## Lung resident mesenchymal cells isolated from patients with the Bronchiolitis Obliterans Syndrome display a deregulated epigenetic profile.

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Supplementary Figure S1.Validation of RT<sup>2</sup> array-based gene expression profile by qRT-PCR.

Values represent fold change in transcript copy number relative to control GAPDH. Mean (SD) expression of the DNMT1, DNMT3A, HAT1 and HDAC4, as measured by RT<sup>2</sup> arrays and TaqMan qRT-PCR in MSC from BALf of BOS 0p vs stable LTRs, **a**), and BOS vs stable LTRs, **b**).

Validation of array-based gene expression profile by qRT-PCR. Values represent fold change in transcript copy number relative to control U6. Mean ( $\pm$ SD) expression of the miR-106a, miR-124, miR-146a, miR-18b and miR-372, as measured by TLDA array and qRT-PCR in MSC from BALf of BOS 0p *vs* stable LTRs, **c**), and BOS *vs* stable LTRs, **d**).



Supplementary Figure S2. Epigenetic changes in human BOS lung biopsies.

a) QRT-PCR results of HDAC1 in lung tissues from BOS patients (n=4) and stable LTRs (n=4). Data are expressed as Mean ( $\pm$ SD). \* p $\leq$  0.05, two-tailed Student's test. b) MiRNA expression analysis in FFPE samples of BOS patients (n=4) and stable LTRs (n=4). Values represent fold change in transcript copy number relative to control U6, analysed by qRT-PCR. Data are expressed as Mean ( $\pm$ SD).

c) Mean (±SD) expression of some miRNAs, measured by qRT-PCR, in MSC from BALf of BOS (n=3) and in FFPE BOS samples (n=4).

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Supplementary Figure S3. Gene expression changes in MSC from BOS patients.

a) Non-supervised hierarchical clustering based on the gene expression levels of pro-fibrotic genes (TGF- $\beta$ 1, FN, Col1A1 and CTGF) and anti-fibrotic genes (PPAR $\gamma$  and CD90) in MSC from BALf of stable LTRs and BOS (n=5 and n=3, respectively). The dendrogram shows the relationships among gene expression patterns: "red" indicates high relative expression, "black" no change, and "green" low relative expression.



## Supplementary Figure S4. Annexin V/PI staining.

Diagrams of FITC-Annexin V/PI flowcytometry of treated and untreated MRC5. The lower left quadrants of eachpanel show the viable cells (negative for PI and FITC-Annexin V binding). The upper quadrants contain thenecrotic cells, positive for PI uptake. The lower right quadrants represent the apoptotic cells, FITC-Annexin V positive and PI negative. One representative experiment for each experimental condition is shown. The averaged percentageof viable, necrotic and apoptotic cells are represented in figure 4d.



Supplementary Figure S5. Cell cycle analysis with PI staining.

Cell cycle phase distributions of MRC5 treated with or without SAHA and/or TGF- $\beta$ 1, analyzed by ModFit LT software. The data are representative as a mean of 3 experiments with similar results, which are shown in the figure 4e.