

Article

Melatonin Protects Band 3 Protein in Human Erythrocytes against H₂O₂-Induced Oxidative Stress

Rossana Morabito ^{1,*}, Alessia Remigante ^{1,2} and Angela Marino ¹

¹ Department of Chemical, Biological, Pharmaceutical and Environmental Sciences, University of Messina, Viale F. Stagno D'Alcontres 31, 98166 Messina, Italy

² Institute of Pharmacology and Toxicology, Paracelsus Medical University, 5020 Salzburg, Austria

* Correspondence: rmorabito@unime.it; Tel.: +39-090-6765214

Received: 27 June 2019; Accepted: 27 July 2019; Published: 28 July 2019



Abstract: The beneficial effect of Melatonin (Mel), recognized as an anti-inflammatory and antioxidant compound, has been already proven to prevent oxidative stress-induced damage associated to lipid peroxidation. As previous studies modeled the impact of oxidative stress on Band 3 protein, an anion exchanger that is essential to erythrocytes homeostasis, by applying H₂O₂ at not hemolytic concentrations and not producing lipid peroxidation, the aim of the present work was to evaluate the possible antioxidant effect of pharmacological doses of Mel on Band 3 protein anion exchange capability. The experiments have been performed on human erythrocytes exposed to 300 μM H₂O₂-induced oxidative stress. To this end, oxidative damage has been verified by monitoring the rate constant for SO₄²⁻ uptake through Band 3 protein. Expression levels of this protein Mel doses lower than 100 μM have also been excluded due to lipid peroxidation, Band 3 protein expression levels, and cell shape alterations, confirming a pro-oxidant action of Mel at certain doses. On the other hand, 100 μM Mel, not provoking lipid peroxidation, restored the rate constant for SO₄²⁻ uptake, Band 3 protein expression levels, and H₂O₂-induced cell shape alterations. Such an effect was confirmed by abolishing the endogenous erythrocytes antioxidant system. Therefore, the present findings show the antioxidant power of Mel at pharmacological concentrations in an in vitro model of oxidative stress not associated to lipid peroxidation, thereby confirming Band 3 protein anion exchange capability measurement as a suitable model to prove the beneficial effect of Mel and support the use of this compound in oxidative stress-related diseases affecting Band 3 protein.

Keywords: Melatonin; Band 3 protein; SO₄²⁻ uptake; oxidative damage; H₂O₂

1. Introduction

The efficiency of Band 3 protein, the anion exchanger present in million copies on erythrocytes membrane and accounting for ion balance, erythrocytes deformability and tissue oxygenation [1], can be monitored by determining the rate constant for SO₄²⁻ uptake, which is slower and more easily detectable than Cl⁻ or HCO₃⁻ uptake [2,3]. SO₄²⁻ uptake measurement has been previously confirmed as a suitable tool to verify the impact of oxidative conditions on erythrocytes homeostasis [4]. In this regard, H₂O₂, used to produce oxidative stress at not hemolytic concentrations [5], impacts on the phospholipid bilayer arrangement and the integral proteins associated to cytoskeleton, including Band 3 protein [6]. Our previous investigations demonstrated a reduction in Band 3 protein anion exchange capability and protein expression levels after a short-term period of exposure (30 min) to 300 μM H₂O₂ associated neither to lipid peroxidation, nor to membrane -SH groups oxidation (mostly belonging to Band 3 protein) or MetHb formation [4]. More recently, SO₄²⁻ uptake measurement has been shown as a valid tool to detect oxidative damage in the case of oxidative stress-related diseases, such as

systemic sclerodermia, diabetes, and canine leishmaniasis [7–10]. In addition, the role of antioxidants in preventing oxidative damage at Band 3 levels has also been investigated [11].

Among antioxidants assumed by food or used in clinics to counteract oxidative conditions often associated to pathologies, melatonin (*N*-acetyl-5-methoxy-tryptamine) has been considered in the present study.

Melatonin (Mel) is involved in photoperiodism as a neuroendocrine transducer, synthesized in the pineal gland as central site of release, detected in serum at nM physiological concentrations and locally produced in other tissues such as gastrointestinal tract [12]. Its anti-inflammatory activity, immune enhancement, prevention of carcinogenesis and tumor promotion are already known [13,14], while its antioxidant power remains not completely clarified [15,16], as both antioxidant and pro-oxidant activity are described, depending on concentration and cell target [17,18]. Though plenty of experiments showed that Mel is a powerful free radical scavenger and antioxidant at different extent detected on lipids and proteins [19], other in vitro studies highlight its context specific pro-oxidant actions, namely in cancer cells [14].

On these premises and taking into account that erythrocytes are a unique and suitable cell model to investigate the response to oxidative stress, due to their simple metabolism and sensitivity to oxidation [20], the aim of the present study was to prove a possible antioxidant effect of Mel on an in vitro model of H₂O₂-induced oxidative stress on human erythrocytes [4]. The hypothesis is that Mel may exert its antioxidant power on an in vitro model that is not associated to lipid peroxidation. Moreover, Band 3 protein could be a target to prove the antioxidant effect which could be, in turn, adopted against oxidative stress-related diseases affecting Band 3 protein [8,9].

In order to understand whether Mel displays anti- or pro-oxidant action on Band 3 protein, lipid peroxidation possibly induced by Mel has been monitored [21]. To this end, pharmacological Mel doses (μ M) have been used according to other in vivo and in vitro studies [15,19]. Successively, the Mel antioxidant effect has been verified by exposing erythrocytes to 300 μ M H₂O₂ and then evaluating cell shape alterations, rate constant for SO₄²⁻ uptake through Band 3 protein along with Band 3 protein expression levels [22,23]. In a separate experimental set, a specific inhibitor of catalase enzyme, playing a pivotal role in defending erythrocytes at H₂O₂ concentrations comprised between 10 and 300 μ M [5,22] has been also used to prove the Mel antioxidant effect in the absence of an endogenous antioxidant system.

2. Results

2.1. Thiobarbituric-Acid-Reactive Substances (TBARS) Levels

As shown in Figure 1, thiobarbituric-acid-reactive substances (TBARS) levels after either 1 μ M or 10 μ M or 50 μ M Mel treatment were significantly higher than control ($p < 0.001$). After treatment with either 100 μ M or 250 μ M or 500 μ M Mel, TBARS levels were significantly lower ($p < 0.001$) than those determined after exposure to either 1 μ M or 10 μ M or 50 μ M Mel, while remaining unchanged with respect to control.

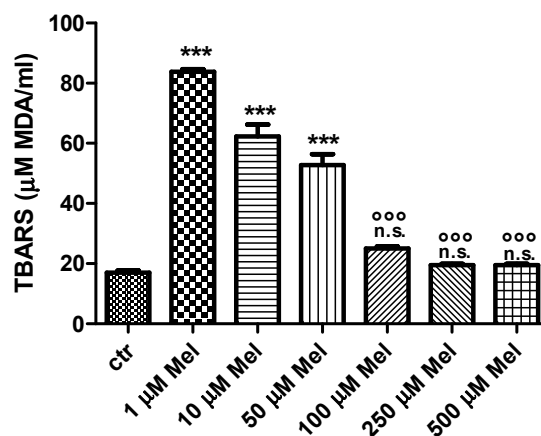


Figure 1. TBARS levels measured as μM malondialdehyde/mL erythrocytes after treatment with Mel at different concentrations. *** $p < 0.001$ vs. ctr; n.s. not significant vs. ctr; $^{\circ\circ\circ}$ $p < 0.001$ vs. either 1 μM or 10 μM or 50 μM Mel, as determined by one way ANOVA followed by Bonferroni's *post hoc* test ($N = 10$).

2.2. Cell Shape

Light microscope observations clearly show cell shape changes after 1 μM Mel treatment (Figure 2b) that were not observed after treatment with 100 μM Mel (Figure 2c).

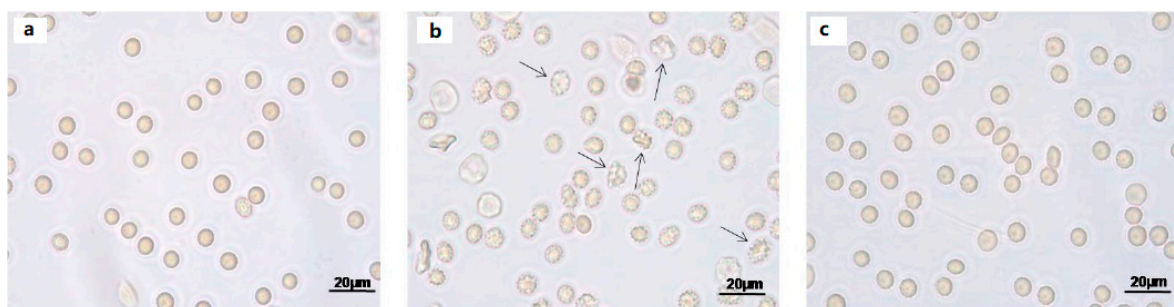


Figure 2. Light microscope observations of untreated (ctr) (a), 1 μM Mel- (b) and 100 μM Mel-treated erythrocytes (c), at 60 min of Mel incubation. Note the significant morphological changes in b (arrows), where 97% altered cells were observed.

2.3. SO_4^- Uptake Measurement

2.3.1. H_2O_2 Treatment

Curves depicted in Figures 3 and 4 describe SO_4^- uptake in different experimental conditions, as a function of time. The rate constant for SO_4^- uptake, accounting for velocity of this process, progressively increased and reached equilibrium within 45 min, with a value of $0.076 \pm 0.001 \text{ min}^{-1}$ in control erythrocytes (Table 1). Pre-exposure to either 1 (Figure 3) or 100 μM Mel (Figure 4) did not alter this parameter, as a rate constant of, respectively, $0.080 \pm 0.001 \text{ min}^{-1}$ and $0.072 \pm 0.001 \text{ min}^{-1}$ was observed ($p < 0.001$, Table 1). Erythrocytes treated with 300 μM H_2O_2 exhibited a rate constant of $0.046 \pm 0.001 \text{ min}^{-1}$ that was significantly lower than control ($p < 0.001$, Table 1) while being comparable to erythrocytes pre-exposed to 1 μM Mel and then to 300 μM H_2O_2 ($0.048 \pm 0.001 \text{ min}^{-1}$). Erythrocytes pre-exposed to 100 μM Mel and then to 300 μM H_2O_2 exhibited a rate constant of $0.074 \pm 0.001 \text{ min}^{-1}$ not significantly different with respect to control, while significantly different with respect to 300 μM H_2O_2 . Treatment with 10 μM DIDS, which was applied at the beginning of SO_4^- medium incubation, completely blocked SO_4^- uptake ($0.017 \pm 0.001 \text{ min}^{-1}$, $p < 0.001$, Table 1).

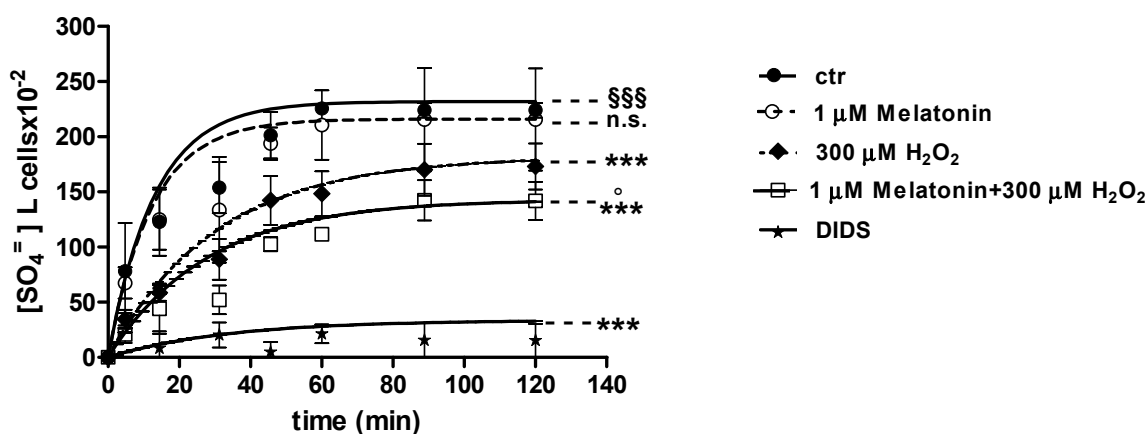


Figure 3. Time course of $\text{SO}_4^{=}$ uptake in untreated erythrocytes (ctr) and erythrocytes treated with $300 \mu\text{M H}_2\text{O}_2$, with or without pre-exposure to $1 \mu\text{M Mel}$. n.s. not significant vs. ctr; *** $p < 0.001$ vs. ctr, $1 \mu\text{M Mel} + 300 \mu\text{M H}_2\text{O}_2$ ° $p < 0.001$ vs. $300 \mu\text{M H}_2\text{O}_2$, \$\$\$ $p < 0.001$, as determined by one way ANOVA followed by Bonferroni's post hoc test ($N = 8$).

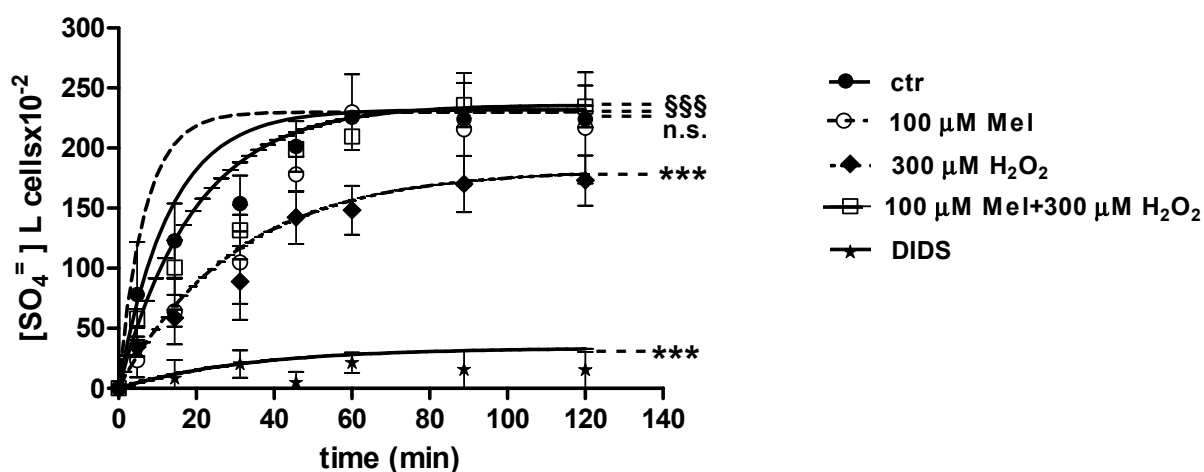


Figure 4. Time course of $\text{SO}_4^{=}$ uptake in untreated erythrocytes (ctr) and erythrocytes treated with $300 \mu\text{M H}_2\text{O}_2$, with or without pre-exposure to $100 \mu\text{M Mel}$. n.s. not significant vs. ctr; *** $p < 0.001$ vs. ctr, $100 \mu\text{M Mel} + 300 \mu\text{M H}_2\text{O}_2$ \$\$\$ $p < 0.001$ vs. $300 \mu\text{M H}_2\text{O}_2$, as determined by one way ANOVA followed by Bonferroni's post hoc test ($N = 8$).

The $\text{SO}_4^{=}$ amount trapped by erythrocytes exposed to 1 and $100 \mu\text{M Mel}$ at 45 min of incubation in $\text{SO}_4^{=}$ medium (192.8 ± 23 and 229.8 ± 14.5 , respectively) was not significantly different with respect to control cells (201.3 ± 21), while, after exposure to $300 \mu\text{M H}_2\text{O}_2$ (142.2 ± 22.2 mM), it was significantly lower than control ($p < 0.001$, Table 1). In DIDS-treated cells, the $\text{SO}_4^{=}$ intracellular amount at 45 min of incubation (4.75 ± 9) was significantly lower than both control and experimental conditions (Table 1).

Light microscope observations at 90 min of incubation in $\text{SO}_4^{=}$ medium show morphological changes in $300 \mu\text{M H}_2\text{O}_2$ -treated erythrocytes (Figure 5b), not seen in erythrocytes treated with $100 \mu\text{M Mel}$ followed by $300 \mu\text{M H}_2\text{O}_2$ (Figure 5c).

Table 1. Rate constant for $\text{SO}_4^{=}$ uptake and amount of $\text{SO}_4^{=}$ amount internalized by erythrocytes in different experimental conditions.

	Rate Constant (min^{-1})	Time (min)	% Decrease	N	$\text{SO}_4^{=}$ Amount Internalized at 45 min of $\text{SO}_4^{=}$ Medium Incubation [$\text{SO}_4^{=}$] L cells $\times 10^{-2}$
Control	0.076 ± 0.001	12	0	10	201.3 ± 21
1 μM Mel	0.080 ± 0.001 n.s.	12.5	0	8	192.8 ± 23 n.s.
100 μM Mel	0.072 ± 0.001 n.s.	13	0	8	229.8 ± 14.5 n.s.
300 μM H_2O_2	0.046 ± 0.001 ***	21	40	8	142.2 ± 22.2 ***
1 μM Mel + 300 μM H_2O_2	0.048 ± 0.001 ***	20	37	8	142.8 ± 21.2 ***
100 μM Mel + 300 μM H_2O_2	0.074 ± 0.001 \$\$\$	13.5	0	10	176.2 ± 6.3 n.s.
50 mM 3-AT + 300 μM H_2O_2	0.018 ± 0.001 ***	55.5	77	9	58 ± 12 ***,###
50 mM 3-AT + 1 μM Mel + 300 μM H_2O_2	0.019 ± 0.001 ***	52.6	75	8	38.4 ± 11 ***
50 mM 3-AT + 100 μM Mel + 300 μM H_2O_2	0.077 ± 0.001 n.s.	12	0	8	219 ± 17 ***
10 μM DIDS	0.017 ± 0.001 ***	55	78	5	4.75 ± 9 ***

Data are presented as means \pm SEM from separate N experiments, where: *** $p < 0.001$ significantly different vs. control; n. s. not significant vs. control; \$\$\$ $p < 0.001$ vs. 300 μM H_2O_2 and control; ### $p < 0.001$ vs. 50 mM 3-AT + 100 μM Mel + 300 μM H_2O_2 as determined by one way ANOVA followed by Bonferroni's Multiple Comparison *post hoc* test.

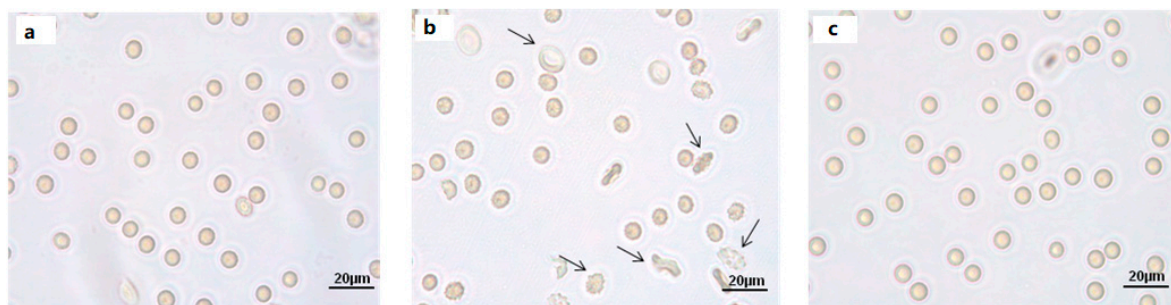


Figure 5. Light microscope observations of untreated (ctr) (a), 300 μM H_2O_2 - (b) and 100 μM Mel + 300 μM H_2O_2 -treated erythrocytes (c), at 90 min of $\text{SO}_4^{=}$ medium incubation. Note the significant morphological changes in b, where, 70% altered cells were observed.

2.3.2. H_2O_2 and 3-AT Treatment

Figures 6 and 7 show the kinetics of $\text{SO}_4^{=}$ uptake in the case of catalase inhibition. With regard to Figure 6, when 50 mM 3-AT and 1 μM Mel were applied before 300 μM H_2O_2 , the rate constant for $\text{SO}_4^{=}$ uptake ($0.019 \pm 0.001 \text{ min}^{-1}$) was significantly lower than control ($0.076 \pm 0.001 \text{ min}^{-1}$), while not being significantly different with respect to 50 mM 3-AT + 300 μM H_2O_2 treatment ($0.018 \pm 0.001 \text{ min}^{-1}$) (Table 1). The $\text{SO}_4^{=}$ amount trapped by erythrocytes treated with 50 mM 3-AT + 1 μM Mel + 300 μM H_2O_2 was 38.4 ± 11 (Table 1).

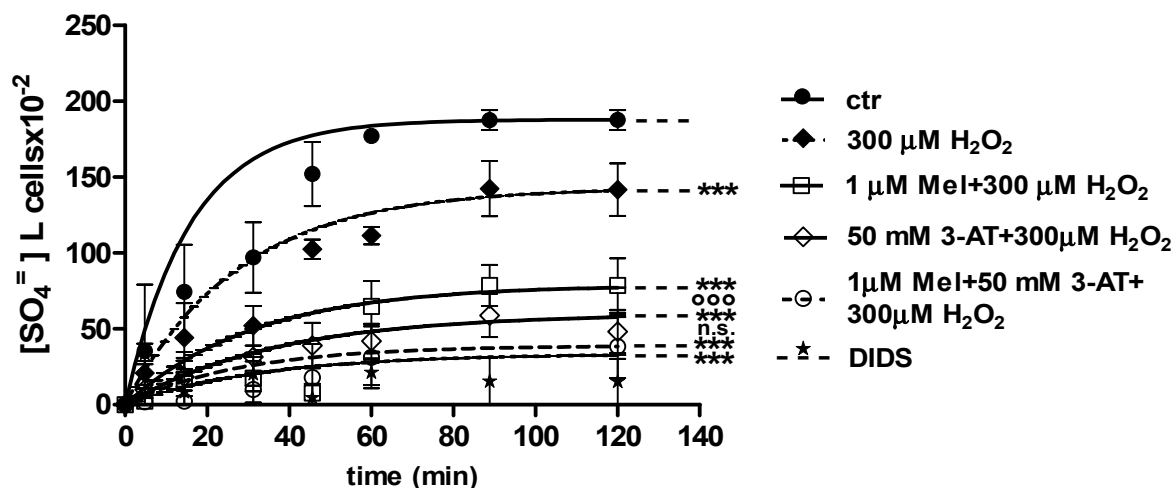


Figure 6. Time course of SO_4^{2-} uptake measured in untreated erythrocytes (ctr) and in erythrocytes treated with 1 μM Mel + 300 μM H_2O_2 with or without pre-exposure to 50 mM 3-AT. *** $p < 0.001$ vs. ctr; 50 mM 3-AT + 300 μM H_2O_2 $^{\circ\circ\circ} p < 0.001$ vs. 300 μM H_2O_2 ; 1 μM Mel + 50 mM 3-AT + 300 μM H_2O_2 n.s. not significant vs. 50 mM 3-AT + 300 μM H_2O_2 as determined by one way ANOVA followed by Bonferroni's post hoc test (N = 8).

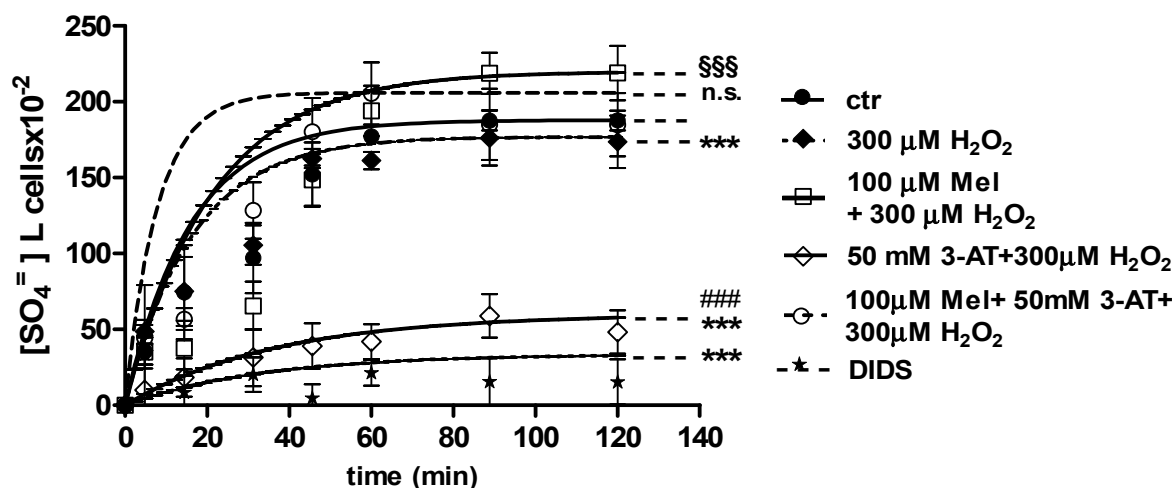


Figure 7. Time course of SO_4^{2-} uptake measured in untreated erythrocytes (ctr) and in erythrocytes treated with 100 μM Mel + 300 μM H_2O_2 with or without 50 mM 3-AT. *** $p < 0.001$ vs. ctr; 100 μM Mel + 50 mM 3-AT + 300 μM H_2O_2 n.s. not significant vs. ctr, 100 μM Mel + 300 μM H_2O_2 $^{\circ\circ\circ} p < 0.001$ vs. 300 μM H_2O_2 ; ### $p < 0.001$ vs. 100 μM Mel + 50 mM 3-AT + 300 μM H_2O_2 as determined by one way ANOVA followed by Bonferroni's post hoc test (N = 8).

With regard to Figure 7, when 50 mM 3-AT and 100 μM Mel were applied before 300 μM H_2O_2 treatment, the rate constant for SO_4^{2-} uptake ($0.077 \pm 0.001 \text{ min}^{-1}$) was not significantly different with respect to both control ($0.076 \pm 0.001 \text{ min}^{-1}$) and 100 μM Mel + 300 μM H_2O_2 treatment ($0.074 \pm 0.001 \text{ min}^{-1}$). This latter value was significantly higher than that one determined after 300 μM H_2O_2 treatment ($0.046 \pm 0.001 \text{ min}^{-1}$, Table 1). SO_4^{2-} amount trapped by erythrocytes treated with 50 mM 3-AT + 100 μM Mel + 300 μM H_2O_2 was 219 ± 17 (Table 1).

2.4. Band 3 Protein Expression Levels Determination

Figures 8 and 9 show that Band 3 protein levels in either 1 μM Mel- or 100 μM Mel-treated erythrocytes were not significantly different with respect to those determined in untreated erythrocytes (control), while, after treatment with 300 μM H_2O_2 , they were significantly lower than control ($p < 0.001$).

A not significant difference ($p < 0.001$) between expression levels in 1 μM Mel + 300 μM H_2O_2 -treated erythrocytes and those measured in 300 μM H_2O_2 -treated erythrocytes was also observed ($p < 0.001$, Figure 8). On the other hand, a significant difference ($p < 0.001$) between expression levels in 100 μM Mel + 300 μM H_2O_2 -treated erythrocytes and those measured in 300 μM H_2O_2 -treated erythrocytes was observed ($p < 0.001$, Figure 9).

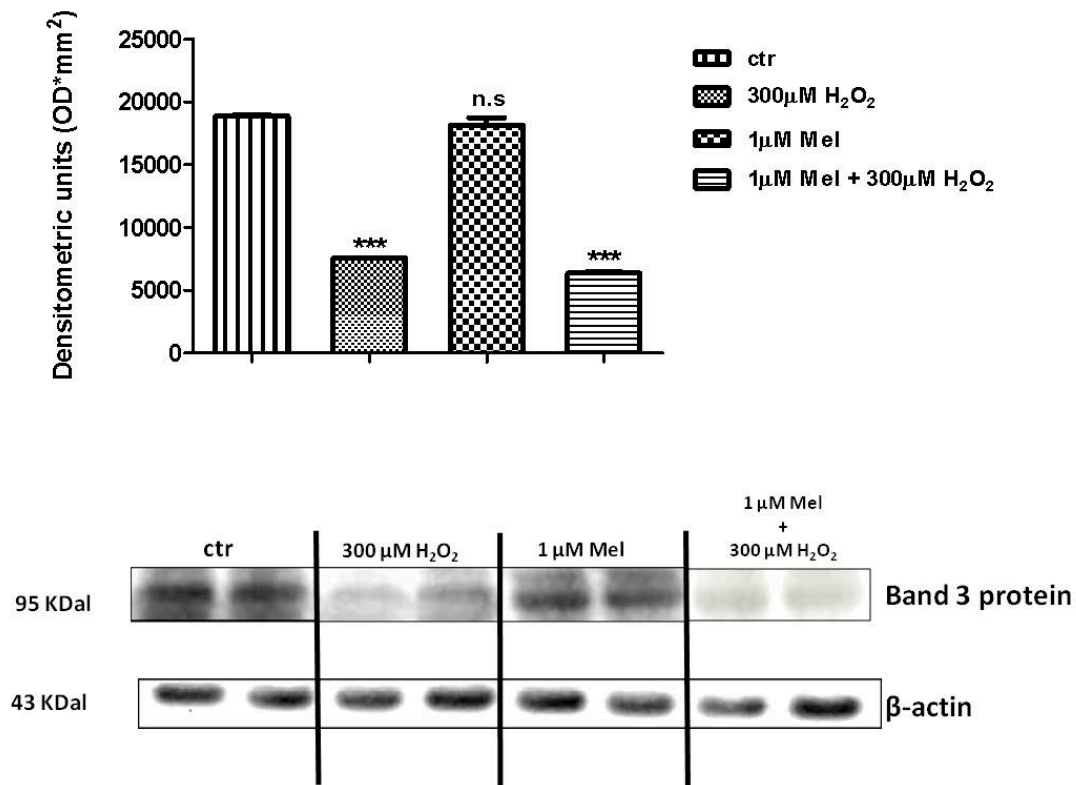


Figure 8. Expression levels of Band 3 protein measured in untreated erythrocytes (ctr), or in 300 μM H_2O_2 - or in 1 μM Mel or in 1 μM Mel + 300 μM H_2O_2 -treated erythrocytes, detected by Western blot analysis. *** $p < 0.001$ vs. ctr; n.s. not significant vs. ctr, as determined by one way ANOVA followed by Bonferroni's *post hoc* test ($N = 5$).

Figure 10 shows that Band 3 protein expression levels in either 50 mM 3-AT + 300 μM H_2O_2 or 50 mM 3-AT + 1 μM Mel or 50 mM 3-AT + 1 μM Mel + 300 μM H_2O_2 -treated erythrocytes were significantly lower than those determined in untreated erythrocytes (control) ($p < 0.001$).

As depicted in Figure 11, Band 3 protein expression levels in either 50 mM 3-AT + 100 μM Mel or 50 mM 3-AT + 100 μM Mel + 300 μM H_2O_2 -treated erythrocytes were not significantly different with respect to those determined in untreated erythrocytes (control), while, after treatment with 50 mM 3-AT + 300 μM H_2O_2 , they were significantly lower than control ($p < 0.001$). Moreover, a significant difference ($p < 0.001$) between expression levels in 50 mM 3-AT + 100 μM Mel + 300 μM H_2O_2 -treated erythrocytes and those measured in 50 mM 3-AT + 300 μM H_2O_2 -treated erythrocytes was observed ($p < 0.001$, Figure 11).

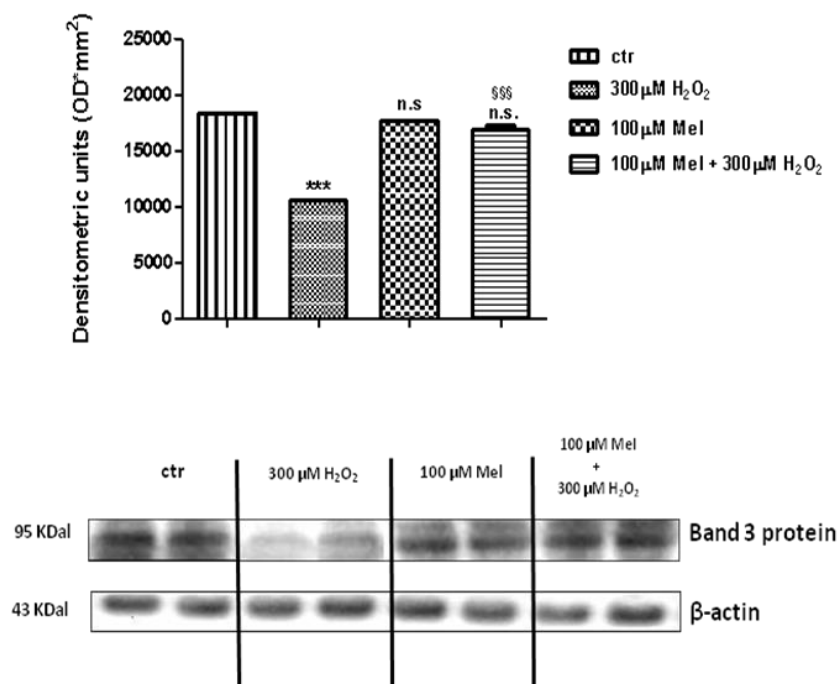


Figure 9. Expression levels of Band 3 protein measured in untreated erythrocytes (ctr), or in 300 μM H₂O₂- or in 100 μM Mel or in 100 μM Mel + 300 μM H₂O₂-treated erythrocytes, detected by Western blot analysis. *** $p < 0.001$ vs. ctr; n.s. not significant vs. ctr; \$\$\$ $p < 0.001$ vs. 300 μM H₂O₂, as determined by one way ANOVA followed by Bonferroni's *post hoc* test (N = 5).

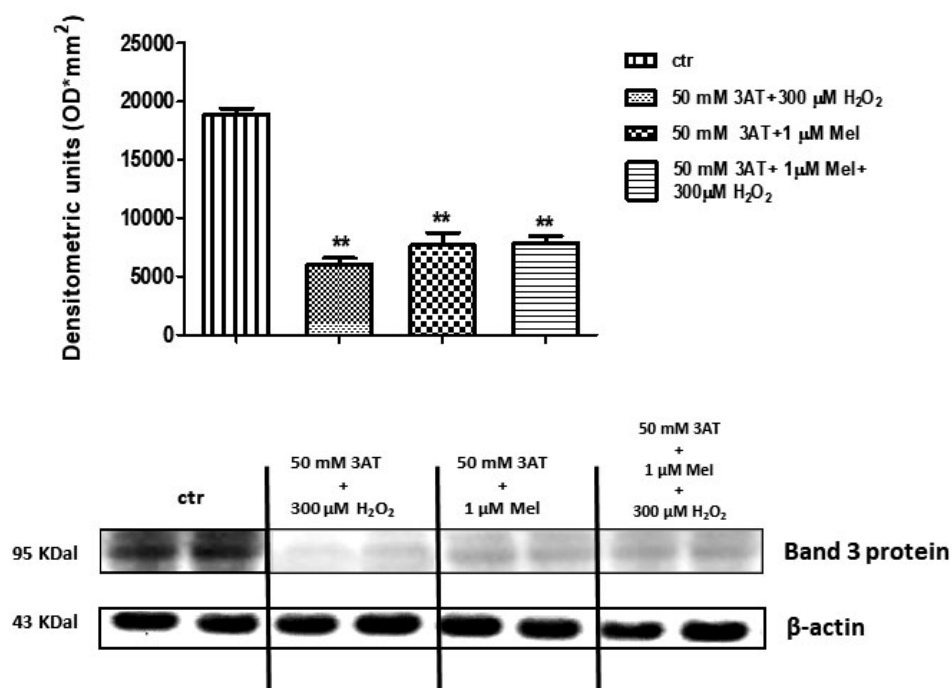


Figure 10. Expression levels of Band 3 protein measured in untreated erythrocytes (ctr), or in 50 mM 3-AT + 300 μM H₂O₂- or in 50 mM 3-AT + 1 μM Mel or in 50 mM 3-AT + 1 μM Mel + 300 μM H₂O₂-treated erythrocytes, detected by Western blot analysis. ** $p < 0.01$ vs. ctr; as determined by one way ANOVA followed by Bonferroni's *post hoc* test (N = 5).

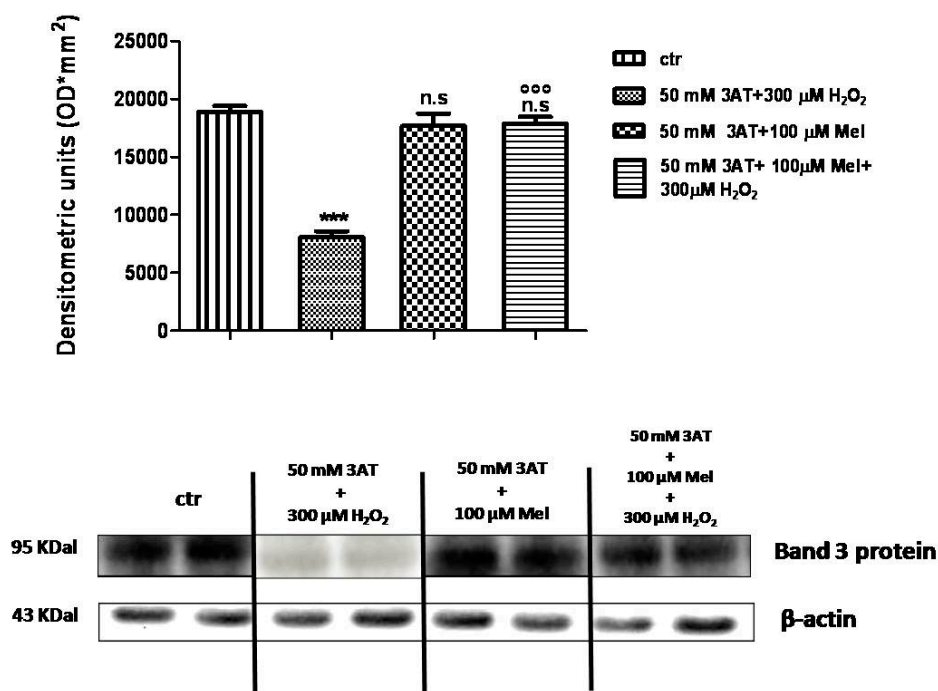


Figure 11. Expression levels of Band 3 protein measured in untreated erythrocytes (ctr), or in 50 mM 3-AT + 300 µM H₂O₂- or in 50 mM 3-AT + 100 µM Mel or in 50 mM 3-AT + 100 µM Mel + 300 µM H₂O₂-treated erythrocytes, detected by Western blot analysis. *** $p < 0.001$ vs. ctr; n.s. not significant vs. ctr; °°° $p < 0.001$ vs. 50 mM 3-AT + 300 µM H₂O₂, as determined by one way ANOVA followed by Bonferroni's *post hoc* test (N = 5).

3. Discussion

High O₂ tension in arterial blood contributes to continuous oxidant species production in erythrocytes, including superoxide (O₂•), hydroxyl (HO•) and H₂O₂ [24], with detrimental effects on cells and tissues when formed in excess [25]. Oxidative damage has been shown to reduce rheological properties, survival of circulating erythrocytes, pathophysiology of abnormal erythrocytes [26] and to affect erythrocytes shape which critically correlates with Band 3 protein function [4,22,27]. Hence, due to their sensitivity to oxidation, erythrocytes are a unique cell model [20] to investigate the impact of oxidative stress and to possibly assess the efficacy of antioxidant therapies. In this regard, Band 3 protein function alterations have been correlated to both in vitro oxidative conditions and oxidative stress-related diseases, i.e., systemic sclerodermia, diabetes and canine leishmaniasis [8,9,28].

As antioxidant supplements may support the endogenous antioxidant system in preserving cell homeostasis under high oxidative conditions [29], the aim of the present investigation was to evaluate the effect of Mel against H₂O₂-induced oxidative damage on anion exchange capability through Band 3 protein in human erythrocytes [4,5,22]. To accomplish this aim, pharmacological doses of Mel have been employed (µM) in line with what already reported in both in vitro and in vivo investigations [15]. In particular, the choice of µM Mel doses is supported by da Silva and co-authors [19], who recently described a beneficial effect of 100 µM Mel on erythrocytes exposed to oxidative stress, as well as by Venegas and co-authors [30] reporting about the therapeutical effect of Mel administered in rats at a µM dose.

It is known that Mel, whose role has been widely investigated [12–17,21,31], exerts both pro-oxidant and antioxidant effects, depending on concentration, time of treatment and cell target [15]. In particular, the evidence of a pro-oxidant action of Mel mediated by ROS production has been obtained on tumoral cells exposed to pharmacological concentrations of Mel (µM to mM range) [15,17,18,21,31–33]. On the other hand, utility and efficacy of Mel in preventing MDA (malondialdehyde) production due to lipid peroxidation in diseases or in experimental models have also been documented [34,35].

As the beneficial effect of Mel in preventing oxidative stress associated to lipid peroxidation has been already demonstrated [34,35], the present study aims to know more about the antioxidant action of Mel against oxidative stress not associated to MDA production by an *in vitro* model performed on human erythrocytes exposed to 300 μM H_2O_2 [36].

It has been already demonstrated that Mel differentially distributes in various subcellular compartments with cell membrane as a major site of concentration (even 1 M), followed by mitochondria, nucleus, and cytosol [15]. On this basis, our hypothesis is that Mel, accumulated at cell membrane level at not pro-oxidant concentrations, may possibly protect ion transport systems against oxidative stress. To this end, not hemolytic concentrations of H_2O_2 (300 μM) reducing anion exchange capability with neither lipid peroxidation, nor Met Hb formation, nor oxidation of membrane –SH groups, have been used to model oxidative damage as previously validated [4,22].

The present findings show that Mel concentrations lower than 100 μM (1 μM) are not useful to prove a possible antioxidant effect on Band 3 protein, as, from one hand, lipid peroxidation and, on the other hand, pro-oxidant action on membrane protein are observed, in line with other investigations [15,21,31]. Both Mel effects, with the first being lipid peroxidation that occurs putatively via ROS production, and the second being pro-oxidant action on membrane proteins, result in cell shape alterations that likely produce a membrane re-arrangement. This arrangement to an extent affects shape, although the rate constant for SO_4^- uptake remains unchanged. Moreover, low Mel concentrations (1 μM) and erythrocytes deformability have been already correlated by Dikmenoglu and co-authors [21]. Similarly to what is shown here, these authors detected significant MDA levels in erythrocytes treated with 1 μM but not by 100 μM Mel. Hence, cell membrane alterations may reasonably depend on this action.

As a second step, 100 μM Mel, pharmacological dose also used in other *in vitro* studies [19] has been considered. Mel doses higher than 100 μM , though not causing lipid peroxidation, have not been used as out of pharmacological range used in both *in vivo* and *in vitro* investigations [15]. Pre-treatment of erythrocytes with 100 μM Mel, not producing lipid peroxidation, impaired the reduction in rate constant for SO_4^- uptake due to 300 μM H_2O_2 , showing for the first time a correlation between the Mel effect, oxidative stress, and Band 3 protein anion exchange capability. In particular, the evidence that H_2O_2 -induced oxidative stress on erythrocytes involves neither lipid peroxidation nor oxidation of membrane –SH groups nor MetHb formation [6] suggests that the beneficial effect of Mel in protecting Band 3 protein should not be related to these mechanisms. Therefore, an interaction of Mel with membrane arrangement can be hypothesized and also supported, as stated above, by light microscope analysis showing cell shape amelioration in erythrocytes treated with 100 μM Mel before oxidative stress application. In line with our observations, other studies have reported the ability of Mel to preserve fluidity in biological membranes, counteracting free radicals-induced rigidity and stabilizing membranes against lipid peroxidation [37]. In this regard, pinealectomy, with consequent reduction in endogenous Mel levels, has been reported to induce an increase in membrane rigidity that can be associated to physiological ageing [38].

The evidence that Mel at 100 μM concentration delays membrane protein degradation and precipitation of hemin onto erythrocytes membrane [39] may explain the preventing effect of Mel against the expected reduction in Band 3 protein expression levels due to 300 μM H_2O_2 [22,40], thus corroborating the hypothesis that the beneficial effect of Mel is exerted at the membrane level.

As oxidative stress alters Band 3 protein aggregation with hemoglobin and cytoskeletal proteins and/or induces protein degradation reducing Band 3 protein expression levels [22,41], it can be suggested that Mel, at 100 μM concentration, would protect Band 3 protein conformation and membrane arrangement, reflecting on erythrocytes morphology. However, to explain Mel action, the role of endogenous antioxidant system should not be neglected. As a pivotal role of catalase in defending erythrocytes against 300 μM H_2O_2 -induced damage has been already proven [22], Mel treatment under oxidative stress has been assayed in the absence of catalase activity. In these experimental conditions, Mel was effective in preventing the reduction of rate constant for SO_4^- uptake due to oxidative stress,

protecting Band 3 protein expression levels even in the absence of catalase activity, thus confirming its antioxidant action. Such observation is supported by Miller and co-authors [42] describing that Mel restores antioxidant enzyme activity in erythrocytes of multiple sclerosis patients and indicating that erythrocytes respond to oxidant challenge with selective induction of certain antioxidant enzymes. Mel has been also reported to alter the activity of glutathione reductase and glucose 6-phosphate dehydrogenase [36,43], with enzymatic activity being increased at 0.02 mM and being decreased at 0.08 mM Mel concentration. With regard to catalase enzyme, Emamgholipour and collaborators [44] have demonstrated that the exposure to H₂O₂ and Mel caused catalase activity and an expression levels increase in peripheral blood mononuclear cells. The antioxidant effect of Mel here showed is exerted at concentrations lower than those reported by Dikmenoglu and co-authors [21] using Mel concentrations comprised between 100 and 1000 µM to prevent MDA formation in erythrocytes exposed to high-concentrated H₂O₂.

In conclusion, these results propose a suitable *in vitro* model to add more knowledge about the antioxidant power of Mel, namely in case of oxidative stress not associated to lipid peroxidation and confirm that erythrocytes homeostasis can be protected by Mel against oxidative stress, when used at a specified range of pharmacological concentrations.

Moreover, a dual effect of Mel on erythrocytes, when used at pharmacological doses, can be suggested: low doses elicit a pro-oxidant action, while high doses exert a beneficial effect. Such effects have been for the first time detected by monitoring the homeostasis of erythrocytes, whereas the vast majority of investigations focused on Mel antioxidant mechanism of action on nucleated cells, with specific regard to nucleus and mitochondria [15].

Finally, as most of the *in vivo* studies demonstrate antioxidant actions of Mel at pharmacological doses, Mel may be suggested as a supplemental therapeutic tool in patients with a deficiency of antioxidant enzymes in red blood cells or in oxidative stress-related diseases associated to Band 3 protein alterations.

4. Materials and Methods

4.1. Solutions and Chemicals

All chemicals were purchased from Sigma (Milan, Italy). H₂O₂ was freshly prepared and diluted from 30% w/w. 3-amino-1,2,4-triazole (3-AT), specific inhibitor of catalase [45], was dissolved in distilled water and diluted from 3 M stock solution through a series of dilution (10 and 100 mM). DIDS (4,4'-diisothiocyanato-stilbene-2,2'-disulfonate) was prepared in DMSO and diluted from 10 mM stock solution. Melatonin, kindly provided by Prof. S. Cuzzocrea (University of Messina, Messina, Italy), was dissolved in 0.5% *v/v* ethanol to prepare 100 mM stock solution. Solvents (ethanol and DMSO) were preventively tested upon erythrocytes at their final concentrations to exclude any damage.

4.2. Erythrocytes Preparation

Human blood was obtained from at least 20 healthy volunteers upon informed consent. Blood was collected in heparinized tubes, washed with an isotonic solution (composition in mM: 145 NaCl, 5 glucose, 20 HEPES (4-(2-hydroxyethyl)-1 piperazineethanesulfonic acid), pH 7.4, osmotic pressure 300 mOsm) and centrifuged thrice (ThermoScientific, 1200 g, 5 min) to remove plasma and buffy coat. Erythrocytes were then suspended to 3% hematocrit in isotonic solution, with or without Mel, and addressed to MDA levels determination.

4.3. Lipid Peroxidation

To prove a possible lipid peroxidative effect of Mel, TBARS (thiobarbituric acid reactive species) levels, resulting from reaction between TBA (thiobarbituric acid) and MDA (malondialdehyde), the end product of the lipid peroxidation, were measured as described by Almroth and collaborators [46] with few modifications. The assay has been performed on erythrocytes washed, suspended at 3%

hematocrit and incubated for 1 h with either 1 μM or 10 μM or 50 μM or 100 μM or 250 μM or 500 μM Mel at 37 °C, in the dark and with gentle shaking. The reaction between MDA and TBA produces TBARS, which is colorimetrically detectable at a 532 nm wavelength. After incubation with Mel at different concentrations, erythrocytes were centrifuged (ThermoScientific, 1000 g, 5 min), suspended to 10% hematocrit in 1 mL distilled water to induce hemolysis, and frozen overnight at -20 °C. After thawing, sample aliquots (200 μL) were treated with 500 μL TBA (1% *v/v* dissolved in 1 N HCl) and incubated at 95 °C for 1 h. Samples were then cooled on ice, centrifuged (Eppendorf, 13,000 g, 15 min, 4 °C), and the supernatant spectrophotometrically read at a 532 nm wavelength (Beckman DU 640).

4.4. SO_4^- Uptake Measurement

After TBARS levels determination, 1 μM and 100 μM Mel doses have been used to carry out the following experiments.

4.4.1. Control Conditions

A turbidimetric method [3] was used to measure SO_4^- uptake through Band 3 protein. After 1 h incubation at 37 °C with or without (control) 100 μM Mel, erythrocytes (3% hematocrit in isotonic solution) were suspended to the same hematocrit in 35 mL isotonic SO_4^- -containing solution, henceforth named SO_4^- medium (composition in mM: 118 Na_2SO_4 , 10 HEPES, 5 glucose, pH 7.4, osmotic pressure 300 mOsm). At specified time intervals (5-10-15-30-45-60-90-120 min), 5 mL samples of erythrocytes suspension were put in a test tube containing 10 μM DIDS, which is a specific and irreversible inhibitor of Band 3 protein [47], and then kept on ice. After the last sample withdrawal, erythrocytes were washed thrice in cold isotonic solution (ThermoScientific, 4 °C, 1000 g, 5 min) to remove SO_4^- from the external medium and hemolyzed in 1 mL distilled water, while proteins were hydrolysed by 4% *v/v* perchloric acid. Cell membranes were discarded by centrifugation (4 °C, 2500 g, 10 min) and the SO_4^- -containing supernatant used for turbidimetric analysis. SO_4^- was precipitated by adding 500 μL supernatant from each sample to 1 mL glycerol and distilled water solution (1:1), 1 mL 4 M NaCl plus HCl (hydrochloric acid 37%) solution (12:1) and 500 μL 1.24 M $\text{BaCl}_2 \cdot 2\text{H}_2\text{O}$. SO_4^- was spectrophotometrically measured at 425 nm wavelength and the absorption converted to $[\text{SO}_4^-]$ L cells 10^{-2} using a calibrated standard curve obtained by precipitating known SO_4^- concentrations.

The rate constant in min^{-1} was then calculated by the following equation: $C_t = C_\infty (1 - e^{-rt}) + C_0$, where C_t , C_∞ and C_0 represent the intracellular SO_4^- concentrations measured at time t , 0 and ∞ respectively; e indicates the Neper number (2.7182818); r is the rate constant describing the process velocity and t is time fixed for each sample withdrawal (5-10-15-30-45-60-90-120 min). Rate constant (min^{-1}) reported in Table 1 *per* each experimental condition represents the time needed to reach 63% of total SO_4^- intracellular concentration [3]. The total amount of SO_4^- trapped by erythrocytes after 45 min of incubation in SO_4^- medium (at equilibrium) has been also considered (Table 1). $[\text{SO}_4^-]$ L cells $\times 10^{-2}$ represents SO_4^- concentration internalized by 10 mL erythrocytes suspended at 3% concentration.

To prove that SO_4^- was effectively internalized through Band 3 protein, erythrocytes were suspended in SO_4^- medium (3% hematocrit) containing 10 μM DIDS. Successively, 5 mL samples were withdrawn at fixed time intervals (5-10-15-30-45-60-90-120 min) and handled as described for control conditions.

4.4.2. Exposure to H_2O_2

To assay the effect of H_2O_2 , erythrocytes, after 1 h incubation with or without Mel (1 or 100 μM), were exposed to 300 μM H_2O_2 for 30 min at 25 °C. Samples were then centrifuged to remove supernatant and erythrocytes re-suspended to 3% hematocrit in SO_4^- medium. SO_4^- uptake was measured as described for control conditions. Results were compared with those from tests performed with H_2O_2 in the absence of Mel.

4.5. Light microscope Observations

Erythrocytes, treated or not with either Mel alone (1 and 100 μM) or with 100 μM Mel \pm 300 μM H_2O_2 , were observed under a light microscope (Leica, 400 \times), at 90 min of $\text{SO}_4^{=}$ medium incubation.

4.6. Erythrocytes Membranes Preparation and SDS-PAGE

Erythrocyte membranes were prepared as previously described [48], with slight modifications. Briefly, after treatment with 300 μM H_2O_2 with or without pre-exposure to Mel (1 or 100 μM), packed erythrocytes were diluted into 1.5 mL of cold hemolysis solution (2.5 mM NaH_2PO_4) containing a protease and a phosphatase inhibitor cocktail (1 mM PMSF, 1 mM NaF, 1 mM sodium orthovanadate) and then repeatedly centrifuged (Eppendorf, 4 $^\circ\text{C}$, 18,000 g, 10 min) to discard hemoglobin. Membrane were then solubilized by 1% (*v/v*) SDS (Sodium Dodecyl Sulfate), and incubated on ice for 20 min. Solubilized membrane proteins, contained in the supernatant, were used for protein content quantification [49] and frozen at -80 $^\circ\text{C}$ until use. Membranes obtained from each experimental condition, once thawed, were solubilized (1:1 volume ratio) in Laemml Buffer [50], heated for 5 min at 95 $^\circ\text{C}$ and 2 μg proteins loaded for anti-Band 3 protein. Samples were then separated on 12% polyacrylamide gel and transferred to a PVDF (polyvinylidene fluoride) membrane.

4.7. Western Blot Analysis

Membranes were incubated at 4 $^\circ\text{C}$ overnight with monoclonal anti-Band 3 protein (1:100,000; Santa Cruz Biotechnology, Dallas, TX, US, produced in mouse), 5% (*w/v*) non-fat dried milk and 0.1% Tween-20 and successively incubated with peroxidase-conjugated goat anti-mouse IgG secondary antibodies (1:5,000, AffiniPure, Cambridge, UK), for 1 h at room temperature. To verify whether blots contained equal amounts of protein, they were also incubated with monoclonal anti- β -actin antibodies (1:1000 Santa Cruz Biotechnology) produced in mouse. Signals were detected by a chemiluminescence detection system (Super Signal West Pico Chemiluminescent Substrate, Pierce Thermo Scientific, Rockford, IL, USA). Protein bands expression (approximately 95 kDa), imported to analysis software (Image Quant TL, v2003) and standardized to β -actin levels, was determined by densitometry (Bio-Rad ChemiDocTM XRS+).

4.8. Catalase Inhibition

In order to prove the antioxidant effect of Mel when catalase activity was inhibited, tests (measurement of rate constant for $\text{SO}_4^{=}$ uptake, light microscope observations and Western blot analysis) were carried out by exposing erythrocytes (3% hematocrit) to 50 mM 3-aminotriazole (3-AT) for 10 min at 25 $^\circ\text{C}$, before incubation with Mel (1 or 100 μM) \pm 300 μM H_2O_2 .

4.9. Experimental Data and Statistics

Data are expressed as arithmetic means \pm S.E.M. for statistical analysis. GraphPad Prism software (version 5.00 for Windows; San Diego, CA, USA) was used. Significant differences between means were tested by one-way analysis of variance (ANOVA), followed by Bonferroni's multiple comparison *post hoc* test. Statistically significant differences were assumed at $p < 0.05$; N represents the number of independent experiments.

Author Contributions: Conceptualization, R.M. and A.M.; Methodology, R.M. and A.R.; Formal Analysis, R.M. and A.R.; Investigation, R.M. and A.M.; Data Curation, R.M. and A.R.; Writing—Original Draft Preparation, R.M.; Writing—Review & Editing, R.M. and A.M.; Supervision, A.M.

Funding: This work was supported by Ministero dell'Istruzione, dell'Università e della Ricerca (MIUR), PRIN 2015 (Prot. 20152HKF3Z).

Conflicts of Interest: The authors declare that they have no competing interests.

References

1. Reithmeier, R.A.F.; Casey, J.R.; Kall, A.C.; Sansom, M.S.P.; Alguel, Y.; Iwata, S. Band 3, the human red cell chloride/bicarbonate anion exchanger (AE1, SLC4A1), in a structural context. *Biochim. Biophys. Acta* **2016**, *1858*, 1507–1532. [\[CrossRef\]](#)
2. Jennings, M.L. Proton fluxes associated with erythrocyte membrane anion exchange. *J. Membr. Biol.* **1976**, *28*, 187–205. [\[CrossRef\]](#)
3. Romano, L.; Passow, H. Characterization of anion transport system in trout red blood cell. *Am. J. Physiol.* **1984**, *246*, 330–338. [\[CrossRef\]](#)
4. Morabito, R.; Romano, O.; La Spada, G.; Marino, A. H₂O₂-induced oxidative stress affects SO₄²⁻ transport in human erythrocytes. *PLoS ONE* **2016**, *11*, e0146485. [\[CrossRef\]](#)
5. Mueller, S.; Riedel, H.D.; Stremmel, W. Determination of catalase activity at physiological hydrogen peroxide concentrations. *Anal. Biochem.* **1997**, *245*, 55–60. [\[CrossRef\]](#)
6. Anong, W.A.; Franco, T.; Chu, H.; Weis, T.L.; Devlin, E.E.; Bodine, D.M.; An, X.; Mohandas, N.; Low, P.S. Adducin forms a bridge between the erythrocyte membrane and its cytoskeleton and regulates membrane cohesion. *Blood* **2009**, *114*, 1904–1912. [\[CrossRef\]](#)
7. De Luca, G.; Gugliotta, T.; Scuteri, A.; Romano, P.; Rinaldi, C.; Sidoti, A.; Amato, A.; Romano, L. The interaction of haemoglobin, magnesium, organic phosphates and band 3 protein in nucleated and anucleated erythrocytes. *Cell. Biochem. Funct.* **2004**, *22*, 179–186. [\[CrossRef\]](#)
8. Morabito, R.; Remigante, A.; Bagnato, G.; Roberts, W.N.; Sciortino, D.; D'Angelo, T.; Iannelli, F.; Iannelli, D.; Cordova, F.; Cirillo, M.; et al. Band 3 protein function and oxidative stress in erythrocytes from Systemic Sclerosis patients with interstitial lung disease. *Eur. J. Clin. Biomed. Sci.* **2017**, *3*, 80–84. [\[CrossRef\]](#)
9. Morabito, R.; Remigante, A.; Cavallaro, M.; Taormina, A.; La Spada, G.; Marino, A. Anion exchange through band 3 protein in canine leishmaniasis at different stages of disease. *Pflugers Arch.* **2017**, *469*, 713–724. [\[CrossRef\]](#)
10. Teti, D.; Crupi, M.; Busá, M.; Valenti, A.; Loddo, S.; Mondello, M.; Romano, L. Chemical and pathological oxidative influences on band 3 protein anion-exchanger. *Cell. Physiol. Biochem.* **2005**, *16*, 77–86. [\[CrossRef\]](#)
11. Morabito, R.; Remigante, A.; Marino, A. Protective role of magnesium against oxidative stress on SO₄²⁻ uptake through Band 3 protein in human erythrocytes. *Cell. Physiol. Biochem.* **2019**, *52*, 1292–1308.
12. Konturek, S.J.; Konturek, P.C.; Brozowska, I.; Pawlik, M.; Sliwowski, Z.; Czenikiewicz-Guzik, M.; Kwiecień, S.; Brzozowski, T.; Bubenik, G.A.; Pawlik, W.W. Localization and biological activities of melatonin in intact and diseased gastrointestinal tract (GIT). *J. Physiol. Pharmacol.* **2007**, *58*, 381–405.
13. Hevia, D.; González-Menéndez, P.; Quiros-González, I.; Miar, A.; Rodríguez-García, A.; Tan, D.X.; Reiter, R.J.; Mayo, J.C.; Sainz, R.M. Melatonin uptake through glucose transporters: A new target for melatonin inhibition of cancer. *J. Pineal Res.* **2015**, *58*, 234–250. [\[CrossRef\]](#)
14. Tan, D.X.; Hardeland, R.; Manchester, L.C.; Paredes, S.D.; Korkmaz, A.; Sainz, R.M.; Mayo, J.C.; Fuentes-Broto, L.; Reiter, R.J. The changing biological roles of melatonin during evolution: From an antioxidant to signals of darkness, sexual selection and fitness. *Biol. Rev. Camb. Philos. Soc.* **2010**, *85*, 607–623. [\[CrossRef\]](#)
15. Zhang, H.M.; Zhang, Y. Melatonin A well-documented antioxidant with conditional pro-oxidant actions. *J. Pineal Res.* **2014**, *57*, 131–146. [\[CrossRef\]](#)
16. Abuja, P.M.; Liebmann, P.; Hayn, M.; Schauenstein, K.; Esterbauer, H. Antioxidant role of melatonin in lipid peroxidation of human LDL. *FEBS Lett.* **1997**, *413*, 289–293. [\[CrossRef\]](#)
17. Chakravarty, S.; Rizvi, S.I. Day and Night GSH and MDA levels in healthy adults and effects of different doses of melatonin on these parameters. *Int. J. Cell Biol.* **2011**, *2011*, 404591. [\[CrossRef\]](#)
18. Duell, P.B.; Wheaton, D.L.; Shultz, A.; Nguyen, H. Inhibition of LDL oxidation by melatonin requires supraphysiologic concentrations. *Clin. Chem.* **1998**, *44*, 1931–1936.
19. Da Silva, D.G.H.; Chaves, N.A.; Miyamoto, S.; de Almeida, E.A. Prolonged erythrocyte auto-incubation as an alternative model for oxidant generation system. *Toxicol. In Vitro* **2019**, *56*, 62–74. [\[CrossRef\]](#)
20. Buehler, P.W.; Alayash, A.I. Redox biology of blood revisited: The role of red blood cells in maintaining circulatory reductive capacity. *Antioxid. Redox Signal.* **2005**, *7*, 1755–1760. [\[CrossRef\]](#)
21. Dikmenoglu, N.; Ileri, E.; Seringec, N.; Ercil, D. Melatonin prevents lipid peroxidation in human erythrocytes but augments deterioration of deformability after in vitro oxidative stress. *Clin. Hemorheol. Microcirc.* **2008**, *40*, 235–242.

22. Morabito, R.; Remigante, A.; Di Pietro, M.L.; Giannetto, A.; La Spada, G.; Marino, A. $\text{SO}_4^{=}$ uptake and catalase role in preconditioning after H_2O_2 -induced oxidative stress in human erythrocytes. *Pflugers Arch.* **2017**, *469*, 235–250. [[CrossRef](#)]
23. Stefanovic, M.; Markham, N.O.; Parry, E.M.; Garrett-Beal, L.J.; Cline, A.P.; Gallagher, P.G.; Low, P.S.; Bodine, D.M. An 11-amino acid beta-hairpinloop in the cytoplasmic domain of band 3 is responsible for ankyrin binding in mouse erythrocytes. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 13972–13977. [[CrossRef](#)]
24. Dumaswala, U.J.; Zhuo, L.; Jacobsen, D.W.; Jain, S.K.; Sukalski, K.A. Protein and lipid oxidation of banked human erythrocytes: Role of glutathione. *Free Radic. Biol. Med.* **1999**, *27*, 1041–1049. [[CrossRef](#)]
25. Winterbourn, C.C. The biological chemistry of hydrogen peroxide. *Methods Enzymol.* **2013**, *528*, 3–25.
26. Shiga, T.; Maeda, N.; Kon, K. Erythrocyte rheology. *Crit. Rev. Oncol. Hematol.* **1990**, *10*, 9–48. [[CrossRef](#)]
27. Morabito, R.; Falliti, G.; Geraci, A.; La Spada, G.; Marino, A. Curcumin protects–SH groups and sulphate transport after oxidative damage in human erythrocytes. *Cell. Physiol. Biochem.* **2015**, *36*, 345–357. [[CrossRef](#)]
28. Romano, L.; Scuteri, A.; Gugliotta, T.; Romano, P.; De Luca, G.; Sidoti, A.; Amato, A. Sulphate influx in the erythrocytes of normotensive, diabetic and hypertensive patients. *Cell. Biol. Int.* **2002**, *26*, 421–426. [[CrossRef](#)]
29. Galano, A.; Reiter, R.J. Melatonin and its metabolites vs. oxidative stress: From individual actions to collective protection. *J. Pineal Res.* **2018**, *65*, e12514. [[CrossRef](#)]
30. Venegas, C.; Garcia, J.A.; Escames, G.; Ortiz, F.; Lopez, A.; Doerrier, C.; Garcia-Corzo, L.; Lopez, L.C.; Reiter, R.J.; Acuna-Castroviejo, D. Extrapineal melatonin: Analysis of its subcellular distribution and daily fluctuations. *J. Pineal Res.* **2012**, *52*, 217–227. [[CrossRef](#)]
31. Da Silva, D.G.H.; Ricci, O., Jr.; Alves de Almeida, E.; Bonini-Domingos, C.R. Potential utility of melatonin as an antioxidant therapy in the management of sickle cell anemia. *J. Pineal Res.* **2015**, *58*, 178–188. [[CrossRef](#)]
32. Ciftci, M.; Dilek, B.; Kufrevioglu, O.I. Effects of melatonin on enzyme activities of glucose 6-phosphate dehydrogenase from human erythrocytes in vitro and from rat erythrocytes in vivo. *Pharmacol. Res.* **2001**, *44*, 7–11. [[CrossRef](#)]
33. Tailleux, A.; Torpier, G.; Bonnefont-Rousselot, D.; Lestavel, S.; Lemdani, M.; Caudeville, B.; Furman, C.; Foricher, R.; Gardes-Albert, M.; Lesieur, D. Daily melatonin supplementation in mice increases atherosclerosis in proximal aorta. *Biochem. Biophys. Res. Commun.* **2002**, *293*, 1114–1123. [[CrossRef](#)]
34. Fulia, F.; Gitto, E.; Cuzzocrea, S.; Reiter, R.J.; Dugo, L.; Gitto, P.; Barberi, S.; Cordaro, S.; Barberi, I. Increased levels of malondialdehyde and nitrite/nitrate in the blood of asphyxiated newborns: Reduction by melatonin. *J. Pineal Res.* **2001**, *31*, 343–349. [[CrossRef](#)]
35. García, J.J.; Piñol-Ripoll, G.; Martínez-Ballarín, E.; Fuentes-Broto, L.; Miana-Mena, F.J.; Venegas, C.; Caballero, B.; Escames, G.; Coto-Montes, A.; Acuña-Castroviejo, D. Melatonin reduces membrane rigidity and oxidative damage in the brain of SAMP8 mice. *Neurobiol. Aging* **2011**, *32*, 2045–2054. [[CrossRef](#)]
36. Cimen, M.Y. Free radical metabolism in human erythrocytes. *Clin. Chim. Acta* **2008**, *390*, 1–11. [[CrossRef](#)]
37. García, J.J.; López-Pingarrón, L.; Almeida-Souza, P.; Tres, A.; Escudero, P.; García-Gil, F.A.; Tan, D.X.; Reiter, R.J.; Ramírez, J.M.; Bernal-Pérez, M. Protective effects of melatonin in reducing oxidative stress and in preserving the fluidity of biological membranes: A review. *J. Pineal Res.* **2014**, *56*, 225–237. [[CrossRef](#)]
38. Reiter, R.J.; Tan, D.; Kim, S.J.; Manchester, L.C.; Qi, W.; Garcia, J.J.; Cabrera, J.C.; El-Sokkary, G.; Rouvier-Garay, V. Augmentation of indices of oxidative damage in life-long melatonin-deficient rats. *Mech. Ageing Dev.* **1999**, *110*, 157–173. [[CrossRef](#)]
39. Tesoriere, L.; D’Arpa, D.; Conti, S.; Giaccone, V.; Pintaudi, A.M.; Livrea, M.A. Melatonin protects human red blood cells from oxidative hemolysis: New insights into the radical-scavenging activity. *J. Pineal Res.* **1999**, *27*, 95–105. [[CrossRef](#)]
40. Srour, M.A.; Bילו, Y.Y.; Jouma, M.; Irhimeh, M.R. Exposure of human erythrocytes to oxygen radicals causes loss of deformability, increased osmotic fragility, lipid peroxidation and protein degradation. *Clin. Hemorheol. Microcirc.* **2000**, *23*, 13–21.
41. Welbourn, E.M.; Wilson, M.T.; Yusof, A.; Metodiev, M.V.; Cooper, C.E. The mechanism of formation, structure and physiological relevance of covalent hemoglobin attachment to the erythrocyte membrane. *Free Radic. Biol. Med.* **2017**, 95–106. [[CrossRef](#)]
42. Miller, E.; Walczak, A.; Majsterek, I.; Kedziora, J. Melatonin reduces oxidative stress in the erythrocytes of multiple patients with secondary progressive clinical course. *J. Neuroimmunol.* **2013**, *257*, 97–101. [[CrossRef](#)]

43. Erat, M.; Ciftci, M. Effect of melatonin on enzyme activities of glutathione reductase from human erythrocytes in vitro and from rat erythrocytes in vivo. *Eur. J. Pharmacol.* **2006**, *537*, 59–63. [[CrossRef](#)]
44. Emamgholipour, S.; Hossein-Nezhad, A.; Ansari, M. Can Melatonin Act as an Antioxidant in Hydrogen Peroxide-Induced Oxidative Stress Model in Human Peripheral Blood Mononuclear Cells? *Biochem. Res. Int.* **2016**, *2016*, 5857940. [[CrossRef](#)]
45. Margoliash, E.; Novogrodsky, A.; Schejter, A. Irreversible reaction of 3-amino-1,2,4-triazole and related inhibitors with the protein of catalase. *Biochem. J.* **1960**, *74*, 339–348. [[CrossRef](#)]
46. Almroth, B.C.; Sturve, J.; Berglund, A.; Forlin, L. Oxidative damage in eelpout (*Zoarces viviparus*), measured as protein carbonyls and TBARS, as biomarkers. *Aquatic Toxicol.* **2005**, *73*, 171–180. [[CrossRef](#)]
47. Jessen, F.; Sjöholm, C.; Hoffmann, E.K. Identification of the anion exchange protein of Ehrlich cells: A kinetic analysis of the inhibitory effects of 4,4'-diisothiocyano-2,2'-stilbene-disulfonic acid(DIDS) and labeling of membrane proteins with 3H-DIDS. *J. Membr. Biol.* **1986**, *92*, 195–205. [[CrossRef](#)]
48. Pantaleo, A.; Ferru, E.; Pau, M.C.; Khadjavi, A.; Mandili, G.; Mattè, A.; Spano, A.; De Franceschi, L.; Pippia, P.; Turrini, F. Band 3 erythrocyte membrane protein acts as redox stress sensor leading to its phosphorylation by p (72) Syk. *Oxid. Med. Cell. Longev.* **2016**, 1–11. [[CrossRef](#)]
49. Bradford, M.M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* **1976**, *72*, 248–254. [[CrossRef](#)]
50. Laemmli, U.K. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **1970**, *227*, 680–685. [[CrossRef](#)]

Sample Availability: Not available.



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