosis D had poor IVF outcomes⁶⁶. Notably, in the same work, higher levels of Vitamin D corresponded with lower levels of glucose in follicular fluid, corroborating the cross-link between Vitamin D and glucose metabolism (as previously mentioned). In this abnormal condition, altered glucose concentration in the follicular fluid may be detrimental for oocyte maturation and growth of granulosa and cumulus cells, affecting directly the oocyte competence⁶⁷. Furthermore, 1,25(OH)₂D₂ seems to alter AMH sensitivity in granulosa cells obtained from women who underwent oocyte retrieval for IVF: according to a recent data analysis, cumulus granulosa cells cultured with 1,25(OH)₂D₃ show a drastic and significant decrease (32%) in AMH Receptor-II mRNA levels⁶⁸. These data are in partial agreement with previous researches that showed how Vitamin D down-regulated AMH gene and up-regulated follicle-stimulating hormone receptor gene expression in hen's ovaries^{69,70}. Probably, a strong confounding factor which may play a detrimental role in data interpretation about this point is body mass index (BMI). Although it is widely accepted that higher BMI is an independent risk factor for infertility, it was also recently found that the body weight of the women with follicular fluid 25(OH) D deficiency measured in single follicles was significantly higher, regardless of the etiology of infertility⁷¹. Nevertheless, the literature overview about this point is still controversial: as example, others advocated a beneficial role of replete follicular Vitamin D levels for IVF outcomes⁷². These apparently contradictory results may depend (at least in part) not only on Vitamin D concentration in follicular fluid, but also on the expression of VDBP. Corroborating this hypothesis, it was found that decreased expression of VDBP in

the follicular fluid is associated with improved IVF outcomes⁷³. In addition, Firouzabadi et al⁷⁴ showed that Vitamin D levels in the follicular fluid were comparable in the pregnant as well non-pregnant women. This element suggests at least two key points: first of all, the poor efficacy of follicular fluid Vitamin D levels in predicting the pregnancy rate during IVF program; second, the dosage of Vitamin D before IVF program could be considered outdated, since the follicular fluid concentration is a reliable mirror of the whole body store of Vitamin D. Moreover, Vitamin D reserve is higher in women with endometriosis, a condition characterized in most of the cases by infertility⁷⁵. Notably, a retrospective cohort study of 188 infertile women undergoing IVF showed a positive correlation between serum 25OH-D levels and IVF success rate among non-Hispanic white women; nevertheless, an opposite correlation was seen among Asian women, where pregnancy rates were higher in those with lower serum 25OH-D levels⁷⁶, confirming that serum concentration of Vitamin D is not reliable and robust to predict IVF outcomes. Considered altogether, the available evidence allows us to hypothesize that a correlation between the serum/ follicular Vitamin D level and the pregnancy rate in IVF cycle does not occur. Probably, the real field in which the game is played should not be considered the ovary, but the endometrium⁷⁷: some authors found that Vitamin D deficiency was not associated with the number of follicles and oocytes or with the morphology of the embryo in IVF procedures and, furthermore, that Vitamin D deficiency and insufficiency was associated with lower pregnancy rates in recipients of egg donation⁷⁸. Adding these two pieces of evidence to the puzzle, it is possible to hypothesize that Vitamin D at physiologic levels has a beneficial role only on endometrial receptivity79, but conversely an excess of this Vitamin plays a detrimental role in the ovarian homeostasis, disturbing the oocytes development and consequently embryo quality. From the molecular point of view, it is possible that the observed effects in clinical practice may derive from the Vitamin D action on hormonal homeostasis. Indeed, 1,25(OH), D, can reduce significantly estrogen receptor (ER)-α, progesterone receptor (PR)-A and -B and steroid receptor coactivator (SRC) expression in human uterine leiomyoma cells⁸⁰, suggesting a clear role of this Vitamin as antagonist of sex steroid hormone receptors. This antagonistic effect on sex steroid hormone receptors was also confirmed in several

oncological conditions, including endometrial^{81,82}, ovarian²⁸, breast^{83,84} and other⁸⁵⁻⁸⁸ cancers. Apart from female fertility, the probability of a negative effect of Vitamin D also on male fertility was suggested after a discovery of a molecular similarity of VDBP to antisperm antibodies89. In addition, several studies⁹⁰⁻⁹² found that both low (<50 nmol/L) and high (>125 nmol/L) Vitamin D serum concentrations decrease not only spermatozoa count but also their progressive movement as well as increase morphological abnormalities, even after adjustment for age, season, BMI, alcohol intake, and smoking. If excess of Vitamin D can be considered a serious danger for both male and female fertility, hypervitaminosis D seems to be equally dangerous also after conception: evidence from animal model suggests that the embryo is not protected against maternal hypervitaminosis D, but rather that 1,25(OH)₂D₃ is transferred through the placental barrier where it reduces the total skeletal calcium store in embryos and may compromise postnatal survival⁹³⁻⁹⁵.

Conclusions

To date, there are not specific guidelines regarding Vitamin D supplementation for women affected by endocrine disturbances and infertility. Our literature analysis leads us to solicit the necessity to be extremely selective in deciding for its supplementation, according to patient's condition. În particular, comorbidities 96-99 and/or specific period of life^{100,101} which may influence the response to Vitamin D should carefully be evaluated, especially considering the effects on immune system^{102,103}. In addition, the "empiric" supplementation of Vitamin D in healthy women seems not to be evidence-based and, probably, may play detrimental effects on fertility^{104,105}. Confounding and opposite results have been obtained so far from IVF clinical studies: an overall analysis allows us to speculate that Vitamin D at physiologic levels has a beneficial role on endometrial receptivity, whereas an excess of this molecule plays a detrimental role on oocytes development and embryo quality, probably due to its anti-estrogenic effect (i.e. Vitamin D is able to reduce significantly ER-α, PR-A and —B and SRC expression in human uterine leiomyoma cells). According to this element, Vitamin D supplementation should be administered in selected populations and during specific moments of the ovarian cycle, in order to support specifically the

luteal phase. Finally, Vitamin D should be supplemented at appropriate dosage and, according to the most updated recommendations¹⁰⁶, only if serum concentration falls below 50 ng/ml (equivalent to 125 nmol/L).

Declaration of Interest

All authors have no proprietary, financial, professional, or other personal interest of any nature in any product, service, or company. The authors alone are responsible for the content and writing of the paper.

References

- HOLICK MF. High prevalence of vitamin D inadequacy and implications for health. Mayo Clin Proc 2006; 81: 353-373.
- McDonnell DP, Mangelsdorf DJ, Pike JW, Haussler MR, O'Malley BW. Molecular cloning of complementary DNA encoding the avian receptor for vitamin D. Science 1987; 235: 1214-1217.
- WOLF G. The discovery of vitamin D: the contribution of Adolf Windaus. J Nutr 2004; 134: 1299-1302.
- PROSSER DE, JONES G. Enzymes involved in the activation and inactivation of vitamin D. Trends Biochem Sci 2004; 29: 664-673.
- HENRY HL. Regulation of vitamin D metabolism. Best Pract Res Clin Endocrinol Metab 2011; 25: 531-541
- NORMAN AW, HENRY H. The role of the kidney and vitamin D metabolism in health and disease. Clin Orthop Relat Res 1974; 98: 258-287.
- MALLOY PJ, TASIC V, TAHA D, TÜTÜNCÜLER F, YING GS, YIN LK, WANG J, FELDMAN D. Vitamin D receptor mutations in patients with hereditary 1,25-dihydroxyvitamin D-resistant rickets. Mol Genet Metab 2014; 111: 33-40.
- CHRISTAKOS S, DHAWAN P, VERSTUYF A, VERLINDEN L, CAR-MELIET G. VITAMIN D: metabolism, molecular mechanism of action, and pleiotropic effects. Physiol Rev. 2016; 96: 365-408.
- ADORINI L, DANIEL KC, PENNA G. Vitamin D receptor agonists, cancer and the immune system: an intricate relationship. Curr Top Med Chem 2006; 6: 1297-1301.
- 10) COLONESE F, LAGANÀ AS, COLONESE E, SOFO V, SALMERI FM, GRANESE R, TRIOLO O. The pleiotropic effects of vitamin D in gynaecological and obstetric diseases: an overview on a hot topic. Biomed Res Int 2015; 2015: 986281.
- Ross TK, Moss VE, PRAHL JM, DELUCA HF. A nuclear protein essential for binding of rat 1,25-dihydroxyvitamin D3 receptor to its response elements. Proc Natl Acad Sci U S A 1992; 89: 256-260.

- MANGELSDORF DJ, THUMMEL C, BEATO M, HERRLICH P, SCHÜTZ G, UMESONO K, BLUMBERG B, KASTNER P, MARK M, CHAMBON P, EVANS RM. The nuclear receptor superfamily: the second decade. Cell 1995; 83: 835-839.
- 13) Lerchbaum E, Obermayer-Pietsch B. Vitamin D and fertility: a systematic review. Eur J Endocrinol 2012; 166: 765-778.
- 14) SCHWARTZ GG, WANG MH, ZHANG M, SINGH RK, SIEGAL GP. 1 alpha,25-Dihydroxyvitamin D (calcitriol) inhibits the invasiveness of human prostate cancer cells. Cancer Epidemiol Biomarkers Prev 1997; 6: 727-732.
- 15) SCHWARTZ GG, SELZER MG, BURNSTEIN KL, ZHUANG SH, BLOCK NL, BINDERUP L. Inhibition of prostate cancer metastasis in vivo: a comparison of 1,23-dihydroxyvitamin D (calcitriol) and EB1089. Cancer Epidemiol Biomarkers Prev 1999; 8: 241-248.
- 16) Mantell DJ, Owens PE, Bundred NJ, Mawer EB, Can-FIELD AE. 1 alpha,25-dihydroxyvitamin D(3) inhibits angiogenesis in vitro and in vivo. Circ Res 2000; 87: 214-220.
- 17) YLIKOMI T, LAAKSI I, LOU YR, MARTIKAINEN P, MIETTINEN S, PENNANEN P, PURMONEN S, SYVÄLÄ H, VIENONEN A, TUOHIMAA P. Antiproliferative action of vitamin D. Vitam Horm 2002; 64: 357-406.
- 18) Weisman Y, Harell A, Edelstein S, David M, Spirer Z, Golander A. 1 alpha, 25-Dihydroxyvitamin D3 and 24,25-dihydroxyvitamin D3 in vitro synthesis by human decidua and placenta. Nature 1979; 281: 317-319.
- PARIKH G, VARADINOVA M, SUWANDHI P, ARAKI T, ROSENWAKS Z, PORETSKY L, SETO-YOUNG D. Vitamin D regulates steroidogenesis and insulin-like growth factor binding protein-1 (IGFBP-1) production in human ovarian cells. Horm Metab Res 2010; 42: 754-757.
- 20) VIGANO P, LATTUADA D, MANGIONI S, ERMELLINO L, VI-GNALI M, CAPORIZZO E, PANINA-BORDIGNON P, BESOZZI M, DI BLASIO AM. Cycling and early pregnant endometrium as a site of regulated expression of the vitamin D system. J Mol Endocrinol 2006; 36: 415-424.
- Pérez-Fernandez R, Alonso M, Segura C, Muñoz I, García-Caballero T, Diguez C. Vitamin D receptor gene expression in human pituitary gland. Life Sci 1997; 60: 35-42.
- 22) EVANS KN, NGUYEN L, CHAN J, INNES BA, BULMER JN, KILBY MD, HEWISON M. Effects of 25-hydroxyvitamin D3 and 1,25-dihydroxyvitamin D3 on cytokine production by human decidual cells. Biol Reprod 2006; 75: 816-822.
- Luk J, Torrealday S, Neal Perry G, Pal L. Relevance of vitamin D in reproduction. Hum Reprod 2012; 27: 3015-3027.
- 24) JOHNSON LE, DELUCA HF. Vitamin D receptor null mutant mice fed high levels of calcium are fertile. J Nutr 2001; 131: 1787-1791.
- 25) AOUILA S, GUIDO C, MIDDEA E, PERROTTA I, BRUNO R, PELLEGRINO M, ANDO S. Human male gamete endocrinology: 1alpha, 25-dihydroxyvitamin D3

- (1,25(OH)2D3) regulates different aspects of human sperm biology and metabolism. Reprod Biol Endocrinol 2009; 7: 140.
- 26) KINUTA K, TANAKA H, MORIWAKE T, AYA K, KATO S, SEINO Y. Vitamin D is an important factor in estrogen biosynthesis of both female and male gonads. Endocrinology 2000; 141: 1317-1324.
- 27) JONES G, PROSSER DE, KAUFMANN M. 25-Hydroxyvitamin D-24-hydroxylase (CYP24A1): Its important role in the degradation of vitamin D. Arch Biochem Biophys 2012; 523: 9-18.
- 28) RODRIGUEZ GC, TURBOV J, ROSALES R, YOO J, HUNN J, ZAPPIA KJ, LUND K, BARRY CP, RODRIGUEZ IV, PIKE JW, CONRADS TP, DARCY KM, MAXWELL GL, HAMILTON CA, SYED V, THAETE LG. Progestins inhibit calcitriol-induced CYP24A1 and synergistically inhibit ovarian cancer cell viability: an opportunity for chemoprevention. Gynecol Oncol 2016; 143: 159-167
- 29) VIENONEN A, MIETTINEN S, BLÄUER M, MARTIKAINEN PM, TOMÁS E, HEINONEN PK, YLIKOMI T. Expression of nuclear receptors and cofactors in human endometrium and myometrium. J Soc Gynecol Investig 2004; 11: 104-112.
- 30) HEANEY RP. Vitamin D in Health and Disease. Clin J Am Soc Nephrol 2008; 3: 1535-1541.
- Teegarden D, Donkin SS. Vitamin D: emerging new roles in insulin sensitivity. Nutr Res Rev 2009; 22: 82-92.
- 32) BARRERA D, AVILA E, HERNÁNDEZ G, HALHALI A, BIRUETE B, LARREA F, DÍAZ L. Estradiol and progesterone synthesis in human placenta is stimulated by calcitriol. J Steroid Biochem Mol Biol 2007; 103: 529-532.
- 33) AVILA E, HERNÁNDEZ G, MÉNDEZ I, GONZÁLEZ L, HALH-ALI A, LARREA F, MORALES A, DÍAZ L. Calcitriol affects hCG gene transcription in cultured human syncytiotrophoblasts. Reprod Biol Endocrinol 2008; 6: 3.
- 34) Du H, DAFTARY GS, LALWANI SI, TAYLOR HS. Direct regulation of HOXA10 by 1,25-(OH)2D3 in human myelomonocytic cells and human endometrial stromal cells. Mol Endocrinol 2005; 19: 2222-2233.
- BAGOT CN, TROY PJ, TAYLOR HS. Alteration of maternal Hoxa10 expression by in vivo gene transfection affects implantation. Gene Ther 2000; 7: 1378-1384.
- 36) Taylor HS, Vanden Heuvel GB, Igarashi P. A conserved Hox axis in the mouse and human female reproductive system: late establishment and persistent adult expression of the Hoxa cluster genes. Biol Reprod 1997; 57: 1338-1345.
- 37) LAGANA AS, STURLESE E, RETTO G, SOFO V, TRIOLO O. Interplay between misplaced müllerian-derived stem cells and peritoneal immune dysregulation in the pathogenesis of endometriosis. Obstet Gynecol Int 2013; 2013: 527041.
- 38) Belkacemi L, Gariépy G, Mounier C, Simoneau L, La-Fond J. Expression of calbindin-D28k (CaBP28k) in trophoblasts from human term placenta. Biol Reprod 2003; 68: 1943-1950.

- 39) HABIB FK, MADDY SQ, GELLY KJ. Characterisation of receptors for 1,25-dihydroxyvitamin D3 in the human testis. J Steroid Biochem 1990; 35: 195-199.
- CORBETT ST, HILL O, NANGIA AK. Vitamin D receptor found in human sperm. Urology 2006; 68: 1345-1349
- 41) BLOMBERG JENSEN M, NIELSEN JE, JØRGENSEN A, RAJPERT-DE MEYTS E, KRISTENSEN DM, JØRGENSEN N, SKAKKEBAEK NE, JUUL A, LEFFERS H. Vitamin D receptor and vitamin D metabolizing enzymes are expressed in the human male reproductive tract. Hum Reprod 2010; 25: 1303-1311.
- 42) HAMDEN K, CARREAU S, JAMOUSSI K, AYADI F, GARMAZI F, MEZGENNI N, ELFEKI A. Inhibitory effects of 1alpha, 25dihydroxyvitamin D3 and Ajuga iva extract on oxidative stress, toxicity and hypo-fertility in diabetic rat testes. J Physiol Biochem 2008; 64: 231-239.
- 43) Hong SH, Lee JE, Kim HS, Jung YJ, Hwang D, Lee JH, Yang SY, Kim SC, Cho SK, An BS. Effect of vitamin D3 on production of progesterone in porcine granulosa cells by regulation of steroidogenic enzymes. J Biomed Res 2016; 30: 203-208.
- 44) DI RENZO GC, GIARDINA I, CLERICI G, MATTEI A, ALAJMI AH, GERLI S. The role of progesterone in maternal and fetal medicine. Gynecol Endocrinol 2012; 28: 925-932.
- 45) DI RENZO GC, MATTEI A, GOJNIC M, GERLI S. Progesterone and pregnancy. Curr Opin Obstet Gynecol 2005; 17: 598-600.
- 46) HAAS DM, RAMSEY PS. Progestogen for preventing miscarriage. Cochrane database Syst Rev 2013; 10: CD003511.
- 47) TIBBETTS TA, CONNEELY OM, O'MALLEY BW. Progesterone via its receptor antagonizes the pro-inflammatory activity of estrogen in the mouse uterus. Biol Reprod 1999; 60: 1158-1165.
- CARP HJ. Progestogens in the prevention of miscarriage. Horm Mol Biol Clin Investig 2016; 27: 55-62.
- 49) VAN DER LINDEN M, BUCKINGHAM K, FAROUHAR C, KREMER JAM, METWALLY M. Luteal phase support for assisted reproduction cycles. Cochrane database Syst Rev 2015; 7: CD009154.
- DI RENZO GC, ROSATI A, MATTEI A, GOJNIC M, GERLI
 The changing role of progesterone in preterm labour. BJOG 2005; 112 Suppl 1: 57-60.
- BHURKE AS, BAGCHI IC, BAGCHI MK. Progesterone-regulated endometrial factors controlling implantation. Am J Reprod Immunol 2016; 75: 237-245.
- 52) Krishnan AV, Moreno J, Nonn L, Malloy P, Swami S, Peng L, Peehl DM, Feldman D. Novel pathways that contribute to the anti-proliferative and chemopreventive activities of calcitriol in prostate cancer. J Steroid Biochem Mol Biol 2007; 103: 694-702.
- 53) MALLOY PJ, PENG L, WANG J, FELDMAN D. Interaction of the vitamin D receptor with a vitamin

- D response element in the Mullerian-inhibiting substance (MIS) promoter: regulation of MIS expression by calcitriol in prostate cancer cells. Endocrinology 2009; 150: 1580-1587.
- 54) DRAKOPOULOS P, VAN DE VIJVER A, SCHUTYSER V, MI-LATOVIC S, ANCKAERT E, SCHIETTECATTE J, BLOCKEEL C, CAMUS M, TOURNAYE H, POLYZOS NP. The effect of serum vitamin D levels on ovarian reserve markers: a prospective cross-sectional study. Hum Reprod 2017; 32: 208-214.
- 55) KAJITANI T, TAMAMORI-ADACHI M, OKINAGA H, CHIKA-MORI M, IIZUKA M, OKAZAKI T. Negative regulation of parathyroid hormone-related protein expression by steroid hormones. Biochem Biophys Res Commun 2011; 407: 472-478.
- 56) ENDO K, ICHIKAWA F, UCHIYAMA Y, KATSUMATA K, OH-KAWA H, KUMAKI K, OGATA E, IKEDA K. Evidence for the uptake of a vitamin D analogue (OCT) by a human carcinoma and its effect of suppressing the transcription of parathyroid hormone-related peptide gene in vivo. J Biol Chem 1994; 269: 32693-32699.
- 57) FALZON M. DNA sequences in the rat parathyroid hormone-related peptide gene responsible for 1,25-dihydroxyvitamin D3-mediated transcriptional repression. Mol Endocrinol 1996; 10: 672-681
- 58) IKEDA K, LU C, WEIR EC, MANGIN M, BROADUS AE. Transcriptional regulation of the parathyroid hormone-related peptide gene by glucocorticoids and vitamin D in a human C-cell line. J Biol Chem 1989; 264: 15743-15746.
- 59) INOUE D, MATSUMOTO T, OGATA E, IKEDA K. 22-Oxacalcitriol, a noncalcemic analogue of calcitriol, suppresses both cell proliferation and parathyroid hormone-related peptide gene expression in human T cell lymphotrophic virus, type I-infected T cells. J Biol Chem 1993; 268: 16730-16736.
- 60) Kim M, Fujiki R, Murayama A, Kitagawa H, Yamaoka K, Yamamoto Y, Mihara M, Takeyama K, Kato S. 1Alpha,25(OH)2D3-induced transrepression by vitamin D receptor through E-box-type elements in the human parathyroid hormone gene promoter. Mol Endocrinol 2007; 21: 334-342.
- 61) NISHISHITA T, OKAZAKI T, ISHIKAWA T, IGARASHI T, HATA K, OGATA E, FUJITA T. A negative vitamin D response DNA element in the human parathyroid hormone-related peptide gene binds to vitamin D receptor along with Ku antigen to mediate negative gene regulation by vitamin D. J Biol Chem 1998; 273: 10901-10907.
- 62) OKAZAKI T, NISHIMORI S, OGATA E, FUJITA T. Vitamin D-dependent recruitment of DNA-PK to the chromatinized negative vitamin D response element in the PTHrP gene is required for gene repression by vitamin D. Biochem Biophys Res Commun 2003; 304: 632-637.
- 63) Casey ML, MacDonald PC. The endothelin-parathyroid hormone-related protein vasoactive peptide system in human endometrium: modulation by transforming growth factor-beta. Hum Reprod 1996; 11 Suppl 2: 62-82.

- 64) CASEY ML, MIBE M, MACDONALD PC. Regulation of parathyroid hormone-related protein gene expression in human endometrial stromal cells in culture. J Clin Endocrinol Metab 1993; 77: 188-194.
- 65) ALEYASIN A, HOSSEINI MA, MAHDAVI A, SAFDARIAN L, FALLAHI P, MOHAJERI MR, ABBASI M, ESFAHANI F. Predictive value of the level of vitamin D in follicular fluid on the outcome of assisted reproductive technology. Eur J Obstet Gynecol Reprod Biol 2011; 159: 132-137.
- 66) ANIFANDIS GM, DAFOPOULOS K, MESSINI CI, CHALVATZAS N, LIAKOS N, POURNARAS S, MESSINIS IE. Prognostic value of follicular fluid 25-OH vitamin D and glucose levels in the IVF outcome. Reprod Biol Endocrinol 2010; 8: 91.
- 67) Gu L, Liu H, Gu X, Boots C, Moley KH, Wang Q. Metabolic control of oocyte development: linking maternal nutrition and reproductive outcomes. Cell Mol Life Sci 2015; 72: 251-271.
- 68) Merhi Z, Doswell A, Krebs K, Cipolla M. Vitamin D alters genes involved in follicular development and steroidogenesis in human cumulus granulosa cells. J Clin Endocrinol Metab 2014; 99: E1137-1145.
- 69) JOHNSON PA, KENT TR, URICK ME, GILES JR. Expression and regulation of anti-mullerian hormone in an oviparous species, the hen. Biol Reprod 2008; 78: 13-19.
- 70) WOJTUSIK J, JOHNSON PA. Vitamin D regulates anti-Mullerian hormone expression in granulosa cells of the hen. Biol Reprod 2012; 86: 91.
- 71) DERIOUEHEM VA, ANTUNES RA, REGINATTO MW, MANCEBO AC, AREAS P, BLOISE E, SOUZA MDO C, ORTIGA-CARVALHO TM. Body weight and 25-hidroxyvitamin D follicular levels: a prospectivestudy of women submitted to in vitro fertilization. JBRA Assist Reprod 2016; 20: 127-131.
- 72) OZKAN S, JINDAL S, GREENSEID K, SHU J, ZEITLIAN G, HICKMON C, PAL L. Replete vitamin D stores predict reproductive success following in vitro fertilization. Fertil Steril 2010; 94: 1314-1319.
- 73) ESTES SJ, YE B, QIU W, CRAMER D, HORNSTEIN MD, MISS-MER SA. A proteomic analysis of IVF follicular fluid in women <or=32 years old. Fertil Steril 2009; 92: 1569-1578.
- 74) FIROUZABADI RD, RAHMANI E, RAHSEPAR M, FIROUZABADI MM. Value of follicular fluid vitamin D in predicting the pregnancy rate in an IVF program. Arch Gynecol Obstet 2014; 289: 201-206.
- 75) Somigliana E, Panina-Bordignon P, Murone S, Di Lucia P, Vercellini P, Vigano P. Vitamin D reserve is higher in women with endometriosis. Hum Reprod 2007; 22: 2273-2278.
- 76) RUDICK B, INGLES S, CHUNG K, STANCZYK F, PAULSON R, BENDIKSON K. Characterizing the influence of vitamin D levels on IVF outcomes. Hum Reprod 2012; 27: 3321-3327.
- 77) RUDICK B, INGLES SA, STANCZYK F, CHUNG K, PAULSON R, BENDIKSON K. Characterizing the role of vitamin D levels on IVF outcomes: stimulation, embryo, or endometrium? Fertil Steril 2010; 94 Supplement: S72.

- 78) RUDICK BJ, INGLES SA, CHUNG K, STANCZYK FZ, PAULSON RJ, BENDIKSON KA. Influence of vitamin D levels on in vitro fertilization outcomes in donor-recipient cycles. Fertil Steril 2014; 101: 447-452.
- 79) Fox C, Morin S, Jeong JW, Scott RT Jr, Lessey BA. Local and systemic factors and implantation: what is the evidence? Fertil Steril 2016; 105: 873-884
- 80) AL-HENDY A, DIAMOND MP, EL-SOHEMY A, HALDER SK. 1,25-dihydroxyvitamin D3 regulates expression of sex steroid receptors in human uterine fibroid cells. J Clin Endocrinol Metab 2015; 100: E572-582.
- KAVANDI L, COLLIER MA, NGUYEN H, SYED V. Progesterone and calcitriol attenuate inflammatory cytokines CXCL1 and CXCL2 in ovarian and endometrial cancer cells. J Cell Biochem 2012; 113: 3143-3152.
- 82) LEE LR, TENG PN, NGUYEN H, HOOD BL, KAVANDI L, WANG G, TURBOV JM, THAETE LG, HAMILTON CA, MAXWELL GL, RODRIGUEZ GC, CONRADS TP, SYED V. Progesterone enhances calcitriol antitumor activity by upregulating vitamin D receptor expression and promoting apoptosis in endometrial cancer cells. Cancer Prev Res 2013; 6: 731-743.
- 83) LOPES N, SOUSA B, MARTINS D, GOMES M, VIEIRA D, VERONESE LA, MILANEZI F, PAREDES J, COSTA JL, SCHMITT F. Alterations in Vitamin D ignaling and metabolic pathways in breast cancer progression: a study of VDR, CYP27B1 and CYP24A1 expression in benign and malignant breast lesions. BMC Cancer 2010: 10: 483.
- 84) SWAMI S, KRISHNAN A V, FELDMAN D. Receptor abundance and suppresses estrogen actions in MCF-7 human breast cancer cells abundance and suppresses estrogen actions in MCF-7. Clin Cancer Res 2000; 6: 3371-3379.
- 85) KING AN, BEER DG, CHRISTENSEN PJ, SIMPSON RU, RAMNATH N. The vitamin D/CYP24A1 story in cancer. Anticancer Agents Med Chem 2010; 10: 213-224.
- 86) ZELJIC K, SUPIC G, STAMENKOVIC RADAK M, JOVIC N, KO-ZOMARA R, MAGIC Z. Vitamin D receptor, CYP27B1 and CYP24A1 genes polymorphisms association with oral cancer risk and survival. J Oral Pathol Med 2012; 41: 779-787.
- 87) OH JJ, BYUN SS, LEE SE, HONG SK, JEONG CW, CHOI WS, KIM D, KIM HJ, MYUNG SC. Genetic variants in the CYP24A1 gene are associated with prostate cancer risk and aggressiveness in a Korean study population. Prostate Cancer Prostatic Dis 2014; 17: 149-156.
- 88) Campbell MJ, Reddy GS, Koeffler HP. Vitamin D3 analogs and their 24-oxo metabolites equally inhibit clonal proliferation of a variety of cancer cells but have differing molecular effects. J Cell Biochem 1997; 66: 413-425.
- 89) Yu HM, Li XJ, Kadam AL, Cheng CY, Koide SS. Human testis vitamin D binding protein involved in infertility. Arch Androl; 33: 119-128.

- 90) HAMMOUD AO, MEIKLE AW, PETERSON CM, STANFORD J, GIBSON M, CARRELL DT. Association of 25-hydroxy-vitamin D levels with semen and hormonal parameters. Asian J Androl 2012; 14: 855-859.
- 91) RAMLAU-HANSEN CH, MOELLER UK, BONDE JP, OLSEN J, THULSTRUP AM. Are serum levels of vitamin D associated with semen quality? Results from a cross-sectional study in young healthy men. Fertil Steril 2011; 95: 1000-1004.
- 92) Lerchbaum E, Pilz S, Trummer C, Rabe T, Schenk M, Heiboer AC, Obermayer-Pietsch B. Serum vitamin D levels and hypogonadism in men. Andrology 2014; 2: 748-754.
- 93) AMLING M, PRIEMEL M, HOLZMANN T, CHAPIN K, RUEGER JM, BARON R, DEMAY MB. Rescue of the skeletal phenotype of vitamin D receptor-ablated mice in the setting of normal mineral ion homeostasis: Formal histomorphometric and biomechanical analyses. Endocrinology 1999; 140: 4982-4987.
- 94) LIEBEN L, STOCKMANS I, MOERMANS K, CARMELIET G. Maternal hypervitaminosis D reduces fetal bone mass and mineral acquisition and leads to neonatal lethality. Bone 2013; 57: 123-131.
- 95) ORNOY A. The effects of maternal hypercortisonism and hypervitaminosis D2 on fetal osteogenesis and ossification in rats. Teratology 1971; 4: 383-394.
- 96) DI SPIGNA G, DEL PUENTE A, COVELLI B, ABETE E, VARRIALE E, SALZANO S, POSTIGLIONE L. Vitamin D receptor polymorphisms as tool for early screening of severe bone loss in women patients with rheumatoid arthritis. Eur Rev Med Pharmacol Sci 2016; 20: 4664-4669.
- 97) BAIDAL DA, RICORDI C, GARCIA-CONTRERAS M, SONNINO A, FABBRI A. Combination high-dose omega-3 fatty acids and high-dose cholecalciferol in new onset type 1 diabetes: a potential role in preservation of beta-cell mass. Eur Rev Med Pharmacol Sci 2016; 20: 3313-3318.
- 98) Gu SG, Wang CJ, Zhao G, Li GY. Role of vitamin D in regulating the neural stem cells of mouse model with multiple sclerosis. Eur Rev Med Pharmacol Sci 2015; 19: 4004-4011.
- 99) TURHAN CAGLAR FN, UNGAN I, KSANSKI V, OPAN S, ÇIFTÇI S, KURAL A, KOYUNCU A, AKTURK F, KARAKAYA O. Evaluation of serum vitamin D levels in patients with X syndrome. Eur Rev Med Pharmacol Sci 2016; 20: 1155-1160.
- 100) TURKELI A, AYAZ O, UNCU A, OZHAN B, BAS VN, TUFAN AK, YILMAZ O, YUKSEL H. Effects of vitamin D levels on asthma control and severity in pre-school children. Eur Rev Med Pharmacol Sci 2016; 20: 26-36.
- 101) Dong CH, Gao QM, Wang ZM, Wang AM, Zhen P. Vitamin D supplementation for osteoporosis in older adults: can we make it help better? Eur Rev Med Pharmacol Sci 2016; 20: 4612-4621.
- 102) SHYMANSKYY IO, LISAKOVSKA OO, MAZANOVA AO, RIASNIY VM, VELIKY MM. Effects of vitamin D3 and vitamin E on prednisolone-induced alterations

- of phagocyte function. Eur Rev Med Pharmacol Sci 2016; 20: 1379-1383.
- 103) MATTOZZI C, PAOLINO G, SALVI M, MACALUSO L, LUCI C, MORRONE S, CALVIERI S, RICHETTA AG. Peripheral blood regulatory T cell measurements correlate with serum vitamin D level in patients with psoriasis. Eur Rev Med Pharmacol Sci 2016; 20: 1675-1679.
- 104) DABROWSKI F, GRZECHOCINSKA B, WIELGOS M. The role of vitamin D in reproductive health—A Trojan horse or the golden fleece? Nutrients 2015; 7: 4139-4153.
- 105) BIZZARRI M. Warning about supplemental of Vitamin D in women looking for pregnancy. Eur Rev Med Pharmacol Sci 2016; 20: 2187-2188.
- 106) Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA, Gallagher JC, Gallo RL, Jones G, Kovacs CS, Mayne ST, Rosen CJ, Shapses SA. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: what clinicians need to know. J Clin Endocrinol Metab 2011; 96: 53-58.